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(54) Title: TOTIPOTENT HEMATOPOIETIC STEM CELL RECEPTORS AND THEIR LIGANDS

(57) Abstract

Isolated mammalian nucleic acid molecules encoding receptor protein tyrosine kinases expressed in primitive hematopoietic cells and not expressed in mature hematopoietic cells are provided. Also included are the receptors encoded by such nucleic acid molecules; the nucleic acid molecules encoding receptor protein tyrosine kinases having the sequences shown in Figure 1a (murine Flk2), Figure 1b (human Flk2) and Figure 2 (murine Flk1); the receptor protein tyrosine kinases having the amino acid sequences shown in Figure 1a, Figure 1b and Figure 2; ligands for the receptors; nucleic acids sequences that encode the ligands; and methods of stimulating the proliferation and/or differentiation of primitive mammalian hematopoietic stem cells comprising contacting the stem cells with a ligand that binds to a receptor protein tyrosine kinase expressed in primitive mammalian hematopoietic cells and not expressed in mature hematopoietic cells.

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**TOTIPOTENT HEMATOPOIETIC STEM CELL
RECEPTORS AND THEIR LIGANDS**

This application is a continuation-in-part of serial number 5 08/125,669, filed September 23, 1993, which is a continuation-in-part of serial number 08/096,759, filed July 22, 1993, which is a continuation-in-part of serial number 08/081,508, filed June 21, 1993, which is a continuation-in-part of serial number 08/080,244, filed June 18, 1993, which is a continuation-in-part 10 of serial number 08/076,022, filed June 9, 1993, which is a continuation-in-part of serial number 08/045,272, filed April 1, 1993, which is a continuation-in-part of serial number 08/005,941, filed January 15, 1993, which is a continuation-in-part of serial number 07/977,451, filed November 19, 1992, which 15 is a continuation-in-part of serial number 07/975,049 filed November 12, 1992, which is a continuation-in-part of serial number 07/906,397 filed June 26, 1992 which is a continuation-in-part of serial number 07/813,593 filed December 24, 1991, which is a continuation-in-part of serial number 07/793,065 filed 20 November 15, 1991, which is a continuation-in-part of serial number 07/728,913 filed June 28, 1991, which is a continuation-in-part of serial number 07/679,666 filed April 2, 1991, all of which are incorporated herein by reference.

25 The invention described in this application was made with U.S. government support from Grant Numbers R01-CA45339 and R01-DK42989 awarded by the National Institutes of Health. The government has certain rights in this invention.

30 FIELD OF THE INVENTION

The present invention relates to hematopoietic stem cell receptors, ligands for such receptors, and nucleic acid molecules encoding such receptors and ligands.

BACKGROUND OF THE INVENTION

5 The mammalian hematopoietic system comprises red and white blood cells. These cells are the mature cells that result from more primitive lineage-restricted cells. The cells of the hematopoietic system have been reviewed by Dexter and Spooncer in the Annual Review of Cell Biology 3, 423-441 (1987).

10 The red blood cells, or erythrocytes, result from primitive cells referred to by Dexter and Spooncer as erythroid burst-forming units (BFU-E). The immediate progeny of the erythroid burst-forming units are called erythroid colony-forming units (CFU-E).

15 The white blood cells contain the mature cells of the lymphoid and myeloid systems. The lymphoid cells include B lymphocytes and T lymphocytes. The B and T lymphocytes result from earlier progenitor cells referred to by Dexter and Spooncer as preT and preB cells.

20 The myeloid system comprises a number of cells including granulocytes, platelets, monocytes, macrophages, and megakaryocytes. The granulocytes are further divided into neutrophils, eosinophils, basophils and mast cells.

25 Each of the mature hematopoietic cells are specialized for specific functions. For example, erythrocytes are responsible for oxygen and carbon dioxide transport. T and B lymphocytes are responsible for cell-and antibody-mediated immune responses, respectively. Platelets are involved in blood clotting. Granulocytes and macrophages act generally as scavengers and accessory cells in the immune response against invading organisms and their by-products.

35 At the center of the hematopoietic system lie one or more

totipotent hematopoietic stem cells, which undergo a series of differentiation steps leading to increasingly lineage-restricted progenitor cells. The more mature progenitor cells are restricted to producing one or two lineages. Some examples of lineage-restricted progenitor cells mentioned by Dexter and Spooncer include granulocyte/macrophage colony-forming cells (GM-CFC), megakaryocyte colony-forming cells (Meg-CFC), eosinophil colony-forming cells (Eos-CFC), and basophil colony-forming cells (Bas-CFC). Other examples of progenitor cells are discussed above.

The hematopoietic system functions by means of a precisely controlled production of the various mature lineages. The totipotent stem cell possesses the ability both to self renew and to differentiate into committed progenitors for all hematopoietic lineages. These most primitive of hematopoietic cells are both necessary and sufficient for the complete and permanent hematopoietic reconstitution of a radiation-ablated hematopoietic system in mammals. The ability of stem cells to reconstitute the entire hematopoietic system is the basis of bone marrow transplant therapy.

It is known that growth factors play an important role in the development and operation of the mammalian hematopoietic system. The role of growth factors is complex, however, and not well understood at the present time. One reason for the uncertainty is that much of what is known about hematopoietic growth factors results from in vitro experiments. Such experiments do not necessarily reflect in vivo realities.

In addition, in vitro hematopoiesis can be established in the absence of added growth factors, provided that marrow stromal cells are added to the medium. The relationship between stromal cells and hematopoietic growth factors in vivo is not understood. Nevertheless, hematopoietic growth factors have been shown to be

highly active in vivo.

From what is known about them, hematopoietic growth factors appear to exhibit a spectrum of activities. At one end of the spectrum are growth factors such as erythropoietin, which is believed to promote proliferation only of mature erythroid progenitor cells. In the middle of the spectrum are growth factors such as IL-3, which is believed to facilitate the growth and development of early stem cells as well as of numerous progenitor cells. Some examples of progenitor cells induced by IL-3 include those restricted to the granulocyte/macrophage, eosinophil, megakaryocyte, erythroid and mast cell lineages.

At the other end of the spectrum is the hematopoietic growth factor that, along with the corresponding receptor, was discussed in a series of articles in the October 5, 1990 edition of Cell. The receptor is the product of the W locus, c-kit, which is a member of the class of receptor protein tyrosine kinases. The ligand for c-kit, which is referred to by various names such as stem cell factor (SCF) and mast cell growth factor (MGF), is believed to be essential for the development of early hematopoietic stem cells and cells restricted to the erythroid and mast cell lineages in mice; see, for example, Copeland et al., Cell 63, 175-183 (1990).

It appears, therefore, that there are growth factors that exclusively affect mature cells. There also appear to be growth factors that affect both mature cells and stem cells. The growth factors that affect both types of cells may affect a small number or a large number of mature cells.

There further appears to be an inverse relationship between the ability of a growth factor to affect mature cells and the ability of the growth factor to affect stem cells. For example, the c-kit ligand, which stimulates a small number of mature

cells, is believed to be more important in the renewal and development of stem cells than is IL-3, which is reported to stimulate proliferation of many mature cells (see above).

5 Prior to the present specification, there have been no reports of growth factors that exclusively stimulate stem cells in the absence of an effect on mature cells. The discovery of such growth factors would be of particular significance.

10 As mentioned above, c-kit is a protein tyrosine kinase (pTK). It is becoming increasingly apparent that the protein tyrosine kinases play an important role as cellular receptors for hematopoietic growth factors. Other receptor pTKs include the receptors of colony stimulating factor 1 (CSF-1) and PDGF.

15 The pTK family can be recognized by the presence of several conserved amino acid regions in the catalytic domain. These conserved regions are summarized by Hanks et al. in *Science* 241, 42-52 (1988), see Figure 1 starting on page 46 and by Wilks in *Proc. Natl. Acad. Sci. USA* 86, 1603-1607 (1989), see Figure 2 on page 1605.

20 Additional protein tyrosine kinases that represent hematopoietic growth factor receptors are needed in order more effectively to stimulate the self-renewal of the totipotent hematopoietic stem cell and to stimulate the development of all cells of the hematopoietic system both in vitro and in vivo. Novel hematopoietic growth factor receptors that are present only on primitive stem cells, but are not present on mature progenitor cells, are particularly desired. Ligands for the novel receptors are also desirable to act as hematopoietic growth factors. Nucleic acid sequences encoding the receptors and ligands are needed to produce recombinant receptors and ligands.

SUMMARY OF THE INVENTION

These and other objectives as will be apparent to those with ordinary skill in the art have been met by providing isolated mammalian nucleic acid molecules encoding receptor protein tyrosine kinases expressed in primitive hematopoietic cells and not expressed in mature hematopoietic cells. Also included are the receptors encoded by such nucleic acid molecules; the nucleic acid molecules encoding receptor protein tyrosine kinases having the sequences shown in Figure 1a.1-1a.6 (hereinafter Figure 1a)(murine Flk2), Figure 1b.1-1b.6 (hereinafter Figure 1b)(human Flk2) and Figure 2.1-2.9 (hereinafter Figure 2)(murine Flk1)(See SEQ. ID. NOS. 1, 3 and 5, respectively); the receptor protein tyrosine kinases having the amino acid sequences shown in Figure 1a, Figure 1b and Figure 2 (See SEQ. ID. NOS. 2, 4 and 6, respectively); ligands for the receptors; nucleic acid sequences that encode the ligands; and methods of stimulating the proliferation of primitive mammalian hematopoietic stem cells comprising contacting the stem cells with a ligand that binds to a receptor protein tyrosine kinase expressed in primitive mammalian hematopoietic cells and not expressed in mature hematopoietic cells.

DESCRIPTION OF THE FIGURES

Figure 1a.1 through 1a.6 shows the cDNA and amino acid sequences of murine Flk2. All subsequent references to Figure 1a are intended to refer to Figure 1a.1 through 1a.6. The amino acid residues occur directly below the nucleotides in the open reading frame. Amino acids -27 to -1 constitute the hydrophobic leader sequence. Amino acids 1 to 517 constitute the extracellular receptor domain. Amino acids 518 to 537 constitute the transmembrane region. Amino acids 538 to 966 constitute the intracellular catalytic domain. Counting amino acid residue -27 as residue number 1, the following amino acid residues in the

intracellular domain are catalytic sub-domains identified by Hanks (see above): 618-623, 811-819, 832-834, 857-862, 872-878. The sequence at residues 709-785 is a signature sequence characteristic of Flk2. The protein tyrosine kinases generally have a signature sequence in this region. (See SEQ. ID. NOS. 1-2)

Figure 1b.1 through 1b.6 shows the complete cDNA and amino acid sequences of human Flk2 receptor. All subsequent references to Figure 1b are intended to refer to Figure 1b.1 through 1b.6. Amino acids -27 to -1 constitute the hydrophobic leader sequence. Amino acids 1 to 516 constitute the extracellular receptor domain. Amino acids 517 to 536 constitute the transmembrane region. Amino acids 537 to 966 constitute the intracellular catalytic domain. (See SEQ. ID. NOS. 3-4)

15

Figure 2.1 through 2.9 shows the cDNA and amino acid sequences of murine Flk1. All subsequent references to Figure 2 are intended to refer to Figure 2.1 through 2.9. Amino acids -19 to -1 constitute the hydrophobic leader sequence. Amino acids 1 to 743 constitute the extracellular receptor domain. Amino acids 744 to 765 constitute the transmembrane region. Amino acids 766 to 1348 constitute the intracellular catalytic domain. (See SEQ. ID. NOS. 5-6)

25

Figure 3 shows the time response of binding between a murine stromal cell line (2018) and APtag-Flk2 as well as APtag-Flk1. APtag without receptor (SEAP) is used as a control. See Example 8.

30

Figure 4 shows the dose response of binding between stromal cells (2018) and APtag-Flk2 as well as APtag-Flk1. APtag without receptor (SEAP) is used as a control. See Example 8.

35

DETAILED DESCRIPTION OF THE INVENTIONReceptors

5 In one embodiment, the invention relates to an isolated mammalian nucleic acid molecule encoding a receptor protein tyrosine kinase expressed in primitive mammalian hematopoietic cells and not expressed in mature hematopoietic cells.

10 The nucleic acid molecule may be a DNA, cDNA, or RNA molecule. The mammal in which the nucleic acid molecule exists may be any mammal, such as a mouse, rat, rabbit, or human.

15 The nucleic acid molecule encodes a protein tyrosine kinase (pTK). Members of the pTK family can be recognized by the conserved amino acid regions in the catalytic domains. Examples of pTK consensus sequences have been provided by Hanks et al. in Science 241, 42-52 (1988); see especially Figure 1 starting on page 46 and by Wilks in Proc. Natl. Acad. Sci. USA 86, 1603-1607 (1989); see especially Figure 2 on page 1605. A methionine residue at position 205 in the conserved sequence WMAPES is characteristic of pTK's that are receptors.

25 The Hanks et al article identifies eleven catalytic sub-domains containing pTK consensus residues and sequences. The pTKs of the present invention will have most or all of these consensus residues and sequences.

30 Some particularly strongly conserved residues and sequences are shown in Table 1.

TABLE 1
Conserved Residues and Sequences in pTKs¹

35	<u>Position</u> ²	<u>Residue or Sequence</u>	<u>Catalytic Domain</u>
----	------------------------------	----------------------------	-------------------------

50	G	I
52	G	I
57	V	I
70	A	II
72	K	II
91	E	III
166	D	VI
171	N	VI
184-186	DFG	VII
208	E	VIII
220	D	IX
225	G	IX
280	R	XI

15 1. See Hanks et al., *Science* 241, 42-52 (1988)
 2. Adjusted in accordance with Hanks et al., *Id.*

20 A pTK of the invention may contain all thirteen of these highly conserved residues and sequences. As a result of natural or synthetic mutations, the pTKs of the invention may contain fewer than all thirteen strongly conserved residues and sequences, such as 11, 9, or 7 such sequences.

25 The receptors of the invention generally belong to the same class of pTK sequences that c-kit belongs to. It has surprisingly been discovered, however, that a new functional class of receptor pTKs exists. The new functional class of receptor pTKs is expressed in primitive hematopoietic cells, but not expressed in mature 30 hematopoietic cells.

35 For the purpose of this specification, a primitive hematopoietic cell is totipotent, i.e. capable of reconstituting all hematopoietic blood cells in vivo. A mature hematopoietic cell is non-self-renewing, and has limited proliferative capacity - i.e., a limited ability to give rise to multiple lineages. Mature hematopoietic cells, for the purposes of this specification, are generally capable of giving rise to only one or two lineages in vitro or in vivo.

5 It should be understood that the hematopoietic system is complex, and contains many intermediate cells between the primitive totipotent hematopoietic stem cell and the totally committed mature hematopoietic cells defined above. As the stem cell develops into increasingly mature, lineage-restricted cells, it gradually loses its capacity for self-renewal.

10 The receptors of the present invention may and may not be expressed in these intermediate cells. The necessary and sufficient condition that defines members of the new class of receptors is that they are present in the primitive, totipotent stem cell or cells, and not in mature cells restricted only to one or, at most, two lineages.

15 An example of a member of the new class of receptor pTKs is called fetal liver kinase 2 (Flk2) after the organ in which it was found. There is approximately 1 totipotent stem cell per 10^4 cells in mid-gestation (day 14) fetal liver in mice. In addition to fetal liver, Flk2 is also expressed in fetal spleen, fetal thymus, adult brain, and adult marrow.

20 For example, Flk2 is expressed in individual multipotential CFU-Blast colonies capable of generating numerous multilineage colonies upon replating. It is likely, therefore, that Flk2 is expressed in the entire primitive (i.e. self-renewing) portion of the hematopoietic hierarchy. This discovery is consistent with Flk2 being important in transducing putative self-renewal signals from the environment.

25 30 It is particularly relevant that the expression of Flk2 mRNA occurs in the most primitive thymocyte subset. Even in two closely linked immature subsets that differ in expression of the IL-2 receptor, Flk2 expression segregates to the more primitive subset lacking an IL-2 receptor. The earliest thymocyte subset is believed to be uncommitted. Therefore, the thymocytes

expressing Flk2 may be multipotential. Flk2 is the first receptor tyrosine kinase known to be expressed in the T-lymphoid lineage.

5 The fetal liver mRNA migrates relative to 28S and 18S ribosomal bands on formaldehyde agarose gels at approximately 3.5 kb, while the brain message is considerably larger. In adult tissues, Flk2 m-RNA from both brain and bone marrow migrated at approximately 3.5 kb.

10

10 A second pTK receptor is also included in the present invention. This second receptor, which is called fetal liver kinase 1 (Flk1), is not a member of the same class of receptors as Flk2, since Flk1 may be found in some more mature hematopoietic cells. The amino acid sequence of murine Flk1 is given in Figure 2. (See SEQ. ID. NOS. 5-6)

15 The present invention includes the Flk1 receptor as well as DNA, cDNA and RNA encoding Flk1. The DNA sequence of murine Flk1 is also given in Figure 2. (See SEQ. ID. NO. 5) Flk1 may be found in the same organs as Flk2, as well as in fetal brain, stomach, kidney, lung, heart and intestine; and in adult kidney, heart, spleen, lung, muscle, and lymph nodes.

20 The receptor protein tyrosine kinases of the invention are known to be divided into easily found domains. The DNA sequence corresponding to the pTKs encode, starting at their 5'-ends, a hydrophobic leader sequence followed by a hydrophilic extracellular domain, which binds to, and is activated by, a specific ligand. Immediately downstream from the extracellular receptor domain, is a hydrophobic transmembrane region. The transmembrane region is immediately followed by a basic catalytic domain, which may easily be identified by reference to the Hanks et al. and Wilks articles discussed above.

30

35

The following table shows the nucleic acid and amino acid numbers that correspond to the signal peptide, the extracellular domain, the transmembrane region and the intracellular domain for murine Flk1 (mFlk1), murine Flk2 (mFlk2) and human Flk2 (hFlk2).

5

mFlk1

	<u>Signal Peptide</u>	<u>Extracellular</u>	<u>Transmembrane</u>	<u>Intracellular</u>
	aa # -19 to -1	1 to 743	744 to 765	766 to 1348
	aa code M A	A E	V V	R A
10	na # 208-264	265-2493	2494-2559	2560-4308

mFlk2

	<u>Signal Peptide</u>	<u>Extracellular</u>	<u>Transmembrane</u>	<u>Intracellular</u>
	aa # -27 to -1	1 to 517	518 to 537	538 to 966
15	aa code M T	N S	F C	H S
	na # 31-111	112-1662	1663-1722	1723-3006

hFlk2

	<u>Signal Peptide</u>	<u>Extracellular</u>	<u>Transmembrane</u>	<u>Intracellular</u>
20	aa # -27 to -1	1 to 516	517 to 536	537 to 966
	aa code M N	Q F	Y C	H S
	na # 58-138	139-1689	1690-1746	1747-3036

The present invention includes the extracellular receptor domain lacking the transmembrane region and catalytic domain. Preferably, the hydrophobic leader sequence is also removed from the extracellular domain. In the case of human and murine Flk2, the hydrophobic leader sequence includes amino acids -27 to -1. (See SEQ. ID. NOS. 2 and 4)

30

These regions and domains may easily be visually identified by those having ordinary skill in the art by reviewing the amino acid sequence in a suspected pTK and comparing it to known pTKs. For example, referring to Figure 1a, the transmembrane region of Flk2, which separates the extracellular receptor domain from the

catalytic domain, is encoded by nucleotides 1663 (T) to 1722 (C). These nucleotides correspond to amino acid residues 545 (Phe) to 564 (Cys). (See SEQ. ID. NOS. 1-2) The amino acid sequence between the transmembrane region and the catalytic sub-domain (amino acids 618-623) identified by Hanks et al. as sub-domain I (i.e., GXGXXG) is characteristic of receptor protein tyrosine kinases.

The extracellular domain may also be identified through commonly recognized criteria of extracellular amino acid sequences. The determination of appropriate criteria is known to those skilled in the art, and has been described, for example, by Hopp et al, Proc. Nat'l Acad. Sci. USA 78, 3824-3828 (1981); Kyte et al, J. Mol. Biol. 157, 105-132 (1982); Emini, J. Virol. 55, 836-839 (1985); Jameson et al, CA BIOS 4, 181-186 (1988); and Karplus et al, Naturwissenschaften 72, 212-213 (1985). Amino acid domains predicted by these criteria to be surface exposed characteristic of extracellular domains.

As will be discussed in more detail below, the nucleic acid molecules that encode the receptors of the invention may be inserted into known vectors for use in standard recombinant DNA techniques. Standard recombinant DNA techniques are those such as are described in Sambrook et al., "Molecular Cloning," Second Edition, Cold Spring Harbor Laboratory Press (1987) and by Ausubel et al., Eds, "Current Protocols in Molecular Biology," Green Publishing Associates and Wiley-Interscience, New York (1987). The vectors may be circular (i.e. plasmids) or non-circular. Standard vectors are available for cloning and expression in a host. The host may be prokaryotic or eucaryotic. Prokaryotic hosts are preferably E. coli. Preferred eucaryotic hosts include yeast, insect and mammalian cells. Preferred mammalian cells include, for example, CHO, COS and human cells.

Ligands

The invention also includes ligands that bind to the receptor pTKs of the invention. In addition to binding, the ligands stimulate the proliferation of additional primitive stem cells, differentiation into more mature progenitor cells, or both.

The ligand may be a growth factor that occurs naturally in a mammal, preferably the same mammal that produces the corresponding receptor. The growth factor may be isolated and purified, or be present on the surface of an isolated population of cells, such as stromal cells. A partial amino acid sequence of a Flk2 ligand is AQSLSF_XFTKFDLD, wherein X is any amino acid. (See SEQ. ID. NO. 11)

The ligand may also be a molecule that does not occur naturally in a mammal. For example, antibodies, preferably monoclonal, raised against the receptors of the invention or against anti-ligand antibodies mimic the shape of, and act as, ligands if they constitute the negative image of the receptor or anti-ligand antibody binding site. The ligand may also be a non-protein molecule that acts as a ligand when it binds to, or otherwise comes into contact with, the receptor.

In another embodiment, nucleic acid molecules encoding the ligands of the invention are provided. The nucleic acid molecule may be RNA, DNA or cDNA.

Stimulating Proliferation of Stem Cells

The invention also includes a method of stimulating the proliferation and/or differentiation of primitive mammalian hematopoietic stem cells as defined above. The method comprises contacting the stem cells with a ligand in accordance with the

present invention. The stimulation of proliferation and/or differentiation may occur in vitro or in vivo.

5 The ability of a ligand according to the invention to stimulate proliferation of stem cells in vitro and in vivo has important therapeutic applications. Such applications include treating mammals, including humans, whose primitive stem cells do not sufficiently undergo self-renewal. Example of such medical problems include those that occur when defects in hematopoietic 10 stem cells or their related growth factors depress the number of white blood cells. Examples of such medical problems include anemia, such as macrocytic and aplastic anemia. Bone marrow damage resulting from cancer chemotherapy and radiation is another example of a medical problem that would be helped by the 15 stem cell factors of the invention.

Functional Equivalents

20 The invention includes functional equivalents of the pTK receptors, receptor domains, and ligands described above as well as of the nucleic acid sequences encoding them. A protein is considered a functional equivalent of another protein for a specific function if the equivalent protein is immunologically cross-reactive with, and has the same function as, the receptors 25 and ligands of the invention. The equivalent may, for example, be a fragment of the protein, or a substitution, addition or deletion mutant of the protein.

30 For example, it is possible to substitute amino acids in a sequence with equivalent amino acids. Groups of amino acids known normally to be equivalent are:

- (a) Ala(A) Ser(S) Thr(T) Pro(P) Gly(G);
- (b) Asn(N) Asp(D) Glu(E) Gln(Q);
- 35 (c) His(H) Arg(R) Lys(K);

- (d) Met(M) Leu(L) Ile(I) Val(V); and
- (e) Phe(F) Tyr(Y) Trp(W).

5 Substitutions, additions and/or deletions in the receptors and ligands may be made as long as the resulting equivalent receptors and ligands are immunologically cross reactive with, and have the same function as, the native receptors and ligands.

10 The equivalent receptors and ligands will normally have substantially the same amino acid sequence as the native receptors and ligands. An amino acid sequence that is substantially the same as another sequence, but that differs from the other sequence by means of one or more substitutions, additions and/or deletions is considered to be an equivalent sequence. Preferably, less than 25%, more preferably less than 15 10%, and most preferably less than 5% of the number of amino acid residues in the amino acid sequence of the native receptors and ligands are substituted for, added to, or deleted from.

20 Equivalent nucleic acid molecules include nucleic acid sequences that encode equivalent receptors and ligands as defined above. Equivalent nucleic acid molecules also include nucleic acid sequences that differ from native nucleic acid sequences in ways that do not affect the corresponding amino acid sequences.

25

ISOLATION OF NUCLEIC ACID MOLECULES AND PROTEINS

Isolation of Nucleic Acid Molecules Encoding Receptors

30 In order to produce nucleic acid molecules encoding mammalian stem cell receptors, a source of stem cells is provided. Suitable sources include fetal liver, spleen, or thymus cells or adult marrow or brain cells.

35 For example, suitable mouse fetal liver cells may be

obtained at day 14 of gestation. Mouse fetal thymus cells may be obtained at day 14-18, preferably day 15, of gestation. Suitable fetal cells of other mammals are obtained at gestation times corresponding to those of mouse.

5

Total RNA is prepared by standard procedures from stem cell receptor-containing tissue. The total RNA is used to direct cDNA synthesis. Standard methods for isolating RNA and synthesizing cDNA are provided in standard manuals of molecular biology such as, for example, in Sambrook et al., "Molecular Cloning," Second Edition, Cold Spring Harbor Laboratory Press (1987) and in Ausubel et al., (Eds), "Current Protocols in Molecular Biology," Greene Associates/Wiley Interscience, New York (1990).

15

The cDNA of the receptors is amplified by known methods. For example, the cDNA may be used as a template for amplification by polymerase chain reaction (PCR); see Saiki et al., *Science*, 239, 487 (1988) or Mullis et al., U.S. patent 4,683,195. The sequences of the oligonucleotide primers for the PCR amplification are derived from the sequences of known receptors, such as from the sequences given in Figures 1a and 1b for Flk2 and in Figure 2 for Flk1, preferably from Flk2. (See SEQ. ID. NOS. 1, 3 and 5, respectively) The oligonucleotides are synthesized by methods known in the art. Suitable methods include those described by Caruthers in *Science* 230, 281-285 (1985).

30

In order to isolate the entire protein-coding regions for the receptors of the invention, the upstream oligonucleotide is complementary to the sequence at the 5' end, preferably encompassing the ATG start codon and at least 5-10 nucleotides upstream of the start codon. The downstream oligonucleotide is complementary to the sequence at the 3' end, optionally encompassing the stop codon. A mixture of upstream and downstream oligonucleotides are used in the PCR amplification.

35

The conditions are optimized for each particular primer pair according to standard procedures. The PCR product is analyzed by electrophoresis for the correct size cDNA corresponding to the sequence between the primers.

5

Alternatively, the coding region may be amplified in two or more overlapping fragments. The overlapping fragments are designed to include a restriction site permitting the assembly of the intact cDNA from the fragments.

10

The amplified DNA encoding the receptors of the invention may be replicated in a wide variety of cloning vectors in a wide variety of host cells. The host cell may be prokaryotic or eukaryotic. The DNA may be obtained from natural sources and, 15 optionally, modified, or may be synthesized in whole or in part.

The vector into which the DNA is spliced may comprise segments of chromosomal, non-chromosomal and synthetic DNA sequences. Some suitable prokaryotic cloning vectors include 20 plasmids from E. coli, such as colE1, pCR1, pBR322, pMB9, pUC, pKSM, and RP4. Prokaryotic vectors also include derivatives of phage DNA such as M13 and other filamentous single-stranded DNA 25 phages.

25

Isolation of Receptors

DNA encoding the receptors of the invention are inserted into a suitable vector and expressed in a suitable prokaryotic or eucaryotic host. Vectors for expressing proteins in bacteria, 30 especially E.coli, are known. Such vectors include the PATH vectors described by Dieckmann and Tzagoloff in J. Biol. Chem. 260, 1513-1520 (1985). These vectors contain DNA sequences that encode anthranilate synthetase (TrpE) followed by a polylinker at the carboxy terminus. Other expression vector systems are based 35 on beta-galactosidase (pEX); lambda P_L; maltose binding protein

(pMAL); and glutathione S-transferase (pGST) - see Gene 67, 31 (1988) and Peptide Research 3, 167 (1990).

5 Vectors useful in yeast are available. A suitable example is the 2μ plasmid.

10 Suitable vectors for use in mammalian cells are also known. Such vectors include well-known derivatives of SV-40, adenovirus, retrovirus-derived DNA sequences and shuttle vectors derived from combination of functional mammalian vectors, such as those described above, and functional plasmids and phage DNA.

15 Further eukaryotic expression vectors are known in the art (e.g., P.J. Southern and P. Berg, J. Mol. Appl. Genet. 1, 327-341 (1982); S. Subramani et al, Mol. Cell. Biol. 1, 854-864 (1981); R.J. Kaufmann and P.A. Sharp, "Amplification And Expression Of Sequences Cotransfected with A Modular Dihydrofolate Reductase Complementary DNA Gene," J. Mol. Biol. 159, 601-621 (1982); R.J. Kaufmann and P.A. Sharp, Mol. Cell. Biol. 159, 601-664 (1982); 20 S.I. Scahill et al, "Expression And Characterization Of The Product Of A Human Immune Interferon DNA Gene In Chinese Hamster Ovary Cells," Proc. Natl. Acad. Sci. USA 80, 4654-4659 (1983); G. Urlaub and L.A. Chasin, Proc. Natl. Acad. Sci. USA 77, 4216-4220, (1980).

25

30 The expression vectors useful in the present invention contain at least one expression control sequence that is operatively linked to the DNA sequence or fragment to be expressed. The control sequence is inserted in the vector in order to control and to regulate the expression of the cloned DNA sequence. Examples of useful expression control sequences are the lac system, the trp system, the tac system, the trc system, major operator and promoter regions of phage lambda, the control region of fd coat protein, the glycolytic promoters of yeast, 35 e.g., the promoter for 3-phosphoglycerate kinase, the promoters

of yeast acid phosphatase, e.g., Pho5, the promoters of the yeast alpha-mating factors, and promoters derived from polyoma, adenovirus, retrovirus, and simian virus, e.g., the early and late promoters of SV40, and other sequences known to control the expression of genes of prokaryotic or eukaryotic cells and their viruses or combinations thereof.

Vectors containing the receptor-encoding DNA and control signals are inserted into a host cell for expression of the receptor. Some useful expression host cells include well-known prokaryotic and eukaryotic cells. Some suitable prokaryotic hosts include, for example, E. coli, such as E. coli SG-936, E. coli HB 101, E. coli W3110, E. coli X1776, E. coli X2282, E. coli DHI, and E. coli MRCl, Pseudomonas, Bacillus, such as Bacillus subtilis, and Streptomyces. Suitable eukaryotic cells include yeast and other fungi, insect, animal cells, such as COS cells and CHO cells, human cells and plant cells in tissue culture.

The human homologs of the mouse receptors described above are isolated by a similar strategy. RNA encoding the receptors are obtained from a source of human cells enriched for primitive stem cells. Suitable human cells include fetal spleen, thymus and liver cells, and umbilical cord blood as well as adult brain and bone marrow cells. The human fetal cells are preferably obtained on the day of gestation corresponding to mid-gestation in mice. The amino acid sequences of the human flk receptors as well as of the nucleic acid sequences encoding them are homologous to the amino acid and nucleotide sequences of the mouse receptors.

30

In the present specification, the sequence of a first protein, such as a receptor or a ligand, or of a nucleic acid molecule that encodes the protein, is considered homologous to a second protein or nucleic acid molecule if the amino acid or nucleotide sequence of the first protein or nucleic acid molecule

is at least about 30% homologous, preferably at least about 50% homologous, and more preferably at least about 65% homologous to the respective sequences of the second protein or nucleic acid molecule. In the case of proteins having high homology, the 5 amino acid or nucleotide sequence of the first protein or nucleic acid molecule is at least about 75% homologous, preferably at least about 85% homologous, and more preferably at least about 95% homologous to the amino acid or nucleotide sequence of the second protein or nucleic acid molecule.

10

Combinations of mouse oligonucleotide pairs are used as PCR primers to amplify the human homologs from the cells to account for sequence divergence. The remainder of the procedure for obtaining the human flk homologs are similar to those described 15 above for obtaining mouse flk receptors. The less than perfect homology between the human flk homologs and the mouse oligonucleotides is taken into account in determining the stringency of the hybridization conditions.

20

Assay for expression of Receptors on Stem Cells

25

In order to demonstrate the expression of flk receptors on the surface of primitive hematopoietic stem cells, antibodies that recognize the receptor are raised. The receptor may be the entire protein as it exists in nature, or an antigenic fragment of the whole protein. Preferably, the fragment comprises the predicted extra-cellular portion of the molecule.

30

Antigenic fragments may be identified by methods known in the art. Fragments containing antigenic sequences may be selected on the basis of generally accepted criteria of potential antigenicity and/or exposure. Such criteria include the hydrophilicity and relative antigenic index, as determined by surface exposure analysis of proteins. The determination of 35 appropriate criteria is known to those skilled in the art, and

has been described, for example, by Hopp et al, Proc. Nat'l Acad. Sci. USA 78, 3824-3828 (1981); Kyte et al, J. Mol. Biol. 157, 105-132 (1982); Emini, J. Virol. 55, 836-839 (1985); Jameson et al, CA BIOS 4, 181-186 (1988); and Karplus et al,

5 Naturwissenschaften 72, 212-213 (1985). Amino acid domains predicted by these criteria to be surface exposed are selected preferentially over domains predicted to be more hydrophobic or hidden.

10 The proteins and fragments of the receptors to be used as antigens may be prepared by methods known in the art. Such methods include isolating or synthesizing DNA encoding the proteins and fragments, and using the DNA to produce recombinant proteins, as described above.

15 Fragments of proteins and DNA encoding the fragments may be chemically synthesized by methods known in the art from individual amino acids and nucleotides. Suitable methods for synthesizing protein fragments are described by Stuart and Young 20 in "Solid Phase Peptide Synthesis," Second Edition, Pierce Chemical Company (1984). Suitable methods for synthesizing DNA fragments are described by Caruthers in Science 230, 281-285 (1985).

25 If the receptor fragment defines the epitope, but is too short to be antigenic, it may be conjugated to a carrier molecule in order to produce antibodies. Some suitable carrier molecules include keyhole limpet hemocyanin, Ig sequences, TrpE, and human 30 or bovine serum albumen. Conjugation may be carried out by methods known in the art. One such method is to combine a cysteine residue of the fragment with a cysteine residue on the carrier molecule.

35 The antibodies are preferably monoclonal. Monoclonal antibodies may be produced by methods known in the art. These

methods include the immunological method described by Kohler and Milstein in *Nature* 256, 495-497 (1975) and Campbell in "Monoclonal Antibody Technology, The Production and Characterization of Rodent and Human Hybridomas" in Burdon et al., Eds, *Laboratory Techniques in Biochemistry and Molecular Biology*, Volume 13, Elsevier Science Publishers, Amsterdam (1985); as well as by the recombinant DNA method described by Huse et al in *Science* 246, 1275-1281 (1989).

10 Polyclonal or monoclonal antisera shown to be reactive with receptor-encoded native proteins, such as with Flk1 and Flk2 encoded proteins, expressed on the surface of viable cells are used to isolate antibody-positive cells. One method for isolating such cells is flow cytometry; see, for example, Loken et al., European patent application 317,156. The cells obtained are assayed for stem cells by engraftment into radiation-ablated hosts by methods known in the art; see, for example, Jordan et al., *Cell* 61, 953-963 (1990).

20 Criteria for Novel Stem Cell Receptor Tyrosine Kinases
Expressed in Stem Cells

Additional novel receptor tyrosine kinase cDNAs are obtained by amplifying cDNAs from stem cell populations using 25 oligonucleotides as PCR primers; see above. Examples of suitable oligonucleotides are PTK1 and PTK2, which were described by Wilks et al. in *Proc. Natl. Acad. Sci. USA* 86, 1603-1607 (1989). Novel cDNA is selected on the basis of differential hybridization screening with probes representing known kinases. The cDNA 30 clones hybridizing only at low stringency are selected and sequenced. The presence of the amino acid triplet DFG confirms that the sequence represents a kinase. The diagnostic methionine residue in the WMAPES motif is indicative of a receptor-like kinase, as described above. Potentially novel sequences obtained 35 are compared to available sequences using databases such as

Genbank in order to confirm uniqueness. Gene-specific oligonucleotides are prepared as described above based on the sequence obtained. The oligonucleotides are used to analyze stem cell enriched and depleted populations for expression. Such cell populations in mice are described, for example, by Jordan et al. in Cell 61, 953-956 (1990); Ikuta et al. in Cell 62, 863-864 (1990); Spangrude et al. in Science 241, 58-62 (1988); and Szilvassy et al. in Blood 74, 930-939 (1989). Examples of such human cell populations are described as CD33⁺CD34⁺ by Andrews et al. in the Journal of Experimental Medicine 169, 1721-1731 (1989). Other human stem cell populations are described, for example, in Civin et al., European Patent Application 395,355 and in Loken et al., European Patent Application 317,156.

15

Isolating Ligands and Nucleic Acid Molecules Encoding Ligands

Cells that may be used for obtaining ligands include stromal cells, for example stromal cells from fetal liver, fetal spleen, fetal thymus and fetal or adult bone marrow. Cell lines expressing ligands are established and screened.

For example, cells such as stromal (non-hematopoietic) cells from fetal liver are immortalized by known methods. Examples of known methods of immortalizing cells include transduction with a temperature sensitive SV40 T-antigen expressed in a retroviral vector. Infection of fetal liver cells with this virus permits the rapid and efficient establishment of multiple independent cell lines. These lines are screened for ligand activity by methods known in the art, such as those outlined below.

Ligands for the receptors of the invention, such as Flk1 and Flk2, may be obtained from the cells in several ways. For example, a bioassay system for ligand activity employs chimeric tagged receptors; see, for example, Flanagan et al., Cell 63,

185-194 (1990). One strategy measures ligand binding directly via a histochemical assay. Fusion proteins comprising the extracellular receptor domains and secretable alkaline phosphatase (SEAP) are constructed and transfected into suitable 5 cells such as NIH/3T3 or COS cells. Flanagan et al. refer to such DNA or amino acid constructs as APtag followed by the name of the receptor - i.e. APtag-c-kit. The fusion proteins bind with high affinity to cells expressing surface-bound ligand. 10 Binding is detectable by the enzymatic activity of the alkaline phosphatase secreted into the medium. The bound cells, which are often stromal cells, are isolated from the APtag-receptor complex.

15 For example, some stromal cells that bind APtag-Flk1 and APtag-Flk2 fusion proteins include mouse fetal liver cells (see example 1); human fetal spleen cells (see example 3); and human fetal liver (example 3). Some stromal fetal thymus cells contain Flk1 ligand (example 3).

20 To clone the cDNA that encodes the ligand, a cDNA library is constructed from the isolated stromal cells in a suitable expression vector, preferably a phage such as CDM8, pSV Sport (BRL Gibco) or piH3, (Seed et al., Proc. Natl. Acad. Sci. USA 84, 3365-3369 (1987)). The library is transfected into suitable host 25 cells, such as COS cells. Cells containing ligands on their surface are detected by known methods, see above.

30 In one such method, transfected COS cells are distributed into single cell suspensions and incubated with the secreted alkaline phosphatase-flk receptor fusion protein, which is present in the medium from NIH/3T3 or COS cells prepared by the method described by Flanagan et al., see above. Alkaline phosphatase-receptor fusion proteins that are not bound to the 35 cells are removed by centrifugation, and the cells are panned on plates coated with antibodies to alkaline phosphatase. Bound

cells are isolated following several washes with a suitable wash reagent, such as 5% fetal bovine serum in PBS, and the DNA is extracted from the cells. Additional details of the panning method described above may be found in an article by Seed et al.,
5 Proc. Natl. Acad. Sci. USA 84, 3365-3369 (1987).

In a second strategy, the putative extracellular ligand binding domains of the receptors are fused to the transmembrane and kinase domains of the human c-fms tyrosine kinase and
10 introduced into 3T3 fibroblasts. The human c-fms kinase is necessary and sufficient to transduce proliferative signals in these cells after appropriate activation i.e. with the Flk1 or Flk2 ligand. The 3T3 cells expressing the chimeras are used to screen putative sources of ligand in a cell proliferation assay.

15

An alternate approach for isolating ligands using the fusion receptor-expressing 3T3 cells and insertional activation is also possible. A retrovirus is introduced into random chromosomal positions in a large population of these cells. In a small fraction, the retrovirus is inserted in the vicinity of the ligand-encoding gene, thereby activating it. These cells proliferate due to autocrine stimulation of the receptor. The ligand gene is "tagged" by the retrovirus, thus facilitating its isolation.

25

Examples

30

Example 1. Cells containing mouse Flk1 and Flk2 ligands. Murine stromal cell line 2018.

35

In order to establish stromal cell lines, fetal liver cells are disaggregated with collagen and grown in a mixture of Dulbecco's Modified Eagle's Medium (DMEM) and 10% heat-inactivated fetal calf serum at 37°C. The cells are immortalized

by standard methods. A suitable method involves introducing DNA encoding a growth regulating- or oncogene-encoding sequence into the target host cell. The DNA may be introduced by means of transduction in a recombinant viral particle or transfection in a 5 plasmid. See, for example, Hammerschmidt et al., *Nature* 340, 393-397 (1989) and Abcouwer et al, *Biotechnology* 7, 939-946 (1989). Retroviruses are the preferred viral vectors, although SV40 and Epstein-Barr virus can also serve as donors of the 10 growth-enhancing sequences. A suitable retrovirus is the ecotropic retrovirus containing a temperature sensitive SV40 T-antigen (tsA58) and a G418 resistance gene described by McKay in *Cell* 66, 713-729 (1991). After several days at 37°C, the 15 temperature of the medium is lowered to 32°C. Cells are selected with G418 (0.5 mg/ml). The selected cells are expanded and maintained.

A mouse stromal cell line produced by this procedure is called 2018 and was deposited on October 30, 1991 in the American Type Culture Collection, Rockville, Maryland, USA (ATCC); 20 accession number CRL 10907.

Example 2. Cells containing human Flk1 and Flk2 ligands.

25 Human fetal liver (18, 20, and 33 weeks after abortion), spleen (18 weeks after abortion), or thymus (20 weeks after abortion) is removed at the time of abortion and stored on ice in a balanced salt solution. After mincing into 1 mm fragments and forcing through a wire mesh, the tissue is washed one time in 30 Hanks Balanced Salt Solution (HBSS).

35 The disrupted tissue is centrifuged at 200 xg for 15 minutes at room temperature. The resulting pellet is resuspended in 10-20 ml of a tissue culture grade trypsin-EDTA solution (Flow Laboratories). The resuspended tissue is transferred to a

sterile flask and stirred with a stirring bar at room temperature for 10 minutes. One ml of heat-inactivated fetal bovine calf serum (Hyclone) is added to a final concentration of 10% in order to inhibit trypsin activity. Collagenase type IV (Sigma) is 5 added from a stock solution (10 mg/ml in HBSS) to a final concentration of 100 ug/ml in order to disrupt the stromal cells. The tissue is stirred at room temperature for an additional 2.5 hours; collected by centrifugation (400xg, 15 minutes); and resuspended in "stromal medium," which contains Iscove's 10 modification of DMEM supplemented with 10% heat-inactivated fetal calf serum, 5% heat-inactivated human serum (Sigma), 4 mM L-glutamine, 1x sodium pyruvate, (stock of 100x Sigma), 1x non-essential amino acids (stock of 100x, Flow), and a mixture of antibiotics kanamycin, neomycin, penicillin, streptomycin. Prior 15 to resuspending the pellet in the stromal medium, the pellet is washed one time with HBSS. It is convenient to suspend the cells in 60 ml of medium. The number of cultures depends on the amount of tissue.

20 Example 3. Isolating Stromal cells

Resuspended Cells (example 2) that are incubated at 37°C with 5% carbon dioxide begin to adhere to the plastic plate 25 within 10-48 hours. Confluent monolayers may be observed within 7-10 days, depending upon the number of cells plated in the initial inoculum. Non-adherent and highly refractile cells adhering to the stromal cell layer as colonies are separately removed by pipetting and frozen. Non-adherent cells are likely 30 sources of populations of self-renewing stem cells containing Flk2. The adherent stromal cell layers are frozen in aliquots for future studies or expanded for growth in culture.

An unexpectedly high level of APtag-Flk2 fusion protein 35 binding to the fetal spleen cells is observed. Two fetal spleen lines are grown in "stromal medium," which is described in

example 2.

5 Non-adherent fetal stem cells attach to the stromal cells and form colonies (colony forming unit - CFU). Stromal cells and CFU are isolated by means of sterile glass cylinders and expanded in culture. A clone, called Fsp 62891, contains the Flk2 ligand. Fsp 62891 was deposited in the American Type Culture Collection, Rockville, Maryland, U.S.A on November 21, 1991, accession number 10

10 CRL 10935.

15 Fetal liver and fetal thymus cells are prepared in a similar way. Both of these cell types produce ligands of Flk1 and, in the case of liver, some Flk2. One such fetal thymus cell line, called F.thy 62891, and one such fetal liver cell line, called FL 15 62891, were deposited in the American Type Culture Collection, Rockville, Maryland, U.S.A on November 21, 1991 and April 2, 1992, respectively, accession numbers CRL 10936 and CRL 11005, respectively.

20 Stable human cell lines are prepared from fetal cells with the same temperature sensitive immortalizing virus used to prepare the murine cell line described in example 1.

Example 4. Isolation of human stromal cell clone

25 Highly refractile cells overgrow patches of stromal cells, presumably because the stromal cells produce factors that allow the formation of the CFU. To isolate stromal cell clones, sterile glass cylinders coated with vacuum grease are positioned over the CFU. A trypsin-EDTA solution (100 ml) is added in order 30 to detach the cells. The cells are added to 5 ml of stromal medium and each (clone) plated in a single well of 6-well plate.

Example 5. Plasmid (AP-taq) for expressing secretable alkaline phosphatase (SEAP)

5 Plasmids that express secretable alkaline phosphatase are described by Flanagan and Leder in Cell 63, 185-194 (1990). The plasmids contain a promoter, such as the LTR promoter; a polylinker, including HindIII and BglII; DNA encoding SEAP; a poly-A signal; and ampicillin resistance gene; and replication 10 site.

Example 6. Plasmid for expressing APtag-Flk2 and APtag-Flk1 fusion proteins

15 Plasmids that express fusion proteins of SEAP and the extracellular portion of either Flk1 or Flk2 are prepared in accordance with the protocols of Flanagan and Leader in Cell 63, 185-194 (1990) and Berger et al., Gene 66, 1-10 (1988). Briefly, 20 a HindIII-Bam HI fragment containing the extracellular portion of Flk1 or Flk2 is prepared and inserted into the HindIII-BglII site of the plasmid described in example 5.

Example 7. Production Of APtag-Flk1 Or -Flk2 Fusion Protein

25 The plasmids from Example 6 are transfected into Cos-7 cells by DEAE-dextran (as described in Current Protocols in Molecular Biology, Unit 16.13, "Transient Expression of Proteins Using Cos Cells," 1991); and cotransfected with a selectable marker, such as pSV7neo, into NIH/3T3 cells by calcium precipitation. The NIH/3T3 cells are selected with 600 μ g/ml G418 in 100 mm plates. Over 300 clones are screened for secretion of placental alkaline phosphatase activity. The assay is performed by heating a portion of the supernatant at 65°C for 10 minutes to inactivate 30 background phosphatase activity, and measuring the OD₄₀₅ after 35 incubating with 1M diethanolamine (pH 9.8), 0.5 mM MgCl₂, 10 mM L-homoarginine (a phosphatase inhibitor), 0.5 mg/ml BSA, and 12

5 mM p-nitrophenyl phosphate. Human placental alkaline phosphatase is used to perform a standard curve. The APtag-Flk1 clones (F-1AP21-4) produce up to 10 µg alkaline phosphatase activity/ml and the APtag-Flk2 clones (F-2AP26-0) produce up to 0.5 µg alkaline phosphatase activity/ml.

Example 8. Assay For APtag-Flk1 Or APtag-Flk2 Binding To Cells

10 The binding of APtag-Flk1 or APtag-Flk2 to cells containing the appropriate ligand is assayed by standard methods. See, for example, Flanagan and Leder, Cell 63:185-194, 1990). Cells (i.e., mouse stromal cells, human fetal liver, spleen or thymus, or various control cells) are grown to confluence in six-well plates and washed with HBHA (Hank's balanced salt solution with 15 0.5 mg/ml BSA, 0.02% NaN₃, 20 mM HEPES, pH 7.0). Supernatants from transfected COS or NIH/3T3 cells containing either APtag-Flk1 fusion protein, APtag-Flk2 fusion protein, or APtag without a receptor (as a control) are added to the cell monolayers and incubated for two hours at room temperature on a rotating 20 platform. The concentration of the APtag-Flk1 fusion protein, APtag-Flk2 fusion protein, or APtag without a receptor is 60 ng/ml of alkaline phosphatase as determined by the standard alkaline phosphatase curve (see above). The cells are then rinsed seven times with HBHA and lysed in 350 µl of 1% Triton X-25 100, 10 mM Tris-HCl (pH 8.0). The lysates are transferred to a microfuge tube, along with a further 150 µl rinse with the same solution. After vortexing vigorously, the samples are 30 centrifuged for five minutes in a microfuge, heated at 65°C for 12 minutes to inactivate cellular phosphatases, and assayed for phosphatase activity as described previously. Results of experiments designed to show the time and dose responses of binding between stromal cells containing the ligands to Flk2 and Flk1 (2018) and APtag-Flk2, APtag-Flk1 and APtag without receptor (as a control) are shown in Figures 3 and 4, respectively.

35

Example 8A. Plasmids for expressing Flk1/fms and Flk2/fms fusion proteins

5 Plasmids that express fusion proteins of the extracellular portion of either Flk1 or Flk2 and the intracellular portion of c-fms (also known as colony-stimulating factor-1 receptor) are prepared in a manner similar to that described under Example 6 (Plasmid for expressing APtag-Flk2 and APtag-Flk1 fusion proteins). Briefly, a Hind III - Bam HI fragment containing the extracellular portion of Flk1 or Flk2 is prepared and inserted into the Hind III - Bgl II site of a pLH expression vector containing the intracellular portion of c-fms.

10

15 8B. Expression of Flk1/fms or Flk2/fms in 3T3 cells

20 The plasmids from Example 8A are transfected into NIH/3T3 cells by calcium. The intracellular portion of c-fms is detected by Western blotting.

25 Example 9. Cloning and Expression of cDNA Coding For Mouse Ligand To Flk1 and Flk2 Receptors

30 cDNA expressing mouse ligand for Flk1 and Flk2 is prepared by known methods. See, for example, Seed, B., and Aruffo, A. PNAS 84:3365-3369, 1987; Simmons, D. and Seed, B. J. Immunol. 141:2797-2800; and D'Andrea, A.D., Lodish, H.F. and Wong, G.G. Cell 57:277-285, 1989).

35 The protocols are listed below in sequence: (a) RNA isolation; (b) poly A RNA preparation; (c) cDNA synthesis; (d) cDNA size fractionation; (e) propagation of plasmids (vector); (f) isolation of plasmid DNA; (g) preparation of vector pSV Sport (BRL Gibco) for cloning; (h) compilation of buffers for the above steps; (i) Transfection of cDNA encoding Ligands in Cos 7 Cells;

(j) panning procedure; (k) Expression cloning of Flk1 or Flk2 ligand by establishment of an autocrine loop.

9a. Guanidinium thiocyanate/LiCl Protocol for RNA Isolation

5

For each ml of mix desired, 0.5 g guanidine thiocyanate (GuSCN) is dissolved in 0.55 ml of 25% LiCl (stock filtered through 0.45 micron filter). 20 μ l of mercaptoethanol is added. (The resulting solution is not good for more than about a week at room temperature.)

The 2018 stromal cells are centrifuged, and 1 ml of the solution described above is added to up to 5×10^7 cells. The cells are sheared by means of a polytron until the mixture is non-viscous. For small scale preparations ($<10^8$ cells), the sheared mixture is layered on 1.5 ml of 5.7M CsCl (RNase free; 1.26 g CsCl added to every ml 10 mM EDTA pH8), and overlaid with RNase-free water if needed. The mixture is spun in an SW55 rotor at 50 krpm for 2 hours. For large scale preparations, 25 ml of the mixture is layered on 12 ml CsCl in an SW28 tube, overlaid as above, and spun at 24 krpm for 8 hours. The contents of the tube are aspirated carefully with a sterile pasteur pipet connected to a vacuum flask. Once past the CsCl interface, a band around the tube is scratched with the pipet tip to prevent creeping of the layer on the wall down the tube. The remaining CsCl solution is aspirated. The resulting pellet is taken up in water, but not redissolved. 1/10 volume of sodium acetate and three volumes of ethanol are added to the mixture, and spun. The pellet is resuspended in water at 70°C, if necessary. The concentration of the RNA is adjusted to 1 mg/ml and frozen.

It should be noted that small RNA molecules (e.g., 5S) do not come down. For small amounts of cells, the volumes are scaled down, and the mixture is overlaid with GuSCN in RNase-free water on a gradient (precipitation is inefficient when RNA is

dilute).

9b. Poly A⁺ RNA preparation

5 (All buffers mentioned are compiled separately below)
A disposable polypropylene column is prepared by washing
with 5M NaOH and then rinsing with RNase-free water. For each
milligram of total RNA, approximately 0.3 ml (final packed bed)
of oligo dT cellulose is added. The oligo dT cellulose is
10 prepared by resuspending approximately 0.5 ml of dry powder in 1
ml of 0.1M NaOH and transferring it into the column, or by
percolating 0.1M NaOH through a previously used column. The
column is washed with several column volumes of RNase-free water
until the pH is neutral, and rinsed with 2-3 ml of loading
15 buffer. The column bed is transferred to a sterile 15 ml tube
using 4-6 ml of loading buffer.

20 Total RNA from the 2018 cell line is heated to 70°C for 2-3
minutes. LiCl from RNase-free stock is added to the mixture to a
final concentration of 0.5M. The mixture is combined with oligo
dT cellulose in the 15 ml tube, which is vortexed or agitated for
10 minutes. The mixture is poured into the column, and washed
with 3 ml loading buffer, and then with 3 ml of middle wash
25 buffer. The mRNA is eluted directly into an SW55 tube with 1.5
ml of 2 mM EDTA and 0.1% SDS, discarding the first two or three
drops.

30 The eluted mRNA is precipitated by adding 1/10 volume of 3M
sodium acetate and filling the tube with ethanol. The contents
of the tube are mixed, chilled for 30 minutes at -20°C, and spun
at 50 k rpm at 5°C for 30 minutes. After the ethanol is decanted,
and the tube air dried, the mRNA pellet is resuspended in 50-100
35 µl of RNase-free water. 5 µl of the resuspended mRNA is heated
to 70°C in MOPS/EDTA/formaldehyde, and examined on an RNase-free
1% agarose gel.

9c. cDNA Synthesis

The protocol used is a variation of the method described by Gubler and Hoffman in *Gene* 25, 263-270 (1983).

5

1. First Strand. 4 µg of mRNA is added to a microfuge tube, heated to approximately 100°C for 30 seconds, quenched on ice. The volume is adjusted to 70µl with RNase-free water. 20 µl of RT1 buffer, 2 µl of RNase inhibitor (Boehringer 36 u/µl), 1 µl of 5 µg/µl of oligo dT (Collaborative Research), 2.5 µl of 20 mM dXTP's (ultrapure - US Biochemicals), 1 µl of 1M DTT and 4 µl of RT-XL (Life Sciences, 24 u/µl) are added. The mixture is incubated at 42°C for 40 minutes, and inactivated by heating at 70°C for 10 minutes.

10

2. Second Strand. 320 µl of RNase-free water, 80 µl of RT2 buffer, 5 µl of DNA Polymerase I (Boehringer, 5 U/µl), 2 µl RNase H (BRL 2 u/µl) are added to the solution containing the first strand. The solution is incubated at 15°C for one hour and at 22°C for an additional hour. After adding 20 µl of 0.5M EDTA, pH 8.0, the solution is extracted with phenol and precipitated by adding NaCl to 0.5M linear polyacrylamide (carrier) to 20 µg/ml, and filling the tube with EtOH. The tube is spun for 2-3 minutes in a microfuge, vortexed to dislodge precipitated material from the wall of the tube, and respun for one minute.

15

20

25

3. Adaptors. Adaptors provide specific restriction sites to facilitate cloning, and are available from BRL Gibco, New England Biolabs, etc. Crude adaptors are resuspended at a concentration of 1 µg/µl. MgSO₄ is added to a final concentration of 10 mM, followed by five volumes of EtOH. The resulting precipitate is rinsed with 70% EtOH and resuspended in TE at a concentration of 1 µg/µl. To kinase, 25 µl of resuspended adaptors is added to 3 µl of 10X kinasing buffer and 20 units of kinase. The mixture is incubated at 37°C overnight. The precipitated cDNA is

30

35

resuspended in 240 μ l of TE (10/1). After adding 30 μ l of 10X low salt buffer, 30 μ l of 10X ligation buffer with 0.1mM ATP, 3 μ l (2.4 μ g) of kinased 12-mer adaptor sequence, 2 μ l (1.6 μ g) of kinased 8-mer adaptor sequence, and 1 μ l of T4 DNA ligase (BioLabs, 400 u/ μ l, or Boehringer, 1 Weiss unit ml), the mixture is incubated at 15°C overnight. The cDNA is extracted with phenol and precipitated as above, except that the extra carrier is omitted, and resuspended in 100 μ l of TE.

10 9d. cDNA Size Fractionation.

A 20% KOAc, 2 mM EDTA, 1 μ g/ml ethidium bromide solution and a 5% KOAc, 2 mM EDTA, 1 μ g/ml ethidium bromide solution are prepared. 2.6 ml of the 20% KOAc solution is added to the back chamber of a small gradient maker. Air bubbles are removed from the tube connecting the two chambers by allowing the 20% solution to flow into the front chamber and forcing the solution to return to the back chamber by tilting the gradient maker. The passage between the chambers is closed, and 2.5 ml of 5% solution is added to the front chamber. Any liquid in the tubing from a previous run is removed by allowing the 5% solution to flow to the end of the tubing, and then to return to its chamber. The apparatus is placed on a stirplate, and, with rapid stirring, the topcock connecting the two chambers and the front stopcock are opened. A polyallomer 5W55 tube is filled from the bottom with the KOAc solution. The gradient is overlaid with 100 μ l of cDNA solution, and spun for three hours at 50k rpm at 22°C. To collect fractions from the gradient, the SW55 tube is pierced close to the bottom of the tube with a butterfly infusion set (with the luer hub clipped off). Three 0.5 ml fractions and then six 0.25 ml fractions are collected in microfuge tubes (approximately 22 and 11 drops, respectively). The fractions are precipitated by adding linear polyacrylamide to 20 μ g/ml and filling the tube to the top with ethanol. The tubes are cooled, spun in a microfuge tube for three minutes, vortexed, and respun

for one minute. The resulting pellets are rinsed with 70% ethanol and respun, taking care not to permit the pellets to dry to completion. Each 0.25 ml fraction is resuspended in 10 μ l of TE, and 1 μ l is run on a 1% agarose minigel. The first three fractions, and the last six which contain no material smaller than 1 kb are pooled.

9e. Propagation of Plasmids

SupF plasmids are selected in nonsuppressing bacterial hosts containing a second plasmid, p3, which contains amber mutated ampicillin and tetracycline drug resistance elements. See Seed, Nucleic Acids Res., 11, 2427-2445 (1983). The p3 plasmid is derived from RPL, is 57 kb in length, and is a stably maintained, single copy episome. The ampicillin resistance of this plasmid reverts at a high rate so that amp^r plasmids usually cannot be used in p3-containing strains. Selection for tetracycline resistance alone is almost as good as selection for ampicillin-tetracycline resistance. However, spontaneous appearance of chromosomal suppressor tRNA mutations presents an unavoidable background (frequency about 10⁻⁹) in this system. Colonies arising from spontaneous suppressor mutations are usually larger than colonies arising from plasmid transformation. Suppressor plasmids are selected in Luria broth (LB) medium containing ampicillin at 12.5 μ g/ml and tetracycline at 7.5 μ g/ml. For scaled-up plasmid preparations, M9 Casamino acids medium containing glycerol (0.8%) is employed as a carbon source. The bacteria are grown to saturation.

Alternatively, pSV Sport (BRL, Gaithersberg, Maryland) may be employed to provide SV40 derived sequences for replication, transcription initiation and termination in COS 7 cells, as well as those sequences necessary for replication and ampicillin resistance in E. coli.

35

9f. Isolation of Vector DNA/Plasmid

One liter of saturated bacterial cells are spun down in J6 bottles at 4.2k rpm for 25 minutes. The cells are resuspended in 40 ml 10 mM EDTA, pH 8. 80 ml 0.2M NaOH and 1% SDS are added, and the mixture is swirled until it is clear and viscous. 40 ml 5M KOAc, pH 4.7 (2.5M KOAc, 2.5M HOAc) is added, and the mixture is shaken semi-vigorously until the lumps are approximately 2-3 mm in size. The bottle is spun at 4.2k rpm for 5 minutes. The supernatant is poured through cheesecloth into a 250 ml bottle, which is then filled with isopropyl alcohol and centrifuged at 4.2k rpm for 5 minutes. The bottle is gently drained and rinsed with 70% ethanol, taking care not to fragment the pellet. After inverting the bottle and removing traces of ethanol, the mixture is resuspended in 3.5 ml Tris base/EDTA (20 mM/10 mM). 3.75 ml of resuspended pellet and 0.75 ml 10 mg/ml ethidium bromide are added to 4.5 g CsCl. VTi80 tubes are filled with solution, and centrifuged for at least 2.5 hours at 80k rpm. Bands are extracted by visible light with 1 ml syringe and 20 gauge or lower needle. The top of the tube is cut off with scissors, and the needle is inserted upwards into the tube at an angle of about 30 degrees with respect to the tube at a position about 3 mm beneath the band, with the bevel of the needle up. After the band is removed, the contents of the tube are poured into bleach. The extracted band is deposited in a 13 ml Sarstedt tube, which is then filled to the top with n-butanol saturated with 1M NaCl extract. If the amount of DNA is large, the extraction procedure may be repeated. After aspirating the butanol into a trap containing 5M NaOH to destroy ethidium, an approximately equal volume of 1M ammonium acetate and approximately two volumes of 95% ethanol are added to the DNA, which is then spun at 10k rpm for 5 minutes. The pellet is rinsed carefully with 70% ethanol, and dried with a swab or lyophilizer.

9g. Preparation of Vector for Cloning

20 μ g of vector is cut in a 200 μ l reaction with 100 units of BstXI (New York Biolabs) at 50°C overnight in a well
5 thermostated, circulating water bath. Potassium acetate
solutions (5 and 20%) are prepared in 5W55 tubes as described
above. 100 μ l of the digested vector is added to each tube and
spun for three hours, 50k rpm at 22°C. Under 300 nm UV light,
10 the desired band is observed to migrate 2/3 of the length of the
tube. Forward trailing of the band indicates that the gradient
is overloaded. The band is removed with a 1 ml syringe fitted
with a 20 gauge needle. After adding linear polyacrylamide and
precipitating the plasmid by adding three volumes of ethanol, the
plasmid is resuspended in 50 μ l of TE. Trial ligations are
carried out with a constant amount of vector and increasing
amounts of cDNA. Large scale ligation are carried out on the
basis of these trial ligations. Usually the entire cDNA prep
requires 1-2 μ g of cut vector.

20 9h. Buffers

Loading Buffer:.5M LiCl, 10 mM Tris pH 7.5, 1 mM EDTA .1% SDS.

Middle Wash Buffer:.15M LiCl, 10 mM Tris pH 7.5, 1 mM EDTA .1%
SDS.

25 RT1 Buffer:.25M Tris pH 8.8 (8.2 at 42°), .25M KCl, 30 mM MgCl₂.

RT2 Buffer:.1M Tris pH 7.5, 25 mM MgCl₂, .5M KCl, .25 mg/ml BSA,
50 mM dithiothreitol (DTT).

10X Low Salt:60 mM Tris pH 7.5, 60 mM MgCl₂, 50 mM NaCl, 2.5
mg/ml BSA 70 mM DME

30 10X Ligation Additions:1 mM ATP, 20 mM DTT, 1 mg/ml BSA 10 mM
spermidine.

10X Kinasing Buffer:.5M Tris pH 7.5, 10 mM ATP, 20 mM DTT, 10 mM
spermidine, 1 mg/ml BSA 100 mM MgCl₂

9i. Transfection of cDNA encoding Ligands in Cos 7 Cells

Cos 7 cells are split 1:5 into 100 mm plates in Dulbecco's modified Eagles medium (DME)/10% fetal calf serum (FCS), and allowed to grow overnight. 3 ml Tris/DME (0.039M Tris, pH 7.4 in DME) containing 400 µg/ml DEAE-dextran (Sigma, D-9885) is prepared for each 100 mm plate of Cos 7 cells to be transfected. 10 µg of plasmid DNA preparation per plate is added. The medium is removed from the Cos-7 cells and the DNA/DEAE-dextran mixture is added. The cells are incubated for 4.5 hours. The medium is removed from the cells, and replaced with 3 ml of DME containing 2% fetal calf serum (FCS) and 0.1 mM chloroquine. The cells are incubated for one hour. After removing the chloroquine and replacing with 1.5 ml 20% glycerol in PBS, the cells are allowed to stand at room temperature for one minute. 3 ml Tris/DME is added, and the mixture is aspirated and washed two times with Tris/DME. 10 ml DME/10% FCS is added and the mixture is incubated overnight. The transfected Cos 7 cells are split 1:2 into fresh 100 mm plates with (DME)/10% FCS and allowed to grow.

9j. Panning Procedure for Cos 7 cells Expressing Ligand1) Antibody-coated plates:

Bacteriological 100 mm plates are coated for 1.5 hours with rabbit anti-human placental alkaline phosphatase (Dako, California) diluted 1:500 in 10 ml of 50 mM Tris.HCl, pH 9.5. The plates are washed three times with 0.15M NaCl, and incubated with 3 mg BSA/ml PBS overnight. The blocking solution is aspirated, and the plates are utilized immediately or frozen for later use.

2) Panning cells:

5 The medium from transfected Cos 7 cells is aspirated, and 3 ml PBS/0.5 mM EDTA/0.02% sodium azide is added. The plates are
10 incubated at 37°C for thirty minutes in order to detach the cells. The cells are triturated vigorously with a pasteur pipet and collected in a 15 ml centrifuge tube. The plate is washed with a further 2 ml PBS/EDTA/azide solution, which is then added to the centrifuge tube. After centrifuging at 200 xg for five
15 minutes, the cells are resuspended in 3 ml of APtaq-Flk1 (F-1AP21-4) or Flk2 (F-2AP26-0) supernatant from transfected NIH/3T3 cells (see Example 7.), and incubated for 1.5 hours on ice. The cells are centrifuged again at 200 xg for five minutes. The supernatant is aspirated, and the cells are resuspended in 3 ml
20 PBS/EDTA/azide solution. The cell suspension is layered carefully on 3 ml PBS/EDTA/azide/2% Ficoll, and centrifuged at 200 xg for four minutes. The supernatant is aspirated, and the cells are resuspended in 0.5 ml PBS/EDTA/azide solution. The cells are added to the antibody-coated plates containing 4 ml
25 PBS/EDTA/azide/5% FBS, and allowed to stand at room temperature one to three hours. Non-adhering cells are removed by washing gently two or three times with 3 ml PBS/5% FBS.

3) Hirt Supernatant:

25 0.4 ml 0.6% SDS and 10 mM EDTA are added to the panned plates, which are allowed to stand 20 minutes. The viscous mixture is added by means of a pipet into a microfuge tube. 0.1 ml 5M NaCl is added to the tube, mixed, and chilled on ice for at
30 least five hours. The tube is spun for four minutes, and the supernatant is removed carefully. The contents of the tube are extracted with phenol once, or, if the first interface is not clean, twice. Ten micrograms of linear polyacrylamide (or other carrier) is added, and the tube is filled to the top with
35 ethanol. The resulting precipitate is resuspended in 0.1 ml

water or TE. After adding 3 volumes of EtOH/NaOAc, the cells are reprecipitated and resuspended in 0.1 ml water or TE. The cDNA obtained is transfected into any suitable E. coli host by electroporation. Suitable hosts are described in various catalogs, and include MC1061/p3 or Electromax DH10B Cells of BRL Gibco. The cDNA is extracted by conventional methods.

5 The above panning procedure is repeated until a pure E. coli clone bearing the cDNA as a unique plasmid recombinant capable of 10 transfecting mammalian cells and yielding a positive panning assay is isolated. Normally, three repetitions are sufficient.

15 9k. Expression cloning of Flk1 or Flk2 ligand by establishment of an autocrine loop

Cells expressing Flk1/fms or Flk2/fms (Example 10) are transfected with 20-30 μ g of a cDNA library from either Flk1 ligand or Flk2 ligand expressing stromal cells, respectively. 20 The cDNA library is prepared as described above (a-h). The cells are co-transfected with 1 μ g pLTR neo cDNA. Following transfection the cells are passaged 1:2 and cultured in 800 μ g/ml of G418 in Dulbecco's medium (DME) supplemented with 10% CS. 25 Approximately 12 days later the colonies of cells are passaged and plated onto dishes coated with poly -D- lysine (1 mg/ml) and human fibronectin (15 μ g/ml). The culture medium is defined serum-free medium which is a mixture (3:1) of DME and Ham's F12 medium. The medium supplements are 8 mM NaHCO₃, 15 mM HEPES pH 7.4, 3 mM histidine, 4 μ M MnCl₂, 10 μ M ethanolamine, 0.1 μ M 30 selenous acid, 2 μ M hydrocortisone, 5 μ g/ml transferrin, 500 μ g/ml bovine serum albumin/linoleic acid complex, and 20 μ g/ml insulin (Ref. Zhan, X, et al. Oncogene 1: 369-376, 1987). The cultures are refed the next day and every 3 days until the only 35 cells capable of growing under the defined medium condition remain. The remaining colonies of cells are expanded and tested for the presence of the ligand by assaying for binding of APtag -

Flk1 or APtag - Flk2 to the cells (as described in Example 8). The DNA would be rescued from cells demonstrating the presence of the Flk1 or Flk2 ligand and the sequence.

5 Example 10. Expression of Ligand cDNA

The cDNA is sequenced, and expressed in a suitable host cell, such as a mammalian cell, preferably COS, CHO or NIH/3T3 cells. The presence of the ligand is confirmed by demonstrating 10 binding of the ligand to APtag-Flk2 fusion protein (see above).

Example 11. Chemical Cross Linking of Receptor and Ligand

Cross linking experiments are performed on intact cells 15 using a modification of the procedure described by Blume-Jensen et al et al., EMBO J., 10, 4121-4128 (1991). Cells are cultured in 100mm tissue culture plates to subconfluence and washed once with PBS-0.1% BSA.

20 To examine chemical cross linking of soluble receptor to membrane-bound ligand, stromal cells from the 2018 stromal cell line are incubated with conditioned media (CM) from transfected 3T3 cells expressing the soluble receptor Flk2-APtag. Cross linking studies of soluble ligand to membrane bound receptor are 25 performed by incubating conditioned media from 2018 cells with transfected 3T3 cells expressing a Flk2-fms fusion construct.

30 Binding is carried out for 2 hours either at room temperature with CM containing 0.02% sodium azide to prevent receptor internalization or at 4°C with CM (and buffers) supplemented with sodium vanadate to prevent receptor dephosphorylation. Cells are washed twice with PBS-0.1% BSA and four times with PBS.

35 Cross linking is performed in PBS containing 250 mM

disuccinimidyl suberate (DSS; Pierce) for 30 minutes at room temperature. The reaction is quenched with Tris-HCl pH 7.4 to a final concentration of 50 mM.

5 Cells are solubilized in solubilization buffer: 0.5% Triton - X100, 0.5% deoxycholic acid, 20 mM Tris pH 7.4, 150 mM NaCl, 10mM EDTA, 1mM PMSF, 50 mg/ml aprotinin, 2 mg/ml bestatin, 2 mg/ml pepstatin and 10mg/ml leupeptin. Lysed cells are immediately transferred to 1.5 ml Nalgene tubes and solubilized
10 by rolling end to end for 45 minutes at 4°C. Lysates are then centrifuged in a microfuge at 14,000g for 10 minutes. Solubilized cross linked receptor complexes are then retrieved from lysates by incubating supernatants with 10% (v/v) wheat germ
15 lectin-Sepharose 6MB beads (Pharmacia) at 4°C for 2 hours or overnight.

20 Beads are washed once with Tris-buffered saline (TBS) and resuspended in 2X SDS-polyacrylamide nonreducing sample buffer. Bound complexes are eluted from the beads by heating at 95°C for 5 minutes. Samples are analyzed on 4-12% gradient gels (NOVEX) under nonreducing and reducing conditions (0.35 M 2-
25 mercaptoethanol) and then transferred to PVDF membranes for 2 hours using a Novex blotting apparatus. Blots are blocked in TBS-3% BSA for 1 hour at room temperature followed by incubation with appropriate antibody.

30 Cross linked Flk2-APtag and Flk2-fms receptors are detected using rabbit polyclonal antibodies raised against human alkaline phosphatase and fms protein, respectively. The remainder of the procedure is carried out according to the instructions provided in the ABC Kit (Pierce). The kit is based on the use of a biotinylated secondary antibody and avidin-biotinylated horseradish peroxidase complex for detection.

Example 12. Expression and purification of Flag-Flk2.

1. Design of the Flag-Flk2 expression plasmids.

5 A synthetic DNA fragment (Fragment 1) is synthesized using complementary oligonucleotides BP1 and BP2 (see below and SEQ. ID. NOS. 7 and 8). The fragment encoded the following features in the 5' to 3' order: Sal I restriction site, 22 base pair (bp) 5' untranslated region containing an eukaryotic ribosome binding site, an ATG initiation codon, preprotrypsinogen signal sequence, 10 coding region for the FLAG peptide (DYKDDDDKI) and Bgl II restriction site.

15 A cDNA fragment (Fragment 2) encoding Asn 27 to Ser 544 of murine Flk2 is obtained by polymerase chain reaction (PCR) using primers designed to introduce an in frame Bgl II site at the 5' end (oligonucleotide BP5, see below and SEQ. ID. NO. 9) and a termination codon followed by a Not I site at the 3' end (oligonucleotide BP10, see below and SEQ. ID. NO. 10). The 20 template for the PCR reaction is full length Flk2 cDNA (Matthews et al., Cell 65:1143 (1991)). Fragment 2 is extensively digested with Bgl II and Not I restriction enzymes prior to ligation.

25 To assemble the complete Flag-Flk2 gene, Fragments 1 and 2 are ligated in a tripartate ligation into Sal I and Not I digested plasmid pSPORT (Gibco/BRL, Grand Island, NY) to give the plasmid pFlag-Flk2.

30 Preferably, the Flag-Flk2 protein is attached at either end to the Fc portion of an immunoglobulin (Ig). The Ig is preferably attached to the Flk2 portion of the Flag-Flk2 protein. To assemble the construct pFlag-Flk2-Ig, the sequences coding for the CH¹ domain of human immunoglobulin G (IgG¹) are placed downstream of the Flk2 coding region in the plasmid pFlag-Flk2 as 35 per the method described by Zettlmeissl et al., DNA and Cell

Biology 9: 347-352 (1990).

The sequences of oligonucleotides used to construct the Flag-Flk2 gene are given below:

5

Oligonucleotide BP1:

5' -AATTCGTCGACTTCTGTACCATGAGTGCACCTCTGATCCTAGCCCTTGTG
GGAGCTGCTGTTGCTGACTACAAAGATGATGATGACAAGATCTA- 3'

10

Oligonucleotide BP2:

5' -AGCTTAGATCTTGTCAATCATCATCTTGTAGTCAGAACAGCAGCTCCCACA
AGGGCTAGGATCAGAAGTGCACTCATGGTGACAGAAAGTCGACG- 3'

Oligonucleotide BP5:

15 5' -TGAGAAGATCTAAACCAAGACCTGCCTGT- 3'

Oligonucleotide BP10:

5' -CCAATGGCGGCCGCTCAGGAGATGTTGTCTTGG- 3'

20

(See SEQ. ID. NOS. 7-10, respectively)

2. Expression of the Flag-Flk2 construct.

25

For transient expression of the Flag-Flk2 construct, the SalI to Not I fragment from pFlag-Flk2 is subcloned into the plasmid pSVSPORT (Gibco/BRL) to give the plasmid pSVFlag-Flk2. For expression of the Flag-Flk2 protein pSVFlag-Flk2 is transfected into COS monkey cells using the DEAE-dextran method.

30

For stable expression in eukaryotic cells, the Sal I-Not I fragment of pFlag-Flk2 is cloned into the EcoRV and Not I sites of the plasmid pCDNA I/Neo (Invitrogen Co., San Diego, CA). The Sal I 3' recessed terminus of pFlag-Flk2 is filled with the Klenow fragment of DNA polymerase I and a mixture of

deoxyribonucleotides to make the site compatible with the EcoRV site of the vector. The resulting construct is introduced into cultured mammalian cells using either the Lipofectin (Gibco/BRL) or the calcium phosphate methods.

5

For expression in insect cells, the SalI to Hind III (from pSPORT polylinker) fragment of pFlag-Flk2 is subcloned into the BamHI-Hind III sites of the baculovirus transfer vector pBlueBac III (Invitrogen). The vector Bam HI site and the insert Sal I site are blunted with Klenow (see above). Production of the recombinant virus and infection of the Sf9 insect cells is performed as per manufacturers directions (Invitrogen).

10 Expression of the Flag-Flk2 protein is detected by Western blotting of SDS-PAGE separated conditioned media (mammalian cells) or cell lysates (insect cells) with the anti-Flag monoclonal antibody (mAb) M1 (International Biotechnology, Inc. [IBI], New Haven, CT).

15 3. Affinity purification of the Flag-Flk2 protein from conditioned media or insect cell lysates is performed using immobilized mAb M1 (IBI) as per manufacturers specifications.

20 3.1 Affinity purification of the Flag-Flk2-Ig¹ protein from conditioned media is performed using immobilized Protein A (Pharmacia LKB, Piscataway, NJ) as per the manufacturers instructions.

II. Use of the Flag-Flk2 protein to search for the Flk2 ligand.

30

1. Binding and cross-linking studies to detect membrane-bound ligand:

A. Binding studies.

35

Murine stromal lines (eg. 2018 cells ATCC CRL 10907 (see below), see example 1, *supra*) considered to be candidates for expression of the Flk2 ligand were deposited at the American Type Culture Collection, ATCC CRL 10907 (see below) and cultured in Dulbecco's modified Eagles medium (DMEM; Gibco/BRL) supplemented with 10% fetal calf serum. The cells are grown to confluence in 10 cm plates and washed once with PBS. Conditioned media containing Flag-Flk2 is incubated with the cells at 4°C for 2 hrs. The cell monolayers are rinsed extensively to remove the non-bound protein, solubilized and centrifuged to remove insoluble cellular material. Glycoproteins in the lysates are partially purified with wheat germ agglutinin-Sepharose (Pharmacia LKB, Piscataway, NJ), boiled in an SDS sample buffer, separated on SDS-PAGE gels and transferred to nitrocellulose membranes. The membranes are probed with the M1 antibody to detect the presence of cell-associated Flag-Flk2 protein.

B. In a cross-linking study, the above protocol is followed except that prior to solubilization the monolayer are treated with the crosslinker disuccinimidyl suberate (DSS; Pierce, Rockford, IL). The presence of a putative ligand is detected by an upward shift in the apparent molecular weight of the Flag-Flk2 band on Western blots.

C. Purified Flag-Flk2 protein labelled with Na¹²⁵I via the Chloramine T method is used to asses the ability of the soluble extracellular domain of the Flk2 receptor to bind transmembrane form of the Flk2 ligand in cultured stromal lines. The labelled protein is added to monolayers of stromal cells on ice for 2 hr in the presence or absence of excess unlabelled protein. Specific binding is calculated by subtracting counts bound in the presence of excess unlabelled protein from the total counts bound.

2. Use of the Flag-Flk2 protein to search for secreted form of the ligand.

A. The Flag-Flk2 protein is used in attempts to identify the Flk2 ligand in conditioned media from stromal cell cultures via modification of the direct N-terminal sequencing method of Pan et al., Bioch. Biophys. Res. Comm. 166:201 (1990). Briefly, 5 the Flag-Flk2 protein N-terminally sequenced by automatic Edman degradation chemistry an an ABI 477A sequencer with on line PTH amino acid analysis. Approximately 15 amino acids are determined. The protein is then immobilized on NuGel PAF silica beads via free NH₄⁺ groups. The immobilized Flag-Flk2 is 10 incubated with conditioned media from putative ligand-producing cells for 30 min at 4°C and washed free off non-bound proteins with phosphate buffered saline adjusted to 2M NaCl. The resulting protein complex is resequenced. For each sequencing cycle, any 15 amino acid not expected at this position in the FLAG-Flk2 protein is considered as possibly originating from a protein complexed to the Flk2 receptor.

B. For conventional affinity chromatography, the Flag-Flk2 protein is immobilized on a stable support such as Sepharose. 20 35S-methionine labelled-conditioned media from stromal cell lines are passed over the affinity matrix and bound material is analyzed by SDS-PAGE gel electrophoresis and autoradiography.

3. Use of the Flag-Flk2 protein in expression cloning 25 experiments.

A method of expression cloning of integral membrane proteins in COS cells has been described (Aruffo and Seed, Proc. Natl. Acad. Sci. 84:8573 (1987)). A cDNA library is prepared from an 30 appropriate stromal cell line such as 2018 and is transfected into COS cells. Cells transiently expressing the Flk2 ligand are affinity adsorbed onto plastic plates coated with the Flag-Flk2 protein. The cells are lysed, the plasmid DNA is recovered and 35 amplified in a bacterial host. The cycle of transfection into COS cells is repeated until a single cDNA clone encoding the ligand

molecule is isolated.

In a modification of the above technique, pools of transfected COS cells are screened for binding of ^{125}I -Flag-Flk2. Positive cells pools are selected and plasmid DNA is recovered and amplified in *E. coli*. The resulting DNA preparation is used in subsequent rounds of transfection and transient expression until all cells are positive for binding of ^{125}I -Flag-Flk2. The cDNA in the final plasmid preparation is then sequenced to determine the sequence of the putative Flk-2 ligand.

Example 13 Isolating the Human Flk2 Ligand from PHA-LCM

13a. Source of the human Flk2 ligand

The Flk2 ligand is isolated from tissue culture medium conditioned by phytohemagglutinin-stimulated human peripheral blood leukocytes (PHA-LCM). The medium is prepared by isolating normal human peripheral blood mononuclear cells (leukocytes) from whole blood by density centrifugation (Ficoll-Hypaque, Pharmacia Biotech, Inc, Piscataway, NJ) and incubating these cells at a concentration of 2×10^6 cells/ml with the lectin phytohemagglutinin (PHA, Gibco Laboratories, Grand Island, NY) in a commercially-prepared, serum-free defined culture medium (AIMV; Gibco Laboratories, Grand Island, NY) for one week. PHA-LCM is harvested by removal of cells and debris by centrifugation.

13b. Isolating the human Flk2 ligand from PHA-LCM

The Flk2 ligand is one of a large number of proteins that are specifically secreted by PHA-activated cells into the medium. Several purification steps using conventional chromatographic techniques are required to isolate the Flk2 ligand. The chromatographic columns used (not listed in specific order) include: Blue Sepharose Fast Flow (Pharmacia Biotech, Inc,

Piscataway, NJ) to remove the medium component albumin, anion exchange (Q-Sepharose Fast Flow, Pharmacia Biotech, Inc, Piscataway, NJ), cation exchange (S-Sepharose Fast Flow, Pharmacia Biotech, Inc, Piscataway, NJ), gel filtration (Superdex 5, Pharmacia Biotech, Inc, Piscataway, NJ), heparin sepharose (Pharmacia Biotech, Inc, Piscataway, NJ), ConA (Pharmacia Biotech, Inc, Piscataway, NJ), wheat germ agglutinin (Pharmacia Biotech, Inc, Piscataway, NJ), and C4 reverse phase (Vydac, The Separations Group, Hesperia, CA).

10

Biological assays are used throughout the purification to identify which column fractions contain the Flk2 ligand. The Flk2 ligand specifically stimulates proliferation *in vitro* of cell lines transfected with constructs expressing the full length Flk2 receptor or a chimeric receptor comprising of the extracellular domain of the Flk2 receptor and the intracellular domain of a different protein tyrosine kinase receptor such as fms, the receptor for CSF-1. For example, the Flk2 ligand specifically stimulates proliferation of murine NIH 3T3 fibroblast cell line transfected with constructs expressing the murine or human Flk2 receptor in either full length or chimeric form (see example 8B). The parent untransfected 3T3 cells do not respond to the Flk2 ligand. The format of the Flk2 receptor 3T3 cell assay uses 96 well tissue culture plates (Becton Dickinson, Lincoln Park, NJ), where column fractions or other test samples are serially diluted across the plates in wells containing a mixture of AIMV and Dulbecco's modification of Eagle's medium (DMEM, Gibco Laboratories, Grand Island, NY). Samples are tested for their ability to stimulate proliferation of Flk2 receptor 3T3 cells initially cultured at 3×10^4 cells/well. Survival of Flk2 receptor 3T3 cells is dependent on the presence of the Flk2 ligand. Viable Flk2 receptor 3T3 cells are quantitated after three to five days in culture either visually or spectrophotometrically (Molecular Devices 15 Corporation, Menlo Park, CA) using a tetraformazan salt (XTT, 20 25 30 35

Diagnostic Chemicals Ltd, Oxford, CT) that when cleaved by actively respiring cells forms diformazan salt which absorbs light at a wavelength (450 nm) that is different from the starting compound (560 nm). Relative (units/ml) and specific (units/mg) activities are defined as the reciprocal dilution at which half-maximal stimulation is detected.

13c. Physical properties of the human Flk2 ligand

The human Flk2 ligand isolated from PHA-LCM is a glycosylated protein and has an apparent molecular weight of 18 kDa, as determined by SDS-PAGE analysis run under reducing (β -mercaptoethanol) and non-reducing conditions. Its N-terminal fourteen amino acid sequence is A Q S L S F X F T K F D L D, wherein X is any amino acid. (See SEQ. ID. NO. 11) Its biological activity is inactivated at 100° C but not 60° C in five minutes and the activity is retained after the Flk2 ligand is subjected to a pH of 2.8 at room temperature for two hours.

The 18 kDa Flk2 ligand may act alone, in combination with other cytokines (e.g., interleukin 1, interleukin 3, interleukin 6, interleukin 11 or the kit ligand), or as a component of a complex of proteins that stimulate the Flk2 receptor in transfected 3T3 cell or in primitive hematopoietic progenitors. The complex of proteins may include a soluble or membrane-bound form of the Flk2 receptor.

A radiolabeled form of the Flk2 ligand may be used to detect and to measure the levels of Flk2 receptor, such as the soluble form of the Flk2 receptor, for example, in serum or urine of patients with bone marrow disorders.

13d. Biological activity of the human Flk2 ligand

In addition to acting on Flk2 receptor-expressing 3T3 cells,

the Flk2 ligand specifically stimulates proliferation of cells that naturally express the Flk2 receptor. In assays using either a human myeloid cell line or a subset of primitive hematopoietic progenitors expressing the surface phenotype CD34, the Flk2 ligand promotes proliferation but not differentiation into mature progeny. These observations suggest that the Flk2 ligand alone or in combination with other cytokines (e.g. Interleukin 1, Interleukin 3, Interleukin 6, Interleukin 11, or the kit ligand) may act to preserve or expand primitive hematopoietic progenitors 5 *in vitro* and *in vivo*.
10

SUPPLEMENTAL ENABLEMENT

15 The invention as claimed is enabled in accordance with the above specification and readily available references and starting materials. Nevertheless, Applicants have deposited with the American Type Culture Collection, Rockville, Md., USA (ATCC) the cell lines listed below:

20 2018, ATCC accession no. CRL 10907, deposited October 30, 1991.

25 Fsp 62891, ATCC accession no. CRL 10935, deposited November 21, 1991.

FL 62891, ATCC accession no. CRL 11005, deposited April 2, 1992.

30 35 These deposits were made under the provisions of the Budapest Treaty on the International Recognition of the Deposit of Microorganisms for the Purposes of Patent Procedure and the regulations thereunder (Budapest Treaty). This assures

5 maintenance of a viable culture for 30 years from date of deposit. The organisms will be made available by ATCC under the terms of the Budapest Treaty, and subject to an agreement between Applicants and ATCC which assures unrestricted availability upon issuance of the pertinent U.S. patent. Availability of the deposited strains is not to be construed as a license to practice the invention in contravention of the rights granted under the authority of any government in accordance with its patent laws.

SEQUENCE LISTING

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(i) APPLICANT: Lemischka, Thor R.

(ii) TITLE OF INVENTION: TOTIPOTENT HEMATOPOIETIC STEM CELL RECEPTORS AND THEIR LIGANDS

(iii) NUMBER OF SEQUENCES: 11

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- (F) ZIP: 10014

(v) COMPUTER READABLE FORM:

- (A) MEDIUM TYPE: Floppy disk
- (B) COMPUTER: IBM PC compatible
- (C) OPERATING SYSTEM: PC-DOS/MS-DOS
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(vii) PRIOR APPLICATION DATA:

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(B) FILING DATE: 15-JAN-1993

(vii) PRIOR APPLICATION DATA:

(A) APPLICATION NUMBER: US 08/045,272
(B) FILING DATE: 01-APR-1993

(vii) PRIOR APPLICATION DATA:

(A) APPLICATION NUMBER: US 08/076022
(B) FILING DATE: 09-JUN-1993

(vii) PRIOR APPLICATION DATA:

(A) APPLICATION NUMBER: US 08/080244
(B) FILING DATE: 18-JUN-1993

(vii) PRIOR APPLICATION DATA:
(A) APPLICATION NUMBER: US 08/081508
(B) FILING DATE: 21-JUN-1993

(vii) PRIOR APPLICATION DATA:
(A) APPLICATION NUMBER: US 08/096759
(B) FILING DATE: 22-JUL-1993

(vii) PRIOR APPLICATION DATA:
(A) APPLICATION NUMBER: US 08/125669
(B) FILING DATE: 23-SEP-1993

(viii) ATTORNEY/AGENT INFORMATION:
(A) NAME: Feit, Irving N.
(B) REGISTRATION NUMBER: 28,601
(C) REFERENCE/DOCKET NUMBER: LEM-3-15P

(ix) TELECOMMUNICATION INFORMATION:
(A) TELEPHONE: 212-645-1405
(B) TELEFAX: 212-645-2054

(2) INFORMATION FOR SEQ ID NO:1:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 3453 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: double
(D) TOPOLOGY: linear

(i.i) MOLECULE TYPE: cDNA

(i.ii) HYPOTHETICAL: NO

(i.iii) ANTI-SENSE: NO

(i.v) FRAGMENT TYPE: N-terminal

(ix) FEATURE:
 (A) NAME/KEY: mat_peptide
 (B) LOCATION: 112...3006

(ix) FEATURE:
 (A) NAME/KEY: sig_peptide
 (B) LOCATION: 31...111

(ix) FEATURE:
 (A) NAME/KEY: CDS
 (B) LOCATION: 31...3009

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:1:

54
 GCGGCCCTGGC TACCGGGCGC TCCGGAGGCC ATG CGG GCG TTG GCG CAG CGC AGC
 Met Arg Ala Leu Ala Gln Arg Ser
 -27 -25 -20
 GAC CGG CGG CTG CTG CTT GTT TTG TCA GTA ATG ATT CTT GAG
 ASP Arg Arg Leu Leu Val Val Leu Ser Val Met Ile Leu Glu
 -15 -10 -5
 102
 ACC GTT ACA AAC CAA GAC CCT GTG ATC AAG TGT GTT TTA ATC AGT
 Thr Val Thr Asn Gln Asp Leu Pro Val Ile Lys Cys Val Leu Ile Ser
 1 5 10
 150
 CAT GAG AAC AAT GGC TCA TCA GCG GGA AAG CCA TCA TCG TAC CGA ATG
 His Glu Asn Asn Gly Ser Ser Ala Gly Lys Pro Ser Ser Tyr Arg Met
 20 25
 198
 246
 GTG CGA GGA TCC CCA GAA GAC CTC CAG TGT ACC CCG AGG CGC CAG AGT
 Val Arg Gly Ser Pro Glu Asp Leu Gln Cys Thr Pro Arg Arg Gln Ser
 30 35 40 45
 294
 GAA GGG ACG GTA TAT GAA GCG ACC GTG GAG GTG GCC GAG TCT GGG

Glu	Gly	Thr	Val	Tyr	Glu	Ala	Ala	Thr	Val	Glu	Val	Ala	Glu	Ser	Gly
50															
TCC	ATC	ACC	CTG	CAA	GTG	CAG	CTC	GCC	ACC	CCA	GGG	GAC	CTT	TCC	TGC
Ser	Ile	Thr	Leu	Gln	Val	Gln	Leu	Ala	Thr	Pro	Gly	Asp	Leu	Ser	Cys
65															
CTC	TGG	GTC	TTT	AAG	CAC	AGC	TCC	CTG	GGC	TGC	CAG	CCG	CAC	TTT	GAT
Leu	Trp	Val	Phe	Lys	His	Ser	Ser	Leu	Gly	Cys	Gln	Pro	His	Phe	Asp
80															
TTA	CAA	AAC	AGA	GGA	ATC	GTT	TCC	ATG	GCC	ATC	TTG	AAC	GTG	ACA	GAG
Leu	Gln	Asn	Arg	Gly	Ile	Val	Ser	Met	Ala	Ile	Leu	Asn	Val	Thr	Gl
95															
ACC	CAG	GCA	GGA	GAA	TAC	CTA	CTC	CAT	ATT	CAG	AGC	GAA	CGC	GCC	AAC
Thr	Gln	Ala	Gly	Glu	Tyr	Leu	Leu	His	Ile	Gln	Ser	Glu	Arg	Ala	Asn
110															
TAC	ACA	GTA	CTG	TTC	ACA	GTG	AAT	GTA	AGA	GAT	ACA	CAG	CTG	TAT	GTG
Tyr	Thr	Val	Leu	Phe	Thr	Val	Asn	Val	Arg	Asp	Thr	Gln	Leu	Tyr	Val
125															
CTA	AGG	AGA	CCT	TAC	TTT	AGG	AAG	ATG	GAA	AAC	CAG	GAT	GCA	CTG	CTC
Leu	Arg	Arg	Pro	Tyr	Phe	Arg	Lys	Met	Glu	Asn	Gln	Asp	Ala	Leu	Leu
140															
TGC	ATC	TCC	GAG	GGT	GTT	CCG	GAG	CCC	ACT	GTG	GAG	TGG	GTG	CTC	TGC
Cys	Ile	Ser	Glu	Gly	Val	Pro	Glu	Pro	Thr	Val	Glu	Trp	Val	Leu	Cys
155															
AGC	TCC	CAC	AGG	GAA	AGC	TGT	AAA	GAA	GAA	GGC	CCT	GCT	GTT	GTC	AGA
Ser	Ser	His	Arg	Glu	Ser	Cys	Lys	Glu	Glu	Gly	Pro	Ala	Val	Val	Arg
170															
AAG	GAG	GAA	AAG	GTA	CTT	CAT	GAG	TTC	GGA	ACA	GAC	ATC	AGA	TGC	
Lys	Glu	Glu	Lys	Val	Ile	His	Glu	Leu	Phe	Gly	Thr	Asp	Ile	Arg	Cys
185															

190	195	200	205
TGT GCT AGA AAT GCA	CTG GGC CGC GAA	ACC AAG CTC	ATA
Cys Ala Arg Asn Ala	Leu Gly Arg Glu	Leu Phe Thr	Ile
210	Cys 215	220	774
GAT CTA AAC CAG GCT CCT CAG AGC	ACA CTG CCC CAG	TTA TTC CTG	AAA
Asp Leu Asn Gln Ala	Pro Gln Ser Thr Leu Pro	Gln Leu Phe	Leu Lys
225	230	235	822
GTC GGG GAA CCC TTG TGG ATC AGG	TGT AAG GCC ATC	CAT GTG AAC	CAT
Val Gly Glu Pro Leu Trp Ile Arg	Cys Lys Ala Ile	His Val Asn	His
240	245	250	870
GGA TTC GGG CTC ACC TGG GAG	CTG GAA GAC AAA	GCC CTG GAG	GAG GGC
Gly Phe Gly Leu Thr Trp Glu	Leu Glu Asp Lys	Ala Leu Glu	Glu Gly
255	260	265	918
AGC TAC TTT GAG ATG AGT ACC	TAC TCC ACA AAC	AGG ACC ATG ATT	CGG
Ser Tyr Phe Glu Met Ser Thr	Tyr Ser Thr Asn	Arg Thr Met Ile	Arg
270	275	280	966
ATT CTC TTG GCC TTT GTG TCT	TCC GTG GGA AGG	AAC GAC ACC	TAT CGG
Ile Leu Leu Ala Phe Val	Ser Ser Val	Gly Arg Asn Asp	Thr Gly Tyr
290	295	300	1014
TAC ACC TGC TCT TCC TCA AAG	CCC AGC CAG TCA	GCG TTG GTG	ACC
Tyr Thr Cys Ser Ser Lys His	Pro Ser Gln Ser	Ala Leu Val	Thr
305	310	315	1062
ATC CTA GAA AAA GGG TTT ATA AAC	GCT ACC AGC TCG	CAA GAA GAG	TAT
Ile Leu Glu Lys Gly Phe Ile	Asn Ala Thr Ser	Gln Glu Glu	Tyr
320	325	330	1110
GAA ATT GAC CCG TAC GAA AAG	TTC TGC TCA GTC	AGG TTT AAA	GCG
Glu Ile Asp Pro Tyr Glu Lys Phe	Cys Phe Ser Val	Arg Phe Lys Ala	Ala
335	340	345	1158

TAC CCA CGA ATC CGA TGC ACG TGG ATC TTC TCT CAA GCC TCA TTT CCT
 TYR Pro Arg Ile Arg Cys Thr Trp Ile Phe Ser Gln Ala Ser Phe Pro 1206
 350 355 360 365

TGT GAA CAG AGA GGC CTG GAG GAT GGG TAC AGC ATA TCT AAA TTT TGC
 Cys Glu Gln Arg Gly Leu Glu Asp Gly Tyr Ser Ile Ser Lys Phe Cys 1254
 370 375 380

GAT CAT AAC AAG CCA GGA GAG TAC ATA TTC TAT GCA GAA AAT GAT
 Asp His Lys Asn Lys Pro Gly Glu Tyr Ile Phe Tyr Ala Glu Asn Asp 1302
 385 390 395

GAC GCC CAG TTC ACC AAA ATG TTC ACG CTG AAT ATA AGA AAG AAA CCT
 Asp Ala Gln Phe Thr Lys Met Phe Thr Leu Asn Ile Arg Lys Lys Pro 1350
 400 405 410

CAA GTG CTA GCA AAT GCC TCA GCC AGC CAG GCG TCC TGT TCC TCT GAT
 Gln Val Leu Ala Asn Ala Ser Ala Ser Gln Ala Ser Cys Ser Ser Asp 1398
 415 420 425

GGC TAC CCG CTA CCC TCT TGG ACC TGG AAG AAG TGT TCG GAC AAA TCT
 Gly Tyr Pro Leu Pro Ser Trp Thr Trp Lys Lys Cys Ser Asp Lys Ser 1446
 430 435 440 445

CCC AAT TGC ACG GAG GAA ATC CCA GAA GGA GTT TGG AAT AAA MAG GCT
 Pro Asn Cys Thr Glu Glu Ile Pro Glu Gly Val Trp Asn Lys Lys Ala 1494
 450 455 460

AAC AGA AAA GTG TTT GGC CAG TGG TCG AGC AGT ACT CTA AAT ATG
 Asn Arg Lys Val Phe Gly Gln Trp Val Ser Ser Thr Leu Asn Met 1542
 465 470 475

ACT GAG GCC GGG AAA GGG CTT CTG GTC AAA TGC TGT GCG TAC AAT TCT
 Ser Glu Ala Gly Lys Gly Leu Leu Val Lys Cys Cys Ala Tyr Asn Ser 1590
 480 485 490

ATG GGC ACG TCT TGC GAA ACC ATC TTT TTA AAC TCA CCA GGC CCC TTC 1638

Met	Gly	Thr	Ser	Cys	Glu	Thr	Ile	Phe	Leu	Asn	Ser	Pro	Gly	Pro	Phe
495															
CCT	TTC	ATC	CAA	GAC	AAC	ATC	TCC	TTC	TAT	GCG	ACC	ATT	GGG	CTC	TGT
Pro	Phe	Ile	Gln	Asp	Asn	Ile	Ser	Phe	Tyr	Ala	Thr	Ile	Gly	Leu	Cys
510															525
CTC	CCC	TTC	ATT	GTT	CTT	CTC	ATT	GTT	TTG	ATC	TGC	CAC	AAA	TAC	AAA
Leu	Pro	Phe	Ile	val	val	Leu	Ile	val	Leu	Ile	Cys	His	Lys	TYR	Lys
															535
															540
AAG	CAA	TTT	AGG	TAC	GAG	AGT	CAG	CTG	CAG	ATC	CAG	GTG	ACT	GGC	
Lys	Gln	Phe	Arg	Arg	Tyr	Glu	Ser	Gln	Leu	Gln	Met	Ile	Gln	Val	Thr
															545
															550
CCC	CTG	GAT	AAC	GAG	TAC	TTC	TAC	GTG	GAC	TTC	AGG	GAC	TAT	GAA	TAT
Pro	Leu	Asp	Asn	Glu	Glu	Tyr	Phe	Tyr	Val	Asp	Phe	Arg	Asp	Tyr	Glu
															560
															565
GAC	CTT	AAG	TGG	GAG	TTG	CCG	AGA	GAG	AAC	TTA	GAG	TTT	GGG	AAG	GTC
Asp	Leu	Lys	Trp	Glu	Glu	Phe	Pro	Arg	Glu	Asn	Leu	Glu	Phe	Gly	Lys
															575
															580
CTG	GGG	TCT	GGC	GCT	TTG	GGG	AGG	GTG	ATG	AAC	GCC	ACG	GCC	TAT	GGC
Leu	Gly	Ser	Gly	Ala	Phe	Gly	Arg	Val	Met	Asn	Ala	Thr	Ala	Tyr	Gly
															590
															595
ATT	AGT	AAA	ACG	GGA	GTC	TCA	ATT	CAG	GTG	GCG	GTG	AAG	ATG	CTA	AAA
Ile	Ser	Lys	Thr	Gly	Val	Ser	Ile	Gln	Val	Ala	Val	Lys	Met	Leu	Lys
															610
															615
GAG	AAA	GCT	GAC	TGT	GAA	AAA	GAA	GCT	CTC	ATG	TCG	GAG	CTC	AAA	
Glu	Lys	Ala	Asp	Ser	Cys	Glu	Lys	Glu	Ala	Leu	Met	Ser	Glu	Leu	Lys
															625
															630
ATG	ATG	ACC	CAC	CTG	GGA	CAC	CAT	GAC	AAC	ATC	GTG	AAT	CTG	CTG	GGG
Met	Met	Thr	His	Leu	Gly	His	His	Asp	Asn	Ile	val	Asn	Ile	Leu	Gly
															635
															640

640	645	650
GCA TGC ACA CTG TCA GGG CCA GTG ATT TTG ATT TTT GAA TAT TGT TGC Ala Cys Thr Leu Ser Gly Pro Val Tyr Leu Ile Phe Glu Tyr Cys Cys		2118
TAT GGT GAC CTC CTC AAC TAC CTA AGA AGT AAA AGA GAG AAG TTT CAC Tyr Gly Asp Leu Leu Asn Tyr Leu Arg Ser Lys Arg Glu Lys Phe His	670	665
		2166
AGG ACA TGG ACA GAG ATT TTT AAG GAA CAT AAT TTC AGT TCT TAC CCT Arg Thr Trp Thr Glu Ile Phe Lys Glu His Asn Phe Ser Ser Tyr Pro	690	695
		2214
ACT TTC CAG GCA CAT TCA AAT TCC AGC ATG CCT GGT TCA CGA GAA GTT Thr Phe Gln Ala His Ser Asn Ser Met Pro Gly Ser Arg Glu Val	705	710
		2262
GAG TTA CAC CCG CCC TTG GAT CAG CTC TCA GGG TTC AAT GGG AAT TCA Gln Leu His Pro Pro Leu Asp Gln Leu Ser Gly Phe Asn Gly Asn Ser	720	725
		2310
ATT CAT TCT GAA GAT GAG ATT GAA TAT GAA AAC CAG AAG AGG CTG GCA Ile His Ser Glu Asp Glu Ile Glu Tyr Glu Asn Gln Lys Arg Leu Ala	735	740
		2358
GAA GAA GAG GAA GAT TTG AAC GTG CTG ACG TTT GAA GAC CTC CTT Glu Glu Glu Glu Asp Leu Asn Val Leu Thr Phe Glu Asp Leu Leu	750	755
		2406
TGC TTT GCG TAC CAA GTG GCC AAA GGC ATG GAA TTC CTG GAG TTC AAG Cys Phe Ala Tyr Gln Val Ala Lys Gly Met Glu Phe Leu Glu Phe Lys	770	775
		2454
TCG TGT GTC CAC AGA GAC CTG GCA GCC AGG AAT GTG TTG GTC ACC CAC Ser Cys Val His Arg Asp Leu Ala Arg Asn Val Leu Val Thr His	785	790
		2502

GGG	AAG	GTG	AAG	ATC	TGT	GAC	TTT	GGG	CTG	GCC	CGA	GAC	ATC	CTG	2550	
Gly	Lys	Val	Val	Lys	Ile	Cys	Asp	Phe	Gly	Leu	Ala	Arg	Asp	Ile	Leu	
800				805						810						
AGC	GAC	TCC	AGC	TAC	GTC	GGC	AGG	GGC	AAC	GCA	CGG	CTG	CCG	GTG	2598	
Ser	Asp	Ser	Ser	Tyr	Val	Val	Arg	Gly	Asn	Ala	Arg	Leu	Pro	Val	Lys	
815				820						825						
TGG	ATG	GCA	CCC	GAG	AGC	TTA	TTT	GAA	GGG	ATC	TAC	ACA	ATC	AAG	AGT	2646
Trp	Met	Ala	Pro	Glu	Ser	Leu	Phe	Glu	Gly	Ile	Tyr	Thr	Ile	Lys	Ser	
830				835						840					845	
GAC	GTC	TGG	TCC	TAC	GGC	ATC	CTT	CTC	TGG	GAG	ATA	TTT	TCA	CTG	GGT	2694
Asp	Val	Trp	Ser	Tyr	Gly	Ile	Leu	Leu	Trp	Glu	Ile	Phe	Ser	Leu	Gly	
				850					855						860	
GTC	AAC	CCT	TAC	CCT	GGC	ATT	CCT	GTC	GAC	GCT	AAC	TTC	TAT	AAA	CTG	2742
Val	Asn	Pro	Tyr	Pro	Gly	Ile	Pro	Vail	Asp	Ala	Asn	Phe	Tyr	Lys	Leu	
				865					870						875	
ATT	CAG	AGT	GGA	TTT	AAA	ATG	GAG	CAG	CCA	TTC	TAT	GCC	ACA	GAA	GGG	2790
Ile	Gln	Ser	Gly	Phe	Lys	Met	Glu	Gln	Pro	Phe	Tyr	Ala	Thr	Glu	Gly	
				880					885						890	
ATA	TAC	TTT	GTA	ATG	CAA	TCC	TGG	GCT	TTT	GAC	TCA	AGG	AAG	CGG	2838	
Ile	Tyr	Phe	Val	Met	Gln	Ser	Cys	Trp	Ala	Phe	Asp	Ser	Arg	Lys	Arg	
				895					900						905	
CCA	TCC	TTC	CCC	AAC	CTG	ACT	TCA	TTT	TTA	GGA	TGT	CAG	CTG	GCA	GAG	2886
Pro	Ser	Phe	Pro	Asn	Leu	Thr	Ser	Phe	Leu	Gly	Cys	Gln	Leu	Ala	Glu	
				910					915						925	
GCA	GAA	GAA	GCA	TGT	ATC	AGA	ACA	TCC	ATC	CAT	CTA	CCA	AAA	CAG	GCG	2934
Ala	Glu	Glu	Ala	Cys	Ile	Arg	Thr	Ser	Ile	His	Leu	Pro	Lys	Gln	Ala	
				930					935						940	
GCC	CCT	CAG	CAG	AGA	GGC	GGG	CTC	AGA	GGC	CAG	TCG	CCA	CAG	CGC	CAG	2982

Ala Pro Gln Gln Arg Gly Leu Arg Ala Gln Ser Pro Gln Arg Gln
 945 950 955
 GTG AAG ATT CAC AGA GAA AGA ACT TAGCGAGGAG GCCTTGGACC CGGCCACCC
 Val Lys Ile His Arg Ser Arg Ser
 960 965
 AGCAGGGCTGT AGACCCGAGA GCCAAGATTA GCCTCGCCCTC TGAGGAAGGCG CCCTACAGCG
 CGTTGCTTCG CTGGACTTTT CTCTAGATGC TGCTCTGCCAT TACTCCAAG TGACTTCTAT
 AAAATCAAC CTCTCCTCGC ACAGGGGGA GAGCCAATAA TGAGACTTGT TGTTGAGGCC
 GCCTACCTG GGGCCTTTC CACCGAGCTTG AGGGAAAGC CATGTATCTG AAATATAGTA
 TATTCTTGTAA AATACGTGAA ACAAAACAAA CCCGTTTTT GCTAAGGGAA AGCTAAATAT
 GATTTTAAA AATCTATGTT TTAAAATACT ATGTAACCTT TTCATCTAT TAGTGATATA
 TTTTATGGAT GGAAATAAAC TTTCTACTGT AAAAAGAAA AAAAAGAAA AAAAAGAAA
 3453

(2) INFORMATION FOR SEQ ID NO: 2:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 992 amino acids
- (B) TYPE: amino acid
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 2:

Met Arg Ala Leu Ala Gln Arg Ser Asp Arg Arg Leu Leu Leu Val
 -27 -25 -20 -15

Val Leu Ser Val Met Ile Leu Glu Thr Val Thr Asn Gln Asp Leu Pro

66

val	Ile	Lys	Cys	val	Leu	Ile	Ser	His	Glu	Asn	Asn	Gly	Ser	Ser	Ala
				10				15							20
Gly	Lys	Pro	Ser	Ser	Tyr	Arg	Met	Val	Arg	Gly	Ser	Pro	Glu	Asp	Leu
				25			30		35						
Gln	Cys	Thr	Pro	Arg	Arg	Gln	Ser	Glu	Gly	Thr	Val	Tyr	Glu	Ala	Ala
				40			45		50						
Thr	Val	Glu	Val	Ala	Glu	Ser	Gly	Ser	Ile	Thr	Leu	Gln	Val	Gln	Leu
				55			60		65						
Ala	Thr	Pro	Gly	Asp	Leu	Ser	Cys	Leu	Trp	Val	Phe	Lys	His	Ser	Ser
				70			75		80			85			
Leu	Gly	Cys	Gln	Pro	His	Phe	Asp	Leu	Gln	Asn	Arg	Gly	Ile	Val	Ser
							90		95				100		
Met	Ala	Ile	Leu	Asn	Val	Thr	Glu	Thr	Gln	Ala	Gly	Glu	Tyr	Leu	Leu
							105		110			115			
His	Ile	Gln	Ser	Glu	Arg	Ala	Asn	Tyr	Thr	Val	Leu	Phe	Thr	Val	Asn
							120		125			130			
Val	Arg	Asp	Thr	Gln	Leu	Tyr	Val	Leu	Arg	Arg	Pro	Tyr	Phe	Arg	Lys
							135		140			145			
Met	Glu	Asn	Gln	Asp	Ala	Leu	Cys	Ile	Ser	Glu	Gly	Val	Pro	Glu	165
							150		155			160			
Pro	Thr	Val	Glu	Trp	Val	Leu	Cys	Ser	Ser	His	Arg	Glu	Ser	Cys	Lys
							170		175			180			
Glu	Glu	Pro	Ala	Val	Val	Arg	Lys	Glu	Glu	Lys	Val	Ileu	His	Glu	
				185				190				195			

Leu Phe GLY Thr Asp Ile Arg Cys Cys Ala Arg Asn Ala Leu Gly Arg
 200 205 210
 Glu Cys Thr Lys Leu Phe Thr Ile Asp Leu Asn Gln Ala Pro Gln Ser
 215 220 225
 Thr Leu Pro Gln Leu Phe Leu Lys Val GLY Glu Pro Leu Trp Ile Arg
 230 235 240 245
 Cys Lys Ala Ile His Val Asn His GLY Phe GLY Leu Thr Trp Glu Leu
 250 255 260
 Glu Asp Lys Ala Leu Glu Gly Ser Tyr Phe Glu Met Ser Thr Tyr
 265 270 275
 Ser Thr Asn Arg Thr Met Ile Arg Ile Leu Leu Ala Phe Val Ser Ser
 280 285 290
 Val GLY Arg Asn Asp Thr GLY Tyr Tyr Thr Cys Ser Ser Lys His
 295 300 305
 Pro Ser Gln Ser Ala Leu Val Thr Ile Leu Glu Lys Gly Phe Ile Asn
 310 315 320 325
 Ala Thr Ser Ser Gln Glu Glu Tyr Glu Ile Asp Pro Tyr Glu Lys Phe
 330 335 340 345
 Cys Phe Ser Val Arg Phe Lys Ala Tyr Pro Arg Ile Arg Cys Thr Trp
 350 355
 Ile Phe Ser Gln Ala Ser Phe Pro Cys Glu Gln Arg GLY Leu Glu Asp
 360 365 370
 GLY Tyr Ser Ile Ser Lys Phe Cys Asp His Lys Asn Lys Pro GLY Glu
 375 380 385
 Tyr Ile Phe Tyr Ala Glu Asn Asp Asp Ala Gln Phe Thr Lys Met Phe

390	Thr Leu Asn Ile Arg Lys Lys Pro Gln Val Leu Ala Asn Ala Ser Ala	410	400
395	Ser Gln Ala Ser Cys Ser Ser Asp Gly Tyr Pro Leu Pro Ser Trp Thr	415	400
400	Ser Gln Ala Ser Cys Ser Ser Asp Gly Tyr Pro Leu Pro Ser Trp Thr	420	395
405	Trp Lys Lys Cys Ser Asp Lys Ser Pro Asn Cys Thr Glu Glu Ile Pro	425	400
410	Glu Gly Val Trp Asn Lys Lys Ala Asn Arg Lys Val Phe Gly Gln Trp	430	405
415	Glu Gly Val Trp Asn Lys Lys Ala Asn Arg Lys Val Phe Gly Gln Trp	435	410
420	Val Ser Ser Ser Thr Leu Asn Met Ser Glu Ala Gly Lys Gly Leu Leu	440	415
425	Val Lys Cys Cys Ala Tyr Asn Ser Met Gly Thr Ser Cys Glu Thr Ile	445	420
430	Val Lys Cys Cys Ala Tyr Asn Ser Met Gly Thr Ser Cys Glu Thr Ile	460	425
435	Val Lys Cys Cys Ala Tyr Asn Ser Met Gly Thr Ser Cys Glu Thr Ile	465	430
440	Val Lys Cys Cys Ala Tyr Asn Ser Met Gly Thr Ser Cys Glu Thr Ile	480	435
445	Val Lys Cys Cys Ala Tyr Asn Ser Met Gly Thr Ser Cys Glu Thr Ile	485	440
450	Val Lys Cys Cys Ala Tyr Asn Ser Met Gly Thr Ser Cys Glu Thr Ile	495	445
455	Val Lys Cys Cys Ala Tyr Asn Ser Met Gly Thr Ser Cys Glu Thr Ile	500	450
460	Val Lys Cys Cys Ala Tyr Asn Ser Met Gly Thr Ser Cys Glu Thr Ile	505	455
465	Val Lys Cys Cys Ala Tyr Asn Ser Met Gly Thr Ser Cys Glu Thr Ile	510	460
470	Val Lys Cys Cys Ala Tyr Asn Ser Met Gly Thr Ser Cys Glu Thr Ile	515	465
475	Val Lys Cys Cys Ala Tyr Asn Ser Met Gly Thr Ser Cys Glu Thr Ile	520	470
480	Val Lys Cys Cys Ala Tyr Asn Ser Met Gly Thr Ser Cys Glu Thr Ile	525	475
485	Val Lys Cys Cys Ala Tyr Asn Ser Met Gly Thr Ser Cys Glu Thr Ile	530	480
490	Val Lys Cys Cys Ala Tyr Asn Ser Met Gly Thr Ser Cys Glu Thr Ile	535	485
495	Val Lys Cys Cys Ala Tyr Asn Ser Met Gly Thr Ser Cys Glu Thr Ile	540	490
500	Val Lys Cys Cys Ala Tyr Asn Ser Met Gly Thr Ser Cys Glu Thr Ile	545	495
505	Val Lys Cys Cys Ala Tyr Asn Ser Met Gly Thr Ser Cys Glu Thr Ile	550	500
510	Val Lys Cys Cys Ala Tyr Asn Ser Met Gly Thr Ser Cys Glu Thr Ile	555	505
515	Val Lys Cys Cys Ala Tyr Asn Ser Met Gly Thr Ser Cys Glu Thr Ile	560	510
520	Val Lys Cys Cys Ala Tyr Asn Ser Met Gly Thr Ser Cys Glu Thr Ile	565	515
525	Val Lys Cys Cys Ala Tyr Asn Ser Met Gly Thr Ser Cys Glu Thr Ile	570	520
530	Val Lys Cys Cys Ala Tyr Asn Ser Met Gly Thr Ser Cys Glu Thr Ile	575	525
535	Val Lys Cys Cys Ala Tyr Asn Ser Met Gly Thr Ser Cys Glu Thr Ile	580	530
540	Val Lys Cys Cys Ala Tyr Asn Ser Met Gly Thr Ser Cys Glu Thr Ile	585	535
545	Val Lys Cys Cys Ala Tyr Asn Ser Met Gly Thr Ser Cys Glu Thr Ile	590	540
550	Val Lys Cys Cys Ala Tyr Asn Ser Met Gly Thr Ser Cys Glu Thr Ile	595	545

600 Val Met Asn Ala Thr Ala Tyr Gly Ile Ser Lys Thr Gly Val Ser Ile
605
610
615 Gln Val Ala Val Lys Met Leu Lys Glu Lys Ala Asp Ser Cys Glu Lys
620
625
630 Glu Ala Leu Met Ser Glu Leu Lys Met Met Thr His Leu Gly His His
635
640
645 Asp Asn Ile Val Asn Leu Leu Gly Ala Cys Thr Leu Ser Gly Pro Val
650
655
660 Tyr Leu Ile Phe Glu Tyr Cys Cys Tyr Gly Asp Leu Leu Asn Tyr Leu
665
670
675 Arg Ser Lys Arg Glu Lys Phe His Arg Thr Trp Thr Glu Ile Phe Lys
680
685
690
695 Glu His Asn Phe Ser Ser Tyr Pro Thr Phe Gln Ala His Ser Asn Ser
700
705
710 Ser Met Pro Gly Ser Arg Glu Val Gln Leu His Pro Pro Leu Asp Gln
715
720
725 Leu Ser Gly Phe Asn Gly Asn Ser Ile His Ser Glu Asp Glu Ile Glu
730
735
740
745 Tyr Glu Asn Gln Lys Arg Leu Ala Glu Glu Glu Glu Asp Leu Asn
750
755
760 Val Leu Thr Phe Glu Asp Leu Leu Cys Phe Ala Tyr Gln Val Ala Lys
765
770
775 Gly Met Glu Phe Leu Glu Phe Lys Ser Cys Val His Arg Asp Leu Ala
780
785
790 Ala Arg Asn Val Leu Val Thr His Gly Lys Val Val Lys Ile Cys Asp

790	795	800	805
Phe Gly Leu Ala Arg Asp Ile Leu Ser Asp 810	815	Ser Ser Tyr Val Val Arg 820	
Gly Asn Ala Arg Leu Pro Val Lys Trp 825	830	Met Ala Pro Glu Ser Leu Phe 835	
Glu Gly Ile Tyr Thr Ile Lys Ser Asp Val Trp Ser Tyr 840	845	Gly Ile Leu 850	
Leu Trp Glu Ile Phe Ser Leu Gly Val Asn Pro Tyr 855	860	Pro Gly Ile Pro 865	
Val Asp Ala Asn Phe Tyr Lys Leu Ile Gln Ser 870	875	Gly Phe Lys Met Glu 880	885
Gln Pro Phe Tyr Ala Thr Glu Gly Ile Tyr 890	895	Val Met Gln Ser Cys 900	
Trp Ala Phe Asp Ser Arg Lys Arg Pro Ser Phe 905	910	Pro Asn Leu Thr Ser 915	
Phe Leu Gly Cys Gln Leu Ala Glu Ala Cys 920	925	Ile Arg Thr 930	
Ser Ile His Leu Pro Lys Gln Ala Ala Pro Gln 935	940	Gln Arg Gly Gly Leu 945	
Arg Ala Gln Ser Pro Gln Arg Gln Val Lys Ile His 950	955	Arg Glu Arg Ser 960	965

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 3501 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: double
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(iii) HYPOTHETICAL: NO

(iv) ANTI-SENSE: NO

(v) FRAGMENT TYPE: N-terminal

(ix) FEATURE:

(A) NAME/KEY: CDS
(B) LOCATION: 58..3039

(ix) FEATURE:

(A) NAME/KEY: mat_peptide
(B) LOCATION: 139..3036

(ix) FEATURE:

(A) NAME/KEY: sig_peptide
(B) LOCATION: 58..138

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 3:

CGAGGGGCA TCCGAGGGCT GGGCCGGGG CCTGGGGGAC CCCGGCTCC GGAGGCC
57
ATG CCG GCG TTG GCG CGC GAC GCG ACC GTG CCG CTC GTT GTT
105
Met Pro Ala Leu Ala Arg Asp Ala Gly Thr Val Pro Leu Val Val
-27 -25 -20 -15

TCT TCT GCA ATG ATA TTT GGG ACT ATT ACA AAT CAA GAT CTG CCT GTG

153

135	140	145	
GAA AAC CAG GAC GCC CTC TGC GTC ATA TCT GAG AGC GTT CCA GAG CCG Glu Asn Gln Asp Ala Leu Val Cys Ile Ser Glu Ser Val Pro Glu Pro 150 155 160 165			633
ATC GTG GAA TGG GTG CTT TGC GAT TCA CAG GGG GAA AGC TGT AAA GAA Ile Val Glu Trp Val Leu Cys Asp Ser Gln Gly Glu Ser Cys Lys Glu 170 175 180 185			681
GAA ACT CCA GCT GTT AAA AAG GAG GAA AAA GTG CTT CAT GAA TTA Glu Ser Pro Ala Val Val Lys Lys Glu Lys Val Leu His Glu Leu 185 190 195			729
TTT GGG ACG GAC ATA AGG TGC TGT GCC AGA AAT GAA CTG GGC AGG GAA Phe Gly Thr Asp Ile Arg Cys Cys Ala Arg Asn Glu Leu Gly Arg Glu 200 205 210			777
TGC ACC AGG CTG TTC ACA ATA GAT CTA AAT CAA ACT CCT CAG ACC ACA Cys Thr Arg Leu Phe Thr Ile Asp Leu Asn Gln Thr Pro Gln Thr Thr 215 220 225			825
TTG CCA CAA TTA TTT CTT AAA GTA GGG GAA CCC TTA TGG ATA AGG TGC Leu Pro Gln Leu Phe Leu Lys Val Gly Glu Pro Leu Trp Ile Arg Cys 230 235 240 245			873
AAA GCT GTT CAT GTG AAC CAT GGA TTC GGG CTC ACC TGG GAA TTA GAA Lys Ala Val His Val Asn His Gly Phe Gly Leu Thr Trp Glu Leu Glu 250 255 260			921
AAC AAA GCA CTC GAG GAG GGC AAC TAC TTT GAG ATG AGT ACC TAT TCA Asn Lys Ala Leu Glu Glu Gly Asn Tyr Phe Glu Met Ser Thr Tyr Ser 265 270 275			969
ACA AAC AGA ACT ATG ATA CGG ATT CTG TTT GCT GTA TCA TCA GTG Thr Asn Arg Thr Met Ile Arg Ile Leu Phe Ala Phe Val Ser Ser Val 280 285 290			1017

GCA	AGA	AAC	GAC	ACC	GGG	TAC	TAC	ACT	TGT	TCC	TCT	TCA	AAG	CAT	CCC	1065
Ala	Arg	Asn	Asp	Thr	Gly	Tyr	Tyr	Thr	Cys	Ser	Ser	Ser	Ser	Lys	His	Pro
295					300					305						
AGT	CAA	TCA	GCT	TTG	GTT	ACC	ATC	GTA	GGG	AAG	GGA	TTT	ATA	AAT	GCT	1113
Ser	Gln	Ser	Ala	Leu	Val	Thr	Ile	Val	Gly	Lys	Gly	Phe	Ile	Asn	Ala	
310				315						320						325
ACC	AAT	TCA	AGT	GAA	GAT	TAT	GAA	ATT	GAC	CAA	TAT	GAA	GAG	TTT	TGT	1161
Thr	Asn	Ser	Ser	Glu	Asp	Tyr	Glu	Ile	Asp	Gln	Tyr	Glu	Glu	Phe	Cys	
				330					335							340
TTT	TCT	GTC	AGG	TTT	AAA	GCC	TAC	CCA	CAA	ATC	AGA	TGT	ACG	TGG	ACC	1209
Phe	Ser	Val	Arg	Phe	Lys	Ala	Tyr	Pro	Gln	Ile	Arg	Cys	Thr	Trp	Thr	
				345					350							355
TTC	TCT	CGA	AAA	TCA	TTT	CCT	TGT	GAG	CAA	AAG	GGT	CTT	GAT	AAC	GGA	1257
Phe	Ser	Arg	Lys	Ser	Phe	Pro	Cys	Glu	Gln	Lys	Gly	Leu	Asp	Asn	Gly	
				360					365							370
TAC	AGC	ATA	TCC	AAG	TTT	TGC	AAT	CAT	AAG	CAC	CAG	CCA	GAA	TAT		1305
Tyr	Ser	Ile	Ser	Lys	Phe	Cys	Asn	Asn	His	Lys	Gln	Pro	Gly	Glu	Tyr	
				375					380							385
ATA	TTC	CAT	GCA	GAA	AAT	GAT	GCC	CAA	TTT	ACC	AAA	ATG	TTC	ACG		1353
Ile	Phe	His	Ala	Glu	Asn	Asp	Asp	Ala	Gln	Phe	Thr	Lys	Met	Phe	Thr	
				390					395							400
CTG	AAT	ATA	AGA	AGG	AAA	CCT	CAA	GTC	GCA	GAA	GCA	TCG	GCA	AGT		1401
Leu	Asn	Ile	Arg	Arg	Lys	Pro	Gln	Val	Leu	Ala	Glu	Ala	Ser	Ala	Ser	
				410						415						420
CAG	GCG	TCC	TGT	TTC	TGG	GAT	GGG	TAC	CCA	TTA	CCA	TCT	TGG	ACC	TGG	1449
Gln	Ala	Ser	Cys	Phe	Ser	Asp	Gly	Tyr	Pro	Leu	Pro	Ser	Trp	Thr	Trp	
				425					430							435
AAG	AAG	TGT	TCA	GAC	AAG	TCT	CCC	AAC	TGC	ACA	GAA	GAG	ATC	ACA	GAA	1497

Lys	Lys	Cys	Ser	Asp	Lys	Ser	Pro	Asn	Cys	Thr	Glu	Glu	Ile	Thr	Glu
440															
GGG	GTC	TGG	AAT	AGA	AAG	GCT	AAC	AGA	AAA	GTC	TTT	GGG	CAG	TGG	GTG
Gly	Val	Trp	Asn	Arg	Lys	Ala	Asn	Arg	Lys	Val	Phe	Gly	Gln	Trp	Val
455															
460															
TCG	AGC	AGT	ACT	CTA	AAC	ATG	AGT	GAA	GCC	ATA	AAA	GGG	TTC	CTG	GTC
Ser	Ser	Ser	Thr	Leu	Asn	Met	Ser	Glu	Ala	Ile	Lys	Gly	Phe	Leu	Val
470															
475															
AAG	TGC	TGT	GCA	TAC	AAT	TCC	CTT	GGC	ACA	TCT	TGT	GAG	ACG	ATC	CTT
Lys	Cys	Cys	Ala	Tyr	Asn	Ser	Leu	Gly	Thr	Ser	Cys	Glu	Thr	Ile	Leu
490															
495															
TAA	AAC	TCT	CCA	GGC	CCC	TTC	CCT	ATC	CAA	GAC	AAC	ATC	TCA	TTC	
Leu	Asn	Ser	PRO	Gly	Pro	Phe	Pro	Ile	Gln	Asp	Asn	Ile	Ser	Phe	
505															
510															
TAT	GCA	ACA	ATT	GGT	GTT	TGT	CTC	CTC	TTC	ATT	GTC	GTT	TTA	ACC	CTG
Tyr	Ala	Thr	Ile	Gly	Val	Cys	Leu	Leu	Phe	Ile	Val	Val	Leu	Thr	Leu
520															
525															
CTA	ATT	TGT	CAC	AAG	TAC	AAA	AAG	CAA	TTT	AGG	TAT	GAA	AGC	CAG	CTA
Leu	Ile	Cys	His	Lys	Tyr	Lys	Lys	Gln	Phe	Arg	Tyr	Glu	Ser	Gln	Leu
535															
540															
CAG	ATG	GTA	CAG	GTG	ACC	GGC	TCC	TCA	GAT	AAT	GAG	TAC	TTC	TAC	GT
Gln	Met	Val	Gln	Val	Thr	Gly	Ser	Ser	Asp	Asn	Glu	Tyr	Phe	Tyr	Val
550															
555															
GAT	TTC	AGA	GAA	TAT	GAA	TAT	GAT	CTC	AAA	TGG	TTT	CCA	AGA	GAA	
Asp	Phe	Arg	Glu	Tyr	Glu	Tyr	Asp	Leu	Lys	Trp	Glu	Phe	Pro	Arg	Glu
570															
575															
AAT	TTA	GAG	TTT	GGG	AAG	GTA	CTA	GGA	TCA	GGT	GCT	TTT	GGA	AAA	GTG
Asn	Leu	Glu	Phe	Gly	Lys	Val	Leu	Gly	Ser	Gly	Ala	Phe	Gly	Lys	Val
1929															

585	590	595
ATG AAC GCA ACA GCT TAT GGA ATT AGC AAA ACA GGA GTC TCA ATC CAG Met Asn Ala Thr Ala Tyr Gly Ile Ser Lys Thr Gly Val Ser Ile Gln		
600	605	610
GTG GCC GTC AAA ATG CTG AAA GAA AAA GCA GAC AGC TCT GAA AGA GAG Val Ala Val Lys Met Leu Lys Ala Asp Ser Ser Glu Arg Glu		
615	620	625
GCA CTC ATG TCA GAA CTC AAG ATG ACC CAG CTG GGA AGC CAC GAG Ala Leu Met Ser Glu Leu Lys Met Met Thr Gln Leu Gly Ser His Glu		
630	635	640
AAT ATT GTG AAC CTG CTG GGG GCG TGC ACA CTG TCA GGA CCA ATT TAC Asn Ile Val Asn Leu Leu Gly Ala Cys Thr Leu Ser Gly Pro Ile Tyr		
650	655	660
TTC ATT TTT GAA TAC TGT TGC TAT GGT GAT CTT CTC AAC TAT CTA AGA Leu Ile Phe Glu Tyr Cys Cys Tyr Gly Asp Leu Leu Asn Tyr Leu Arg		
665	670	675
AGT AAA AGA GAA AAA TTT CAC AGG ACT TGG ACA GAG ATT TTC AAG GAA Ser Lys Arg Glu Lys Phe His Arg Thr Trp Thr Glu Ile Phe Lys Glu		
680	685	690
CAC AAT TTC AGT TTT TAC CCC ACT TTC CAA TCA CAT CCA AAT TCC AGC His Asn Phe Ser Phe Tyr Pro Thr Phe Gln Ser His Pro Asn Ser Ser		
695	700	705
ATG CCT GGT TCA AGA GAA GTT CAG ATA CAC CCG GAC TCG GAT CAA ATC Met Pro Gly Ser Arg Glu Val Gln Ile His Pro Asp Ser Asp Gln Ile		
710	715	720
TCA GGG CTT CAT GGG AAT TCA TTT CAC TCT GAA GAT GAA ATT GAA TAT Ser Gly Leu His Gly Asn Ser Phe His Ser Glu Asp Glu Ile Glu Tyr		
730	735	740

GAA AAC CAA AAA AGG CTG GAA GAG GAC TTG AAT GTG CTT ACA	2409
Glu Asn Gln Lys Arg Leu Glu Glu Asp Leu Asn Val Leu Thr	
745 750 755	
TTT GAA GAT CTT CTC TGC TTT GCA TAT CAA GTT GCC AAA GGA ATG GAA	2457
Phe Glu Asp Leu Cys Phe Ala Tyr Gln Val Ala Lys Glu Met Glu	
760 765 770	
TTT CTG GAA TTT AAG TCG TGT GTT CAC AGA GAC CTG GCC AGG AAC	2505
Phe Leu Glu Phe Lys Ser Cys Val His Arg Asp Leu Ala Arg Asn	
775 780 785	
GTC CTT GTC ACC CAC GGG AAA GTG GTG AAG ATA TGT GAC TTT GGA TTG	2553
Val Leu Val Thr His Gly Lys Val Val Lys Ile Cys Asp Phe Gly Leu	
790 795 800 805	
GCT CGA GAT ATC ATG AGT GAT TCC AAC TAT GTT GTC AGG GGC AAT GCC	2601
Ala Arg Asp Ile Met Ser Asp Ser Asn Tyr Val Val Arg Gly Asn Ala	
810 815 820	
CGT CTG CCT GTA AAA TGG ATG GCC CCC GAA AGC CTG TTT GAA GGC ATC	2649
Arg Leu Pro Val Lys Trp Met Ala Pro Glu Ser Leu Phe Glu Gly Ile	
825 830 835 840	
TAC ACC ATT AAG AGT GAT GTC TGG TCA TAT GGA ATA TTA CTG TGG GAA	2697
Tyr Thr Ile Lys Ser Asp Val Trp Ser Tyr Gly Ile Leu Trp Glu	
840 845 850 855	
ATC TTC TCA CTT GGT GTG ATC CCT TAC CCT GGC ATT CCG GTT GAT GCT	2745
Ile Phe Ser Leu Gly Val Asn Pro Tyr Pro Gly Ile Pro Val Asp Ala	
855 860 865	
AAC TTC TAC AAA CTG ATT CAA AAT GGA TTT AAA ATG GAT CAG CCA TT	2793
Asn Phe Tyr Lys Leu Ile Gln Asn Gly Phe Lys Met Asp Gln Pro Phe	
870 875 880 885	
TAT GCT ACA GAA GAA ATA TAC ATT ATA ATG CAA TCC TGC TGG GCT TT	2841

Tyr	Ala	Thr	Glu	Ile	Glu	Ile	Tyr	Ile	Ile	Met	Gln	Ser	Cys	Trp	Ala	Phe
890																900
GAC	TCA	AGG	AAA	CGG	CCA	TCC	TTG	CCT	AAT	TTG	ACT	TCG	TTT	TTA	GGA	2889
Asp	Ser	Arg	Lys	Arg	Pro	Ser	Phe	Pro	Asn	Leu	Thr	Ser	Phe	Leu	Gly	
																905
TGT	CAG	CTG	GCA	GAT	GCA	GAA	GCG	ATG	TAT	CAG	AAT	GTG	GAT	GCG		2937
Cys	Gln	Leu	Ala	Asp	Ala	Glu	Glu	Ala	Met	Tyr	Gln	Asn	Val	Asp	Gly	
																920
CGT	GTT	TCG	GAA	TGT	CCT	CAC	ACC	TAC	CAA	AAC	AGG	CGA	CCT	TTC	AGC	2985
Arg	Val	Ser	Glu	Cys	Pro	His	Thr	Tyr	Gln	Asn	Arg	Arg	Arg	Pro	Phe	Ser
																925
AGA	GAG	ATG	GAT	TTG	GGG	CTA	CTC	TCT	CCG	CAG	GCT	CAG	GTC	GAA	GAT	3033
Arg	Glu	Met	Asp	Leu	Gly	Leu	Leu	Ser	Pro	Gln	Ala	Gln	Val	Glu	Asp	
																930
TCG	TAGAGGAACA	ATTAGCTTT	AAGGACTTCA	TCCCTCCACC	TATCCCTAAC											
Ser																3086
AGGCTGTAGA	TTACCAAAAC	AAGATTAAATT	TCATCACTAA	AAGAAAATCT	ATTATCAACT											3146
GCTGCTTCAC	CAGACTTTTC	TCTAGAAGGCC	GTCTGGGTT	ACTCTTGT	TT TCAAAGGGAC											3206
TTTTGTAAAA	TCAAAATCATC	CTGTCACAAG	GCAGGGAGGAG	CTGATAATGA	ACTTTATTGG											3266
AGCATTGATC	TGCATCCAAG	GCCTTCTCAG	GCCGGCTTGA	GTGAATTGTG	TACCTGAAGT											3326
ACAGTATTATT	CTTGTAAATA	CATAAAACAA	AAGCATTTTTG	CTAAGGAGAA	GCTAATAATGA											3386
TTTTTTAAGT	CTATGTTTA	AAATAATATG	TAATTTTTC	AGCTTATTAG	TGATATATT											3446
TATGGGTGGG	AATAAAATT	CTACTACAGA	AAAAAA	AAAAAA	AAAAAA											3501

(2) INFORMATION FOR SEQ ID NO:4:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 993 amino acids
- (B) TYPE: amino acid
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:4:

Met Pro Ala Leu Ala Arg Asp Ala Gly Thr Val Pro Leu Leu Val Val
-27 -25 -20 -15

Phe Ser Ala Met Ile Phe Gly Thr Ile Thr Asn Gln Asp Leu Pro Val
-10 -5 -5 1 5

Ile Lys Cys Val Leu Ile Asn His Lys Asn Asn Asp Ser Ser Val Gly
10 15 20

Lys Ser Ser Tyr Pro Met Val Ser Glu Ser Pro Glu Asp Leu Gly
25 30 35

Cys Ala Leu Arg Pro Gln Ser Ser Gly Thr Val Tyr Glu Ala Ala Ala
40 45 50

Val Glu Val Asp Val Ser Ala Ser Ile Thr Leu Gln Val Leu Val Asp
55 60 65

Ala Pro Gly Asn Ile Ser Cys Leu Trp Val Phe Lys His Ser Ser Leu
70 75 80 85

Asn Cys Gln Pro His Phe Asp Leu Gln Asn Arg Gly Val Val Ser Met
90 95 100

Val Ile Leu Lys Met Thr Glu Thr Gln Ala Gly Glu Tyr Leu Leu Phe
105 110 115

Ile Gln Ser Glu Ala Thr Asn Tyr Thr Ile Leu Phe Thr Val Ser Ile
120 125 130
Arg Asn Thr Leu Leu Tyr Thr Leu Arg Arg Pro Tyr Phe Arg Lys Met
135 140 145
Glu Asn Gln Asp Ala Leu Val Cys Ile Ser Glu Ser Val Pro Glu Pro
150 155 160 165
Ile Val Glu Trp Val Leu Cys Asp Ser Gln Gly Glu Ser Cys Lys Glu
170 175 180
Glu Ser Pro Ala Val Val Lys Lys Glu Lys Val Leu His Glu Leu
185 190 195
Phe Gly Thr Asp Ile Arg Cys Cys Ala Arg Asn Glu Leu Gly Arg Glu
200 205 210
Cys Thr Arg Leu Phe Thr Ile Asp Leu Asn Gln Thr Pro Gln Thr Thr
215 220 225
Leu Pro Gln Leu Phe Leu Lys Val Gly Glu Pro Leu Trp Ile Arg Cys
230 235 240 245
Lys Ala Val His Val Asn His Gly Phe Gly Leu Thr Trp Glu Leu Glu
250 255 260
Asn Lys Ala Leu Glu Gly Asn Tyr Phe Glu Met Ser Thr Tyr Ser
265 270 275
Thr Asn Arg Thr Met Ile Arg Ile Leu Phe Ala Phe Val Ser Ser Val
280 285 290
Ala Arg Asn Asp Thr Gly Tyr Tyr Thr Cys Ser Ser Lys His Pro
295 300 305
Ser Gln Ser Ala Leu Val Thr Ile Val Gly Lys Gly Phe Ile Asn Ala

310	315	320	325
Thr Asn Ser Ser Glu Asp Tyr Glu Ile Asp Gln Tyr Glu Glu Phe Cys			
330	335	340	345
Phe Ser Val Arg Phe Lys Ser Phe Ala Tyr Pro Gln Ile Arg Cys Thr Trp Thr	345	350	355
Phe Ser Arg Lys Ser Phe Pro Cys Glu Gln Lys Gly Leu Asp Asn Gly	360	365	370
Tyr Ser Ile Ser Lys Phe Cys Asn His Lys His Gln Pro Gly Glu Tyr	375	380	385
Ile Phe His Ala Glu Asn Asp Asp Ala Gln Phe Thr Lys Met Phe Thr	390	395	400
Leu Asn Ile Arg Arg Lys Pro Gln Val Leu Ala Glu Ala Ser Ala Ser	410	415	420
Gln Ala Ser Cys Phe Ser Asp Gly Tyr Pro Leu Pro Ser Trp Thr Trp	425	430	435
Lys Lys Cys Ser Asp Lys Ser Pro Asn Cys Thr Glu Glu Ile Thr Glu	440	445	450
Gly Val Trp Asn Arg Lys Ala Asn Arg Lys Val Phe Gly Gln Trp Val	455	460	465
Ser Ser Ser Thr Leu Asn Met Ser Glu Ala Ile Lys Gly Phe Leu Val	470	475	480
Lys Cys Cys Ala Tyr Asn Ser Leu Gly Thr Ser Cys Glu Thr Ile Leu	490	495	500
Leu Asn Ser Pro Gly Pro Phe Pro Phe Ile Gln Asp Asn Ile Ser Phe	505	510	515

Tyr Ala Thr Ile Gly Val Cys Leu Leu Phe Ile Val Val Leu Thr Leu
 520 525 530
 Leu Ile Cys His Lys Tyr Lys Lys Gln Phe Arg Tyr Glu Ser Gln Leu
 535 540 545
 Gln Met Val Gln Val Thr GLY Ser Ser Asp Asn Glu Tyr Phe Tyr Val
 550 555 560 565
 Asp Phe Arg Glu Tyr Glu Tyr Asp Leu Lys Trp Glu Phe Pro Arg Glu
 570 575 580 580
 Asn Leu Glu Phe Gly Lys Val Leu GLY Ser Gly Ala Phe GLY Lys Val
 585 590 595
 Met Asn Ala Thr Ala Tyr Gly Ile Ser Lys Thr GLY Val Ser Ile Gln
 600 605 610
 Val Ala Val Lys Met Leu Lys Glu Lys Ala Asp Ser Ser Glu Arg Glu
 615 620 625
 Ala Leu Met Ser Glu Leu Lys Met Met Thr Gln Leu GLY Ser His Glu
 630 635 640 645
 Asn Ile Val Asn Leu Leu Gly Ala Cys Thr Leu Ser Gly Pro Ile Tyr
 650 655 660
 Leu Ile Phe Glu Tyr Cys Cys Tyr GLY Asp Leu Leu Asn TYR Leu Arg
 665 670 675
 Ser Lys Arg Glu Lys Phe His Arg Thr Trp Thr Glu Ile Phe Lys Glu
 680 685 690
 His Asn Phe Ser Phe Tyr Pro Thr Phe Gln Ser His Pro Asn Ser Ser
 695 700 705
 Met Pro Gly Ser Arg Glu Val Gln Ile His Pro Asp Ser Asp Gln Ile

83

Cys Gln Leu Ala Asp Ala Glu Glu Ala Met Tyr Gln Asn Val Asp Gly
920 925 930

Arg Val Ser Glu Cys Pro His Thr Tyr Gln Asn Arg Arg Pro Phe Ser
935 940 945

Arg Glu Met Asp Leu Gly Leu Leu Ser Pro Gln Ala Gln Val Glu Asp
950 955 960 965

Ser

(2) INFORMATION FOR SEQ ID NO:5:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 5406 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: double
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(iii) HYPOTHETICAL: NO

(iv) ANTI-SENSE: NO

(v) FRAGMENT TYPE: N-terminal

(ix) FEATURE:

- (A) NAME/KEY: CDS
- (B) LOCATION: 208..4311

(ix) FEATURE:

- (A) NAME/KEY: mat_peptide
- (B) LOCATION: 265..4308

(ix) FEATURE:

- (A) NAME/KEY: sig_peptide

(B) LOCATION: 208..264

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:5:

CTCTGGAGCCA CAGCCCGGATA ACCTGGTGA	CCCCATTCCG CGGACACCCG CGGAGCCG	120
GCTCTCTGCC CAGGGCGAG ACTTCTTGC	GGGCCGGCTC TCCCCGGTCT CGGCCAGGA	180
GCTCTCTGCC CAGGGCGAG GTGCCAGG ATG	GAG AGC AAG GGC CTC CTA GCT	231
	Met Glu Ser Lys GLY Leu Leu Ala	
	-19 -15	
GTC GCT CTG TGG TTC TGC GTG	GAG ACC CGA GCC TCT GTG GGT TGT	279
val Ala Leu Trp Phe Cys Val	Glut Arg Ala Ser Val GLY Leu	
-10 -5	-1	5
CCT GGC GAT TTT CTC CAT CCC	CCC AAG CTC AGC ACA CAG AAA GAC ATA	327
Pro GLY Asp Phe Leu His Pro	Pro Lys Leu Ser Thr GLN Lys Asp Ile	
10	15	20
CTG ACA ATT TTG GCA AAT ACA	ACC CTT CAG ATT ACT TGC AGG GGA CAG	375
Leu Thr Ile Leu Ala Asn Thr	Leu GLN Ile Thr Cys Arg GLY GLN	
25	30	35
CGG GAC CTG GAC TGG CTT	CCC AAT GCT CAG CGT GAT TCT GAG GAA	423
Arg Asp Leu ASP Trp Leu	PRO Asn Ala GLN Arg Asp Ser Glu Glu	
40	45	50
AGG GTA TTG GTG ACT GAA TGC	GGC GGT GAC AGT ATC TTG TGC AAA	471
Arg Val Leu Val Thr Glu Cys	GLY GLY GLY Asp Ser Ile Phe Cys Lys	
55	60	65
ACA CTC ACC ATT CCC ACG GTG GTT GGA	ATG GAT ACTT GCC TAC AAG	519
Thr Leu Thr Ile Pro Arg Val Val	GLY ASN Asp Thr GLY Ala TYR Lys	

70	75	80	85
TCG TAC CGG GAC GTC GAC ATA GCC TCC ACT GTT TAT GTC TAT GTT Cys Ser Tyr Arg Asp Val Asp Ile Ala Ser Thr Val Tyr Val Val 90			567
CGA GAT TAC AGA TCA CCA TTC ATC GCC TCT GTC AGT GAC CAG CAT GGC Arg Asp Tyr Arg Ser Pro Phe Ile Ala Ser Val Ser Asp Gln His Gly 105			615
ATC GTG TAC ATC ACC GAG AAC AAG AAC AAA ACT GTG GTG ATC CCC TGC Ile Val Tyr Ile Thr Glu Asn Lys Asn Lys Thr Val Val Ile Pro Cys 120			663
CGA GGG TCG ATT TCA AAC CTC AAT GTG TCT CTT GCT AGG TAT CCA Arg Gly Ser Ile Ser Asn Leu Asn Val Ser Leu Cys Ala Arg Tyr Pro 135			711
GAA AAG AGA TTT GTT CCG GAT GGA AAC AGA ATT TCC TGG GAC AGC GAG Glu Lys Arg Phe Val Pro Asp Gly Asn Arg Ile Ser Trp Asp Ser Glu 150			759
ATA GGC TTT ACT CTC CCC AGT TAC ATG ATC AGC TAT GCC GGC ATG GTC Ile Gly Phe Thr Leu Pro Ser Tyr Met Ile Ser Tyr Ala Gly Met Val 170			807
TTC TGT GAG GCA AAG ATC AAT GAT GAA ACC TAT CAG TCT ATC ATG TAC Phe Cys Glu Ala Lys Ile Asn Asp Glu Thr Tyr Gln Ser Ile Met Tyr 185			855
ATA GTT GTG GTT GTA GGA TAT AGG ATT TAT GAT GTG ATT CTG AGC CCC Ile Val Val Val Gly Tyr Arg Ile Tyr Asp Val Ile Leu Ser Pro 200			903
CCG CAT GAA ATT GAG CTA TCT GCC GGA GAA AAA CTT GTC TTA AAT TGT Pro His Glu Ile Glu Leu Ser Ala Gly Glu Lys Leu Val Leu Asn Cys 215			951

66

ACA GCG AGA ACA GAG CTC AAT GTG GGG CTT GAT TTC ACC TGG CAC TCT	999
Thr Ala Arg Thr Glu Leu Asn Val Gly Leu Asp Phe Thr Trp His Ser	245
230 235	
CCA CCT TCA AAG TCT CAT AAG AAG ATT GTA AAC CGG GAT GTG AAA	1047
Pro Pro Ser Lys Ser His His Lys Lys Ile Val Asn Arg Asp Val Lys	260
250 255	
CCC TTT CCT GGG ACT GTG GCG AAG ATG TTT TTG AGC ACC TTG ACA ATA	1095
Pro Phe Pro GLY Thr Val Ala Lys Met Phe Leu Ser Thr Leu Thr Ile	275
265 270	
GAA AGT GTG ACC AAG AGT GAC CAA GGG GAA TAC ACC TGT GTA GCG TCC	1143
Glu Ser Val Thr Lys Ser Asp Gln Gly Glu Tyr Thr Cys Val Ala Ser	290
280 285	
AGT GGA CGG ATG ATC AAG AGA AAT AGA ACA TTT GTC CGA GTT CAC ACA	1191
Ser Gly Arg Met Ile Lys Arg Asn Arg Thr Phe Val Arg Val His Thr	305
295 300	
AAG CCT TTT ATT GCT TTC GGT AGT GGG ATG AAA TCT TTG GTG GAA GCC	1239
Lys Pro Phe Ile Ala Phe Gly Ser Gly Met Lys Ser Leu Val Glu Ala	325
310 315	
ACA GTG GGC AGT CAA GTC CGA ATC CCT GTG AAG TAT CTC AGT TAC CCA	1287
Thr Val Gly Ser Gln Val Arg Ile Pro Val Lys Tyr Leu Ser Tyr Pro	340
330 335	
GCT CCT GAT ATC AAA TGG TAC AGA AAT GGA AGG CCC ATT GAG TCC AAC	1335
Ala Pro Asp Ile Lys Trp Tyr Arg Asn Gly Arg Pro Ile Glu Ser Asn	355
345 350	
TAC ACA ATG ATT GTT GGC GAT GAA CTC ACC ATC ATG GAA GTG ACT GAA	1383
Tyr Thr Met Ile Val Gly Asp Glu Leu Thr Ile Met Glu Val Thr Glu	370
360 365	
AGA GAT GCA GGA AAC TAC ACG GTC ATC CTC ACC AAC CCC ATT TCA ATG	1431

Arg	Asp	Ala	Gly	Asn	Tyr	Thr	Val	Ile	Leu	Thr	Asn	Pro	Ile	Ser	Met
375															
GAG	AAA	CAG	AGC	CAC	ATG	GTC	TCT	CTG	GTT	GTG	AAT	GTC	CCA	CCC	CAG
Glu	Lys	Gln	Ser	His	Met	Val	Ser	Leu	Val	Val	Asn	Val	Pro	Pro	Gln
390															405
ATC	GGT	GAG	AAA	GCC	TTG	ATC	TCG	CCT	ATG	GAT	TCC	TAC	CAG	TAT	GGG
Ile	Gly	Glu	Lys	Ala	Leu	Ile	Ser	Pro	Met	Asp	Ser	Tyr	Gln	Tyr	Gly
410															420
ACC	ATG	CAG	ACA	TTG	ACA	TGC	ACA	GTC	TAC	GCC	AAC	CCT	CCC	CTG	CAC
Thr	Met	Gln	Thr	Leu	Thr	Cys	Thr	Val	Val	TYR	Ala	Asn	Pro	Pro	Leu
425															His
CAC	ATC	CAG	TGG	TAC	TGG	CAG	CTA	GAA	GAA	GCC	TGC	TCC	TAC	AGA	CCC
His	Ile	Gln	Trp	Tyr	Trp	Gln	Leu	Glu	Glu	Ala	Cys	Ser	Tyr	Arg	Pro
440															450
GGC	CAA	ACA	AGC	CCG	TAT	GCT	TGT	AAA	GAA	TGG	AGA	CAC	GTG	GAG	GAT
Gly	Gln	Thr	Ser	Pro	Tyr	Ala	CYS	LYS	GLU	TRP	ARG	HIS	VAL	GLU	ASP
455															465
TTC	CAG	GGG	GGA	AAC	AAG	ATC	GAA	GTC	ACC	AAA	AAC	CAA	TAT	GCC	CTG
Phe	Gln	Gly	Gly	Asn	Lys	Ile	Glu	Val	Thr	LYS	Asn	Gln	Tyr	Ala	Leu
470															485
ATT	GAA	GGA	AAA	AAC	AAA	ACT	GTA	AGT	ACG	CTG	GTC	ATC	CAA	GCT	GCC
Ile	Glu	Gly	Lys	Asn	Lys	Thr	Val	Ser	Thr	Leu	Val	Ile	Gln	Ala	Ala
															500
AAC	GTG	TCA	GCG	TTG	TAC	AAA	TGT	GAA	GCC	ATC	AAC	AAA	GCG	GGA	CGA
Asn	Val	Ser	Ala	Leu	Tyr	Lys	Cys	Glu	Ala	Ile	Asn	Lys	Ala	Gly	Arg
505															515
GGA	GAG	AGG	GTC	ATC	TCC	TTC	CAT	GTG	ATC	AGG	GGT	CCT	GAA	ATT	ACT
Gly	Glu	Arg	Val	Ile	Ser	Phe	His	Val	Ile	Arg	Gly	Pro	Glu	Ile	Thr
															1863

	520	525	530	
GTG CAA CCT GCT GCC CAG CCA ACT GAG CAG GAG AGT GTG TCC CTG TTG				1911
val Gln Pro Ala Ala Gln Pro Thr Glu Gln Glu Ser Val Ser Leu Leu	535	540	545	
TGC ACT GCA GAC AGA AAT ACG TTT GAG AAC CTC ACG TGG TAC AAG CTT				1959
Cys Thr Ala Asp Arg Asn Thr Phe Glu Asn Leu Thr Trp Tyr Lys Leu	550	555	560	565
GGC TCA CAG GCA ACA TCG GTC CAC ATG GGC GAA TCA CTC ACA CCA GTT				2007
Gly Ser Gln Ala Thr Ser Val His Met Gly Glu Ser Leu Thr Pro Val	570	575	580	
TGC AAG AAC TTG GAT GCT CTT TGG AAA CTG AAT GGC ACC ATG TTT TCT				2055
Cys Lys Asn Leu Asp Ala Leu Trp Lys Leu Asn Gly Thr Met Phe Ser	585	590	595	
AAC AGC ACA AAT GAC ATC TTG ATT GTG GCA TTT CAG AAT GCC TCT CTG				2103
Asn Ser Thr Asn Asp Ile Leu Ile Val Ala Phe Gln Asn Ala Ser Leu	600	605	610	
CAG GAC CAA GGC GAC TAT GTT TGC TCT GCT CAA GAT AAG ACC AAG				2151
Gln Asp Gln Gly Asp Tyr Val Cys Ser Ala Gln Asp Lys Lys Thr Lys	615	620	625	
AAA AGA CAT TGC GTC AAA CAG CTC ATC ATC CTA GAG CGC ATG GCA				2199
Lys Arg His Cys Leu Val Lys Gln Leu Ile Ile Leu Glu Arg Met Ala	630	635	640	645
CCC ATG ATC ACC GGA AAT CTG GAG AAT CAG ACA ACC ATT GGC GAG				2247
Pro Met Ile Thr Gly Asn Leu Glu Asn Gln Thr Thr Ile Gly Glu	650	655	660	
ACC ATT GAA GTG ACT TGC CCA GCA TCT GGA AAT CCT ACC CCA CAC ATT				2295
Thr Ile Glu Val Thr Cys Pro Ala Ser Gly Asn Pro Thr Pro His Ile	665	670	675	

ACA	TGG	TTC	AAA	GAC	AAC	GAG	ACC	CTG	GTA	GAA	GAT	TCA	GGC	ATT	GTA	2343
Thr	Trp	Phe	Lys	Asp	Asn	Glu	Thr	Leu	Val	Glu	Asp	Ser	Gly	Ile	Val	
																680
CTG	AGA	GAT	GGG	AAC	CGG	AAC	CTG	ACT	ATC	CGC	AGG	GTG	AAG	GAG		2391
Leu	Arg	Asp	Gly	Asn	Arg	Asn	Leu	Thr	Ile	Arg	Arg	Val	Arg	Lys	Glu	
																685
GAT	GGA	GGC	CTC	TAC	ACC	TGC	CAG	GCC	TGC	AAT	GTC	CTT	GGC	TGT	GCA	2439
Asp	Gly	Gly	Leu	Tyr	Thr	Cys	Gln	Ala	Cys	Asn	Val	Leu	Gly	Cys	Ala	
																710
AGA	GCC	GAG	ACG	CTC	TTC	ATA	ATA	GAA	GGT	GCC	CAG	GAA	AAG	ACC	AAC	2487
Arg	Ala	Glu	Thr	Leu	Phe	Ile	Ile	Glu	Gly	Ala	Gln	Glu	Lys	Thr	Asn	
																730
TTG	GAA	GTC	ATT	ATC	CTC	GTC	GGC	ACT	GCA	GTG	ATT	GCC	ATG	TTC	TTC	2535
Leu	Glu	val	Ile	Ile	Ile	Leu	Val	Gly	Thr	Ala	Val	Ile	Ala	Met	Phe	
																745
TGC	CTC	CTT	GTC	ATT	CTC	GTA	CGG	ACC	GTT	AAG	CGG	GCC	AAT	GAA		2583
Trp	Leu	Leu	Leu	Val	Ile	Leu	Val	Arg	Thr	Val	Lys	Arg	Ala	Asn	Glu	
																760
GGG	GAA	CTG	AAG	ACA	GGC	TAC	TTG	TCT	ATT	GTC	ATG	GAT	CCA	GAT	GAA	2631
Gly	Glu	Leu	Lys	Thr	Gly	Tyr	Leu	Ser	Ile	Val	Met	Asp	Pro	Asp	Glu	
																775
TTC	CCC	TTG	GAT	GAG	CGC	TGT	GAA	CGC	TTG	CCT	TAT	GAT	GCC	ACG	AAG	2679
Leu	Pro	Leu	Asp	Glu	Arg	Cys	Glu	Arg	Leu	Pro	Tyr	Asp	Ala	Ser	Lys	
																790
TGG	GAA	TTC	CCC	AGG	GAC	CGG	CTG	AAA	CTA	GGA	AAA	CCT	CTT	GGC	CGC	
Trp	Glu	Phe	Pro	Arg	Asp	Arg	Leu	Lys	Leu	Gly	Lys	Pro	Leu	Gly	Arg	
																810
GGT	GCC	TTC	GGC	CAA	GTG	ATT	GAG	GCA	GAC	GCT	TTT	GGA	ATT	GAC	AAG	2775

Gly Ala Phe Gly Gln Val Ile Glu Ala Asp Ala Phe Gly Ile Asp Lys
 825 830 835
 ACA GCC ACT TGC AAA ACA GTA GCC GTC AAG ATG TTG AAA GAA GGA GCA
 Thr Ala Thr Cys Lys Thr Val Ala Val Lys Met Leu Lys Glu Gly Ala
 840 845 850
 ACA CAC AGC GAG CAT CGA GCC CTC ATG TCT GAA CTC AAG ATC CTC ATC
 Thr His Ser Glu His Arg Ala Leu Met Ser Glu Leu Lys Ile Leu Ile
 855 860 865
 CAC ATT GGT CAC CAT CTC AAT GTG GTG AAC CTC CTA GGC GCC TGC ACC
 His Ile Gly His His Leu Asn Val Val Asn Leu Leu Gly Ala Cys Thr
 870 875 880 885
 AAG CCG GGA GGG CCT CTC ATG GTG ATT GTG GAA TTC TCG AAG TTT GCA
 Lys Pro Gly Gly Pro Leu Met Val Ile Val Glu Phe Ser Lys Phe Gly
 890 895 900
 AAC CTA TCA ACT TAC TTA CGG GGC AAG AGA AAT GAA TTT GTT CCC TAT
 Asn Leu Ser Thr Tyr Tyr Leu Arg Gly Lys Arg Asn Glu Phe Val Pro Tyr
 905 910 915
 AAG AGC AAA GGG GCA CGC TTC CGC CAG GGC AAG GAC TAC GTT GGG GAG
 Lys Ser Lys Gly Ala Arg Phe Arg Gln Gly Lys Asp Tyr Val Gly Glu
 920 925 930
 CTC TCC GTG GAT CTG AAA AGA CGC TTG GAC AGC ATC ACC AGC CAG
 Leu Ser Val Asp Leu Lys Arg Arg Leu Asp Ser Ile Thr Ser Ser Gln
 935 940 945
 AGC TCT GCC AGC TCA GGC TTT GTT GAG GAG AAA TCG CTC AGT GAT GTA
 Ser Ser Ala Ser Ser G1Y Phe Val Glu Glu Lys Ser Leu Ser Asp Val
 950 955 960
 GAG GAA GAA GCT TCT GAA GAA CTG TAC AAG GAC TTC CTG ACC TTG
 Glu Glu Glu Ala Ser Glu Glu Leu Tyr Lys Asp Phe Leu Thr Leu
 3207

2

970	GAG CAT CTC ATC TGT TAC AGC TTC CAA GTG GCT AAG GGC ATG GAG TTC	975	980
Gl u His Leu Ile Cys Tyr Ser Phe Gln Val Ala Lys Gl y Met Gl u Phe	990	995	3255
985			
TrG GCA TCA AGG AAG TGT ATC CAC AGG GAC CTC GCA GCA AAC ATT	1000		3303
Leu Ala Ser Arg Lys Cys Ile His Arg Asp Leu Ala Ala Arg Asn Ile	1005	1010	
1010			
CTC CTA TCG GAG AAG AAT GTG GTT AAG ATC TGT GAC TTC GGC TTG GCC	1015		3351
Leu Leu Ser Glu Lys Asn Val Val Lys Ile Cys Asp Phe Gl y Leu Ala	1020	1025	
1020			
CGG GAC ATT TAT AAA GAC CCG GAT TAT GTC AGA AAA GGA GAT GCC CGA	1030		3399
Arg Asp Ile Tyr Lys Asp Pro Asp Tyr Val Arg Lys Gl y Asp Ala Arg	1035	1040	
1035			
CTC CCTT TTG AAG TGG ATG GCC CCG GAA ACC ATT TTT GAC AGA GTA TAC	1045		3447
Leu Pro Leu Lys Trp Met Ala Pro Glu Thr Ile Phe Asp Arg Val Tyr	1050	1055	
1050			
ACA ATT CAG AGC GAT GTG TGG TCT TTC GGT GTG TTG CTC TGG GAA ATA	1060		3495
Thr Ile Gln Ser Asp Val Trp Ser Phe Gl y Val Leu Leu Trp Gl u Ile	1065	1070	
1065			
T TT TCC TTA GGT GCC TCC CCA TAC CCT GGG GTC AAG ATT GAT GAA GAA	1075		3543
Phe Ser Leu Gl y Ala Ser Pro Tyr Pro Gl y Val Lys Ile Asp Gl u Gl u	1080	1085	
1080			
T TT TGT AGG AGA TTG AAA GAA GGA ACT AGA ATG CGG GCT CCT GAC TAC	1090		3591
Phe Cys Arg Arg Leu Lys Gl u Gl y Thr Arg Met Arg Ala Pro Asp Tyr	1095	1100	
1095			
ACT ACC CCA GAA ATG TAC CAG ACC ATG CTG GAC TGC TGG CAT GAG GAC	1110		3639
Thr Thr Pro Glu Met Tyr Gln Thr Met Leu Asp Cys Trp His Gl u Asp	1115	1120	
1115			

72

CCC AAC CAG AGA CCC TCG TTT TCA GAG TTG GTG GAG CAT TTG GGA AAC	3687
Pro Asn Gln Arg Pro Ser Phe Ser Glu Leu Val Glu His Leu Gly Asn	
1130 1135 1140	
CTC CTG CAA GCA AAT GCG CAG CAG GAT GGC AAA GAC TAT ATT GTT CTT	3735
Leu Leu Gln Ala Asn Ala Gln Gln Asp Gly Lys Asp Tyr Ile Val Leu	
1145 1150 1155	
CCA ATG TCA GAG ACA CTG AGC ATG GAA GAG GAT TCT GGA CTC TCC CTG	3783
Pro Met Ser Glu Thr Leu Ser Met Glu Glu Asp Ser Gly Leu Ser Leu	
1160 1165 1170	
CCT ACC TCA CCT GTT TCC TGT ATG GAG GAA GAG GAA GTG TGC GAC CCC	3831
Pro Thr Ser Pro Val Ser Cys Met Glu Glu Glu Val Cys Asp Pro	
1175 1180 1185	
AAA TTC CAT TAT GAC AAC ACA GCA GGA ATC AGT CAT TAT CTC CAG AAC	3879
Lys Phe His Tyr Asp Asn Thr Ala Gly Ile Ser His Tyr Leu Gln Asn	
1190 1195 1200 1205	
AGT AAG CGA AAG AGC CGG CCA GTG AGT GTA AAA ACA TTT GAA GAT ATC	3927
Ser Lys Arg Lys Ser Arg Pro Val Ser Val Lys Thr Phe Glu Asp Ile	
1210 1215 1220	
CCA TTG GAG GAA CCA GAA GAA GTG ATC CCA GAT GAC AGC CAG ACA	3975
Pro Leu Glu Glu Pro Glu Val Lys Val Ile Pro Asp Asp Ser Gln Thr	
1225 1230 1235	
GAC AGT GGG ATG GTC CTT GCA TCA GAA GAG CTG AAA ACT CTG GAA GAC	4023
Asp Ser Gly Met Val Leu Ala Ser Glu Glu Leu Lys Thr Leu Glu Asp	
1240 1245 1250	
AGG AAC AAA TTA TCT CCA TCT TTT GGT GGA ATG ATG CCC AGT AAA AGC	4071
Arg Asn Lys Leu Ser Pro Ser Phe Gly Gly Met Met Pro Ser Lys Ser	
1255 1260 1265	
AGG GAG TCT GTG GCC TCG GAA GGC TCC AAC CAG ACC AGT GGC TAC CAG	4119

Arg Glu Ser val Ala ser Glu Gly ser Asn Gln Thr Ser Gly Tyr Tyr Gln
 1270 1275 1280 1285
 TCT GGG TAT CAC TCA GAT GAC ACA GAC ACC ACC GTG TAC TCC AGC GAC
 Ser Gly Tyr His Ser Asp Asp Thr Asp Thr Thr Val Tyr Ser Ser Asp 4167
 1290 1295 1300
 GAG GCA GGA CTT TTA AAG ATG GTG GAT GCT GCA GTT CAC GCT GAC TCA
 Glu Ala Gly Leu Leu Lys Met Val Asp Ala Ala Val His Ala Asp Ser 4215
 1305 1310 1315
 GGG ACC ACA CTG CAG CTC ACC TCC TGT TTA AAT GGA AGT GGT CCT GTC
 Gly Thr Thr Leu Gln Leu Thr Ser Cys Leu Asn Gly Ser Gly Pro Val 4263
 1320 1325 1330
 CCG GCT CCG CCC CCA ACT CCT GGA AAT CAC GAG AGA GGT GCT GCT TAGATTTCA 4318
 Pro Ala Pro Pro Pro Thr Pro Gly Asn His Glu Arg Gly Ala Ala
 1335 1340 1345
 ACTGTGTTC TTTCCACCAC CCGGAAGTAG CCACATTGA TTTTCATTG TGGAGGGAGGG 4378
 ACCTCAGGACT GCAAGGAGCT TGTCCCTCAGG GCATTCCAG AGAAAGATGCC AGAACCCAA
 GAATGTGTG ACTCTACTCT CTTTTCCATT CATTAAAG TCCTATATAA TGTGCCCTGC 4438
 TGTGGTCTCA CTACCAGTTA AAGCAAAGA CTTTCAAACAA CGTGGACTCT GTCCCTCCAAG 4498
 AAGTGGCAAC GGCACCTCTG TGAAACTGGA TCGAATGGGC AATGCTTTGT GTGTTGAGGA
 TGGGTGAGAT GTCCCAGGG CGAAGTCTGTC TACCTTGGAG GCTTTGTGGA GGATGCCGCT 4678
 ATGAGCCAAG TGTTAAGTGT GGGATGTGGA CTGGGAGGAA GGAAGGGCA AGCCGTCGG 4738
 AGAGCGGTG GAGCCTGCAG ATGCATTGTC CTGGCTCTGG TGGAGGTGG CCTTGTGGCCT 4798
 GTCAGGAAAC GCAAAGGGG CCGGAGGGT TTGGTTTGG AAGGTTTGG TGCTCTTCAC 4858

AGTCGGGTTA CAGGGAGT CCCTGTGGCG TTTCTACTC CTAATGAGAG TTCCTTCGGG 4918
 ACTCTTACGT GTCTCCTGGC CTGGCCCG GAAGGAAATG ATGCAGCTTG CTCCCTTCCTC 4978
 ATCTCTCAGG CTGTGCCTTA ATTAGAACCA CCAAAAGAGA GGAACGTCGG CAGAGGCTCC 5038
 TGACGGGCC GAAGAATTGT GAGAACAGAA CAGAAACTCA GGTTTCTGC TGGGTGGAGA 5098
 CCCACGTGGC GCCCTGGTGG CAGGTCTGAG GGTtCTCTGT CAAGTGGGG TAAAGGCTCA 5158
 GGCTGGTGT CTTCCCTAT CTCCACTCCT GTCAGGGCCC CAAGTCCTCA GTATTTCAGC 5218
 TTTGTGGCTT CCTGATGGCA GAAAATCTT ATTGGTTGG TTTGGCTCTCC AGATAATCAC 5278
 TAGCCAGATT TCGAAATTAC TTTTAGCCG AGGTTATGAT AACATCTACT GTATCCTTTA 5338
 GAATTAAAC CTATAAAACT ATGTCTACTG GTTCTGCCT GTGTGCTTAT GTTAAAAAA 5398
 AAAA
Q
 AAAA
 5406

(2) INFORMATION FOR SEQ ID NO: 6:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 1367 amino acids
- (B) TYPE: amino acid
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 6:

Met Glu Ser Lys Gly Leu Leu Ala Val Ala Leu Trp Phe Cys Val Glu
 -19 -15 -10 -5
 Thr Arg Ala Ala Ser Val Gly Leu Pro Gly Asp Phe Leu His Pro Pro
 5 10

Lys Leu Ser Thr Gln Lys Asp Ile Leu Thr Ile Leu Ala Asn Thr Thr
15 20 25

Leu Gln Ile Thr Cys Arg Gly Gln Arg Asp Leu Asp Trp Leu Trp Pro
30 35 40 45

Asn Ala Gln Arg Asp Ser Glu Glu Arg Val Leu Val Thr Glu Cys Gly
50 55 60

Gly Gly Asp Ser Ile Phe Cys Lys Thr Leu Thr Ile Pro Arg Val Val
65 70 75

Gly Asn Asp Thr Gly Ala Tyr Lys Cys Ser Tyr Arg Asp Val Asp Ile
80 85 90

Ala Ser Thr Val Tyr Val Tyr Val Arg Asp Tyr Arg Ser Pro Phe Ile
95 100 105

Ala Ser Val Ser Asp Gln His Gly Ile Val Tyr Ile Thr Glu Asn Lys
110 115 120 125

Asn Lys Thr Val Val Ile Pro Cys Arg Gly Ser Ile Ser Asn Leu Asn
130 135 140

Val Ser Leu Cys Ala Arg Tyr Pro Glu Lys Arg Phe Val Pro Asp Gly
145 150 155

Asn Arg Ile Ser Trp Asp Ser Glu Ile Gly Phe Thr Leu Pro Ser Tyr
160 165 170

Met Ile Ser Tyr Ala Gly Met Val Phe Cys Glu Ala Lys Ile Asn Asp
175 180 185

Glu Thr Tyr Gln Ser Ile Met Tyr Ile Val Val Val Gly Tyr Arg
190 195 200 205

Ile Tyr Asp Val Ile Leu Ser Pro His Glu Ile Glu Leu Ser Ala

96

210	Gly Glu Lys Leu Val Leu Asn Cys Thr Ala Arg Thr Glu Leu Asn Val	225
		230
240	Gly Leu Asp Phe Thr Trp His Ser Pro Pro Ser Lys Ser His His Lys	245
		250
255	Lys Ile Val Asn Arg Asp Val Lys Pro Phe Pro Gly Thr Val Ala Lys	260
		265
270	Met Phe Leu Ser Thr Leu Thr Ile Glu Ser Val Thr Lys Ser Asp Gln	275
		280
	Gly Glu Tyr Thr Cys Val Ala Ser Ser Gly Arg Met Ile Lys Arg Asn	295
		300
290	Arg Thr Phe Val Arg Val His Thr Lys Pro Phe Ile Ala Phe Gly Ser	305
		310
305	Gly Met Lys Ser Leu Val Glu Ala Thr Val Gly Ser Gln Val Arg Ile	320
		325
320	Pro Val Lys Tyr Leu Ser Tyr Pro Ala Pro Asp Ile Lys Trp Tyr Arg	335
		340
335	Asn Gly Arg Pro Ile Glu Ser Asn Tyr Thr Met Ile Val Gly Asp Glu	350
		355
350	Leu Thr Ile Met Glu Val Thr Glu Arg Asp Ala Gly Asn Tyr Thr Val	370
		375
375	Ile Leu Thr Asn Pro Ile Ser Met Glu Lys Gln Ser His Met Val Ser	390
		395
390	Leu Val Val Asn Val Pro Pro Gln Ile Gly Glu Lys Ala Leu Ile Ser	400
		405
405		410

Pro Met Asp Ser Tyr Gln Tyr Gly Thr Met Gln Thr Leu Thr Cys Thr
415 420 425

Val Tyr Ala Asn Pro Pro Leu His His Ile Gln Trp Tyr Trp Gln Leu
430 435 440 445

Glu Glu Ala Cys Ser Tyr Arg Pro Gly Gln Thr Ser Pro Tyr Ala Cys
450 455 460

Lys Glu Trp Arg His Val Glu Asp Phe Gln Gly Gly Asn Lys Ile Glu
465 470 475 475

Val Thr Lys Asn Gln Tyr Ala Leu Ile Glu Gly Lys Asn Lys Thr Val
480 485 490

Ser Thr Leu Val Ile Gln Ala Ala Asn Val Ser Ala Leu Tyr Lys Cys
495 500 505

Glu Ala Ile Asn Lys Ala Gly Arg Gly Glu Arg Val Ile Ser Phe His
510 515 520 525

Val Ile Arg Gly Pro Glu Ile Thr Val Gln Pro Ala Ala Gln Pro Thr
530 535 540

Glu Gln Glu Ser Val Ser Leu Leu Cys Thr Ala Asp Arg Asn Thr Phe
545 550 555

Glu Asn Leu Thr Trp Tyr Lys Leu Gly Ser Gln Ala Thr Ser Val His
560 565 570

Met Gly Glu Ser Leu Thr Pro Val Cys Lys Asn Leu Asp Ala Leu Trp
575 580 585

Lys Leu Asn Gly Thr Met Phe Ser Asn Ser Thr Asn Asp Ile Leu Ile
590 595 600 605

Val Ala Phe Gln Asn Ala Ser Leu Gln Asp Gln Gly Asp Tyr Val Cys

Lys Leu Gly Lys Pro Leu Gly Arg Gly Ala Phe Gly Gln Val Ile Glu
 815 820 825
 Ala Asp Ala Phe Gly Ile Asp Lys Thr Ala Thr Cys Lys Thr Val Ala
 830 835 840 845
 Val Lys Met Leu Lys Glu GLY Ala Thr His Ser Glu His Arg Ala Leu
 850 855 860 865
 Met Ser Glu Leu Lys Ile Leu Ile His Ile Gly His His Leu Asn Val
 865 870 875 880
 Val Asn Leu Leu Gly Ala Cys Thr Lys Pro Gly Gly Pro Leu Met Val
 880 885 890 895
 Ile Val Glu Phe Ser Lys Phe Gly Asn Leu Ser Thr Tyr Leu Arg Gly
 900 905 910
 Lys Arg Asn Glu Phe Val Pro Tyr Lys Ser Lys Gly Ala Arg Phe Arg
 915 920 925
 Gln Gly Lys Asp Tyr Val Gly Glu Leu Ser Val Asp Leu Lys Arg Arg
 930 935 940
 Leu Asp Ser Ile Thr Ser Ser Gln Ser Ser Ala Ser Ser Gly Phe Val
 945 950 955
 Glu Glu Lys Ser Leu Ser Asp Val Glu Glu Ala Ser Glu Glu
 960 965 970
 Leu Tyr Lys Asp Phe Leu Thr Leu Glu His Leu Ile Cys Tyr Ser Phe
 975 980 985
 Gln Val Ala Lys GLY Met Glu Phe Leu Ala Ser Arg Lys Cys Ile His
 990 995 1000 1005
 Arg Asp Leu Ala Ala Arg Asn Ile Leu Ser Glu Lys Asn Val Val

Ser Val Lys Thr Phe Glu Asp Ile Pro Leu Glu Glu Pro Glu Val Lys
 1215 1220 1225
 Val Ile Pro Asp Asp Ser Gln Thr Asp Ser Gly Met Val Leu Ala Ser
 1230 1235 1240 1245
 Glu Glu Leu Lys Thr Leu Glu Asp Arg Asn Lys Leu Ser Pro Ser Phe
 1250 1255 1260
 Gly Gly Met Met Pro Ser Lys Ser Arg Glu Ser Val Ala Ser Glu Gly
 1265 1270 1275
 Ser Asn Gln Thr Ser Gly Tyr Gln Ser Gly Tyr His Ser Asp Asp Thr
 1280 1285 1290 1295
 Asp Thr Thr Val Tyr Ser Ser Asp Glu Ala Gly Leu Leu Lys Met Val
 1295 1300 1305
 Asp Ala Ala Val His Ala Asp Ser Gly Thr Thr Leu Gln Leu Thr Ser
 1310 1315 1320 1325
 Cys Leu Asn Gly Ser Gly Pro Val Pro Ala Pro Pro Pro Thr Pro Gly
 1330 1335 1340
 Asn His Glu Arg Gly Ala Ala
 1345

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(2) INFORMATION FOR SEQ ID NO:7:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 96 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: cDNA

- (iii) HYPOTHETICAL: NO
- (iv) ANTI-SENSE: NO

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:7:

AATTCTCGA CTTCCTGTCA CCATGAGTGC ACTTCTGATC CTAGCCCTTG TGGAGCTGC
TGTGCTGAC TACAAAGATG ATGATGACAA GATCTA

(2) INFORMATION FOR SEQ ID NO:8:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 96 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(iii) HYPOTHETICAL: NO

(iv) ANTI-SENSE: NO

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:8:

AGCTTAGATC TTGTCTCAT CATCTTGTA GTCAGCAACA GCAGCTCCCA CAGAGGCTAG
GATCAGAAGT GCACTCATGG TGACAGAAAG TCGACCC

(2) INFORMATION FOR SEQ ID NO:9:

(i) SEQUENCE CHARACTERISTICS:

—

- (A) LENGTH: 30 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(i) MOLECULE TYPE: cDNA

(ii) HYPOTHETICAL: NO

(iv) ANTI-SENSE: NO

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:9:

TGAGAAGATC TCAAACCAAG ACCTGCCTGT

(2) INFORMATION FOR SEQ ID NO:10:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 34 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(iii) HYPOTHETICAL: NO

(iv) ANTI-SENSE: NO

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:10:

CCAATGGCGG CCGCTCAGGA GATGTTGTCT TGGA

(2) INFORMATION FOR SEQ ID NO:11:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 14 amino acids
- (B) TYPE: amino acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

(iii) HYPOTHETICAL: NO

(iv) ANTI-SENSE: NO

(v) FRAGMENT TYPE: N-terminal

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:11:
Ala Gln Ser Leu Ser Phe Xaa Phe Thr Lys Phe Asp Leu Asp
1 5 10

CLAIMS

What is claimed is:

5

1. A protein that binds to the Flk2 receptor comprising the amino acid sequence AQSLSF^XFTKFDLD shown in SEQ. ID. NO. 11, wherein X is any amino acid.

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Fig. 1a.1

GCGGCCTGGC TACCGCGCGC TCCGGAGGCC ATG CGG GCG TTG GCG CAG CGC AGC
 Met Arg Ala Leu Ala Gln Arg Ser
 -27 -25 -20

GAC CGG CGG CTG CTG CTG CTT GTT TTG TCA GTA ATG ATT CTT GAG
 Asp Arg Arg Leu Leu Leu Val Val Leu Ser Val Met Ile Leu Glu
 -15 -10 -5

ACC GTT ACA AAC CAA GAC CTG CCT GTG ATC AAG TGT GTT TTA ATC AGT
 Thr Val Thr Asn Gln Asp Leu Pro Val Ile Lys Cys Val Leu Ile Ser
 1 5 10

CAT GAG AAC AAT GGC TCA TCA GCG GGA AAG CCA TCA TCG TAC CGA ATG
 His Glu Asn Asn Gly Ser Ser Ala Gly Lys Pro Ser Ser Tyr Arg Met
 15 20 25

GTG CGA GGA TCC CCA GAA GAC CTC CAG TGT ACC CCG AGG CGC CAG AGT
 Val Arg Gly Ser Pro Glu Asp Leu Gln Cys Thr Pro Arg Arg Gln Ser
 30 35 40 45

GAA GGG ACG GTA TAT GAA GCG GCC ACC GTG GAG GTG GCC GAG TCT GGG
 Glu Gly Thr Val Tyr Glu Ala Ala Thr Val Glu Val Ala Glu Ser Gly
 50 55 60

TCC ATC ACC CTG CAA GTG CAG CTC GCC ACC CCA GGG GAC CTT TCC TGC
 Ser Ile Thr Leu Gln Val Gln Leu Ala Thr Pro Gly Asp Leu Ser Cys
 65 70 75

CTC TGG GTC TTT AAG CAC AGC TCC CTG GGC TGC CAG CCG CAC TTT GAT
 Leu Trp Val Phe Lys His Ser Ser Leu Gly Cys Gln Pro His Phe Asp
 80 85 90

TTA CAA AAC AGA GGA ATC GTT TCC ATG GCC ATC TTG AAC GTG ACA GAG
 Leu Gln Asn Arg Gly Ile Val Ser Met Ala Ile Leu Asn Val Thr Glu
 95 100 105

ACC CAG GCA GGA GAA TAC CTA CTC CAT ATT CAG AGC GAA CGC GCC AAC
 Thr Gln Ala Gly Glu Tyr Leu Leu His Ile Gln Ser Glu Arg Ala Asn
 110 115 120 125

TAC ACA GTA CTG TTC ACA GTG AAT GTA AGA GAT ACA CAG CTG TAT GTG
 Tyr Thr Val Leu Phe Thr Val Asn Val Arg Asp Thr Gln Leu Tyr Val
 130 135 140

CTA AGG AGA CCT TAC TTT AGG AAG ATG GAA AAC CAG GAT GCA CTG CTC
 Leu Arg Arg Pro Tyr Phe Arg Lys Met Glu Asn Gln Asp Ala Leu Leu
 145 150 155

Fig. 1a.2

TGC ATC TCC GAG GGT GTT CCG GAG CCC ACT GTG GAG TGG GTG CTC TGC
 Cys Ile Ser Glu Gly Val Pro Glu Pro Thr Val Glu Trp Val Leu Cys
 160 165 170

AGC TCC CAC AGG GAA AGC TGT AAA GAA GAA GGC CCT GCT GTT GTC AGA
 Ser Ser His Arg Glu Ser Cys Lys Glu Glu Gly Pro Ala Val Val Arg
 175 180 185

AAG GAG GAA AAG GTA CTT CAT GAG TTG TTC GGA ACA GAC ATC AGA TGC
 Lys Glu Glu Lys Val Leu His Glu Leu Phe Gly Thr Asp Ile Arg Cys
 190 195 200 205

TGT GCT AGA AAT GCA CTG GGC CGC GAA TGC ACC AAG CTG TTC ACC ATA
 Cys Ala Arg Asn Ala Leu Gly Arg Glu Cys Thr Lys Leu Phe Thr Ile
 210 215 220

GAT CTA AAC CAG GCT CCT CAG AGC ACA CTG CCC CAG TTA TTC CTG AAA
 Asp Leu Asn Gln Ala Pro Gln Ser Thr Leu Pro Gln Leu Phe Leu Lys
 225 230 235

GTG GGG GAA CCC TTG TGG ATC AGG TGT AAG GCC ATC CAT GTG AAC CAT
 Val Gly Glu Pro Leu Trp Ile Arg Cys Lys Ala Ile His Val Asn His
 240 245 250

GGA TTC GGG CTC ACC TGG GAG CTG GAA GAC AAA GCC CTG GAG GAG GGC
 Gly Phe Gly Leu Thr Trp Glu Leu Glu Asp Lys Ala Leu Glu Glu Gly
 255 260 265

AGC TAC TTT GAG ATG AGT ACC TAC TCC ACA AAC AGG ACC ATG ATT CGG
 Ser Tyr Phe Glu Met Ser Thr Tyr Ser Thr Asn Arg Thr Met Ile Arg
 270 275 280 285

ATT CTC TTG GCC TTT GTG TCT TCC GTG GGA AGG AAC GAC ACC GGA TAT
 Ile Leu Leu Ala Phe Val Ser Ser Val Gly Arg Asn Asp Thr Gly Tyr
 290 295 300

TAC ACC TGC TCT TCC TCA AAG CAC CCC AGC CAG TCA GCG TTG GTG ACC
 Tyr Thr Cys Ser Ser Lys His Pro Ser Gln Ser Ala Leu Val Thr
 305 310 315

ATC CTA GAA AAA GGG TTT ATA AAC GCT ACC AGC TCG CAA GAA GAG TAT
 Ile Leu Glu Lys Gly Phe Ile Asn Ala Thr Ser Ser Gln Glu Glu Tyr
 320 325 330

GAA ATT GAC CCG TAC GAA AAG TTC TGC TTC TCA GTC AGG TTT AAA GCG
 Glu Ile Asp Pro Tyr Glu Lys Phe Cys Phe Ser Val Arg Phe Lys Ala
 335 340 345

Fig. 1a.3

TAC CCA CGA ATC CGA TGC ACG TGG ATC TTC TCT CAA GCC TCA TTT CCT
 Tyr Pro Arg Ile Arg Cys Thr Trp Ile Phe Ser Gln Ala Ser Phe Pro
 350 355 360 365

 TGT GAA CAG AGA GGC CTG GAG GAT GGG TAC AGC ATA TCT AAA TTT TGC
 Cys Glu Gln Arg Gly Leu Glu Asp Gly Tyr Ser Ile Ser Lys Phe Cys
 370 375 380

 GAT CAT AAG AAC AAG CCA GGA GAG TAC ATA TTC TAT GCA GAA AAT GAT
 Asp His Lys Asn Lys Pro Gly Glu Tyr Ile Phe Tyr Ala Glu Asn Asp
 385 390 395

 GAC GCC CAG TTC ACC AAA ATG TTC ACG CTG AAT ATA AGA AAG AAA CCT
 Asp Ala Gln Phe Thr Lys Met Phe Thr Leu Asn Ile Arg Lys Lys Pro
 400 405 410

 CAA GTG CTA GCA AAT GCC TCA GCC AGC CAG GCG TCC TGT TCC TCT GAT
 Gln Val Leu Ala Asn Ala Ser Ala Ser Gln Ala Ser Cys Ser Ser Asp
 415 420 425

 GGC TAC CCG CTA CCC TCT TGG ACC TGG AAG AAG TGT TCG GAC AAA TCT
 Gly Tyr Pro Leu Pro Ser Trp Thr Trp Lys Lys Cys Ser Asp Lys Ser
 430 435 440 445

 CCC AAT TGC ACG GAG GAA ATC CCA GAA GGA GTT TGG AAT AAA AAG GCT
 Pro Asn Cys Thr Glu Glu Ile Pro Glu Gly Val Trp Asn Lys Lys Ala
 450 455 460

 AAC AGA AAA GTG TTT GGC CAG TGG GTG TCG AGC AGT ACT CTA AAT ATG
 Asn Arg Lys Val Phe Gly Gln Trp Val Ser Ser Ser Thr Leu Asn Met
 465 470 475

 AGT GAG GCC GGG AAA GGG CTT CTG GTC AAA TGC TGT GCG TAC AAT TCT
 Ser Glu Ala Gly Lys Gly Leu Leu Val Lys Cys Cys Ala Tyr Asn Ser
 480 485 490

 ATG GGC ACG TCT TGC GAA ACC ATC TTT TTA AAC TCA CCA GGC CCC TTC
 Met Gly Thr Ser Cys Glu Thr Ile Phe Leu Asn Ser Pro Gly Pro Phe
 495 500 505

 CCT TTC ATC CAA GAC AAC ATC TCC TTC TAT GCG ACC ATT GGG CTC TGT
 Pro Phe Ile Gln Asp Asn Ile Ser Phe Tyr Ala Thr Ile Gly Leu Cys
 510 515 520 525

 CTC CCC TTC ATT GTT CTC ATT GTG TTG ATC TGC CAC AAA TAC AAA
 Leu Pro Phe Ile Val Val Leu Ile Val Leu Ile Cys His Lys Tyr Lys
 530 535 540

Fig. 1a.4

AAG CAA TTT AGG TAC GAG AGT CAG CTG CAG ATG ATC CAG GTG ACT GGC
 Lys Gln Phe Arg Tyr Glu Ser Gln Leu Gln Met Ile Gln Val Thr Gly
 545 550 555

CCC CTG GAT AAC GAG TAC TTC TAC GTT GAC TTC AGG GAC TAT GAA TAT
 Pro Leu Asp Asn Glu Tyr Phe Tyr Val Asp Phe Arg Asp Tyr Glu Tyr
 560 565 570

GAC CTT AAG TGG GAG TTC CCG AGA GAG AAC TTA GAG TTT GGG AAG GTC
 Asp Leu Lys Trp Glu Phe Pro Arg Glu Asn Leu Glu Phe Gly Lys Val
 575 580 585

CTG GGG TCT GGC GCT TTC GGG AGG GTG ATG AAC GCC ACG GCC TAT GGC
 Leu Gly Ser Gly Ala Phe Gly Arg Val Met Asn Ala Thr Ala Tyr Gly
 590 595 600 605

ATT AGT AAA ACG GGA GTC TCA ATT CAG GTG GCG GTG AAG ATG CTA AAA
 Ile Ser Lys Thr Gly Val Ser Ile Gln Val Ala Val Lys Met Leu Lys
 610 615 620

GAG AAA GCT GAC AGC TGT GAA AAA GAA GCT CTC ATG TCG GAG CTC AAA
 Glu Lys Ala Asp Ser Cys Glu Lys Glu Ala Leu Met Ser Glu Leu Lys
 625 630 635

ATG ATG ACC CAC CTG GGA CAC CAT GAC AAC ATC GTG AAT CTG CTG GGG
 Met Met Thr His Leu Gly His His Asp Asn Ile Val Asn Leu Leu Gly
 640 645 650

GCA TGC ACA CTG TCA GGG CCA GTG TAC TTG ATT TTT GAA TAT TGT TGC
 Ala Cys Thr Leu Ser Gly Pro Val Tyr Leu Ile Phe Glu Tyr Cys Cys
 655 660 665

TAT GGT GAC CTC CTC AAC TAC CTA AGA AGT AAA AGA GAG AAG TTT CAC
 Tyr Gly Asp Leu Leu Asn Tyr Leu Arg Ser Lys Arg Glu Lys Phe His
 670 675 680 685

AGG ACA TGG ACA GAG ATT TTT AAG GAA CAT AAT TTC AGT TCT TAC CCT
 Arg Thr Trp Thr Glu Ile Phe Lys Glu His Asn Phe Ser Ser Tyr Pro
 690 695 700

ACT TTC CAG GCA CAT TCA AAT TCC AGC ATG CCT GGT TCA CGA GAA GTT
 Thr Phe Gln Ala His Ser Asn Ser Ser Met Pro Gly Ser Arg Glu Val
 705 710 715

CAG TTA CAC CCG CCC TTG GAT CAG CTC TCA GGG TTC AAT GGG AAT TCA
 Gln Leu His Pro Pro Leu Asp Gln Leu Ser Gly Phe Asn Gly Asn Ser
 720 725 730

Fig. 1a.5

ATT	CAT	TCT	GAA	GAT	GAG	ATT	GAA	TAT	GAA	AAC	CAG	AAG	AGG	CTG	GCA
Ile	His	Ser	Glu	Asp	Glu	Ile	Glu	Tyr	Glu	Asn	Gln	Lys	Arg	Leu	Ala
735						740						745			
GAA	GAA	GAG	GAG	GAA	GAT	TTG	AAC	GTG	CTG	ACG	TTT	GAA	GAC	CTC	CTT
Glu	Glu	Glu	Glu	Glu	Asp	Leu	Asn	Val	Leu	Thr	Phe	Glu	Asp	Leu	Leu
750						755					760				765
TGC	TTT	GCG	TAC	CAA	GTG	GCC	AAA	GGC	ATG	GAA	TTC	CTG	GAG	TTC	AAG
Cys	Phe	Ala	Tyr	Gln	Val	Ala	Lys	Gly	Met	Glu	Phe	Leu	Glu	Phe	Lys
						770				775				780	
TCG	TGT	GTC	CAC	AGA	GAC	CTG	GCA	GCC	AGG	AAT	GTG	TTG	GTC	ACC	CAC
Ser	Cys	Val	His	Arg	Asp	Leu	Ala	Ala	Arg	Asn	Val	Leu	Val	Thr	His
						785				790				795	
GGG	AAG	GTG	GTG	AAG	ATC	TGT	GAC	TTT	GGA	CTG	GCC	CGA	GAC	ATC	CTG
Gly	Lys	Val	Val	Lys	Ile	Cys	Asp	Phe	Gly	Leu	Ala	Arg	Asp	Ile	Leu
						800			805				810		
AGC	GAC	TCC	AGC	TAC	GTC	GTC	AGG	GGC	AAC	GCA	CGG	CTG	CCG	GTG	AAG
Ser	Asp	Ser	Ser	Tyr	Val	Val	Arg	Gly	Asn	Ala	Arg	Leu	Pro	Val	Lys
						815			820				825		
TGG	ATG	GCA	CCC	GAG	AGC	TTA	TTT	GAA	GGG	ATC	TAC	ACA	ATC	AAG	AGT
Trp	Met	Ala	Pro	Glu	Ser	Leu	Phe	Glu	Gly	Ile	Tyr	Thr	Ile	Lys	Ser
						830			835			840			845
GAC	GTC	TGG	TCC	TAC	GGC	ATC	CTT	CTC	TGG	GAG	ATA	TTT	TCA	CTG	GGT
Asp	Val	Trp	Ser	Tyr	Gly	Ile	Leu	Leu	Trp	Glu	Ile	Phe	Ser	Leu	Gly
						850			855				860		
GTG	AAC	CCT	TAC	CCT	GGC	ATT	CCT	GTC	GAC	GCT	AAC	TTC	TAT	AAA	CTG
Val	Asn	Pro	Tyr	Pro	Gly	Ile	Pro	Val	Asp	Ala	Asn	Phe	Tyr	Lys	Leu
						865			870				875		
ATT	CAG	AGT	GGA	TTT	AAA	ATG	GAG	CAG	CCA	TTC	TAT	GCC	ACA	GAA	GGG
Ile	Gln	Ser	Gly	Phe	Lys	Met	Glu	Gln	Pro	Phe	Tyr	Ala	Thr	Glu	Gly
						880			885				890		
ATA	TAC	TTT	GTA	ATG	CAA	TCC	TGC	TGG	GCT	TTT	GAC	TCA	AGG	AAG	CGG
Ile	Tyr	Phe	Val	Met	Gln	Ser	Cys	Trp	Ala	Phe	Asp	Ser	Arg	Lys	Arg
						895			900				905		

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Fig. 1a.6

CCA TCC TTC CCC AAC CTG ACT TCA TTT TTA GGA TGT CAG CTG GCA GAG
Pro Ser Phe Pro Asn Leu Thr Ser Phe Leu Gly Cys Gln Leu Ala Glu
910 915 920 925

GCA GAA GAA GCA TGT ATC AGA ACA TCC ATC CAT CTA CCA AAA CAG GCG
Ala Glu Glu Ala Cys Ile Arg Thr Ser Ile His Leu Pro Lys Gln Ala
930 935 940

GCC CCT CAG CAG AGA GGC GGG CTC AGA GCC CAG TCG CCA CAG CGC CAG
Ala Pro Gln Gln Arg Gly Gly Leu Arg Ala Gln Ser Pro Gln Arg Gln
945 950 955

GTG AAG ATT CAC AGA GAA AGA AGT TAGCGAGGAG GCCTTGGACC CCGCCACCC
Val Lys Ile His Arg Glu Arg Ser
960 965

AGCAGGCTGT AGACCGCAGA GCCAAGATTA GCCTCGCCTC TGAGGAAGCG CCCTACAGCG
CGTTGCTTCG CTGGACTTTT CTCTAGATGC TGTCTGCCAT TACTCCAAAG TGACTTCTAT

AAAATCAAAC CTCTCCTCGC ACAGGCGGGA GAGCCAATAA TGAGACTTGT TGGTGAGCCC

GCCTACCCCTG GGGGCCTTTC CACGAGCTTG AGGGGAAAGC CATGTATCTG AAATATAGTA

TATTCTTGTA AATACGTGAA ACAAAACAAA CCCGTTTTT GCTAAGGGAA AGCTAAATAT

GATTTTAAA AATCTATGTT TTAAAATACT ATGTAACCTT TTCATCTATT TAGTGATATA

TTTTATGGAT GGAAATAAAC TTTCTACTGT AAAAAAAA AAAAAAAA AAAAAAA

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Fig. 1b.1

CGAGGCGGCA TCCGAGGGCT GGGCCGGCGC CCTGGGGGAC CCCGGGCTCC GGAGGCC

 ATG CCG GCG TTG GCG CGC GAC GCG GGC ACC GTG CCG CTG CTC GTT GTT
 Met Pro Ala Leu Ala Arg Asp Ala Gly Thr Val Pro Leu Leu Val Val
 -27 -25 -20 -15

 TTT TCT GCA ATG ATA TTT GGG ACT ATT ACA AAT CAA GAT CTG CCT GTG
 Phe Ser Ala Met Ile Phe Gly Thr Ile Thr Asn Gln Asp Leu Pro Val
 -10 -5 1 5

 ATC AAG TGT GTT TTA ATC AAT CAT AAG AAC AAT GAT TCA TCA GTG GGG
 Ile Lys Cys Val Leu Ile Asn His Lys Asn Asn Asp Ser Ser Val Gly
 10 15 20

 AAG TCA TCA TCA TAT CCC ATG GTA TCA GAA TCC CCG GAA GAC CTC GGG
 Lys Ser Ser Ser Tyr Pro Met Val Ser Glu Ser Pro Glu Asp Leu Gly
 25 30 35

 TGT GCG TTG AGA CCC CAG AGC TCA GGG ACA GTG TAC GAA GCT GCC GCT
 Cys Ala Leu Arg Pro Gln Ser Ser Gly Thr Val Tyr Glu Ala Ala Ala
 40 45 50

 GTG GAA GTG GAT GTA TCT GCT TCC ATC ACA CTG CAA GTG CTG GTC GAT
 Val Glu Val Asp Val Ser Ala Ser Ile Thr Leu Gln Val Leu Val Asp
 55 60 65

 GCC CCA GGG AAC ATT TCC TGT CTC TGG GTC TTT AAG CAC AGC TCC CTG
 Ala Pro Gly Asn Ile Ser Cys Leu Trp Val Phe Lys His Ser Ser Leu
 70 75 80 85

 AAT TGC CAG CCA CAT TTT GAT TTA CAA AAC AGA GGA GTT GTT TCC ATG
 Asn Cys Gln Pro His Phe Asp Leu Gln Asn Arg Gly Val Val Ser Met
 90 95 100

 GTC ATT TTG AAA ATG ACA GAA ACC CAA GCT GGA GAA TAC CTA CTT TTT
 Val Ile Leu Lys Met Thr Glu Thr Gln Ala Gly Glu Tyr Leu Leu Phe
 105 110 115

 ATT CAG AGT GAA GCT ACC AAT TAC ACA ATA TTG TTT ACA GTG AGT ATA
 Ile Gln Ser Glu Ala Thr Asn Tyr Thr Ile Leu Phe Thr Val Ser Ile
 120 125 130

 AGA AAT ACC CTG CTT TAC ACA TTA AGA AGA CCT TAC TTT AGA AAA ATG
 Arg Asn Thr Leu Leu Tyr Thr Leu Arg Arg Pro Tyr Phe Arg Lys Met
 135 140 145

Fig. 1b.2

GAA AAC CAG GAC GCC CTG GTC TGC ATA TCT GAG AGC GTT CCA GAG CCG
 Glu Asn Gln Asp Ala Leu Val Cys Ile Ser Glu Ser Val Pro Glu Pro
 150 155 160 165

ATC GTG GAA TGG GTG CTT TGC GAT TCA CAG GGG GAA AGC TGT AAA GAA
 Ile Val Glu Trp Val Leu Cys Asp Ser Gln Gly Glu Ser Cys Lys Glu
 170 175 180

GAA AGT CCA GCT GTT AAA AAG GAG GAA AAA GTG CTT CAT GAA TTA
 Glu Ser Pro Ala Val Val Lys Lys Glu Glu Lys Val Leu His Glu Leu
 185 190 195

TTT GGG ACG GAC ATA AGG TGC TGT GCC AGA AAT GAA CTG GGC AGG GAA
 Phe Gly Thr Asp Ile Arg Cys Cys Ala Arg Asn Glu Leu Gly Arg Glu
 200 205 210

TGC ACC AGG CTG TTC ACA ATA GAT CTA AAT CAA ACT CCT CAG ACC ACA
 Cys Thr Arg Leu Phe Thr Ile Asp Leu Asn Gln Thr Pro Gln Thr Thr
 215 220 225

TTG CCA CAA TTA TTT CTT AAA GTA GGG GAA CCC TTA TGG ATA AGG TGC
 Leu Pro Gln Leu Phe Leu Lys Val Gly Glu Pro Leu Trp Ile Arg Cys
 230 235 240 245

AAA GCT GTT CAT GTG AAC CAT GGA TTC GGG CTC ACC TGG GAA TTA GAA
 Lys Ala Val His Val Asn His Gly Phe Gly Leu Thr Trp Glu Leu Glu
 250 255 260

AAC AAA GCA CTC GAG GAG GGC AAC TAC TTT GAG ATG AGT ACC TAT TCA
 Asn Lys Ala Leu Glu Glu Asn Tyr Phe Glu Met Ser Thr Tyr Ser
 265 270 275

ACA AAC AGA ACT ATG ATA CGG ATT CTG TTT GCT TTT GTA TCA TCA GTG
 Thr Asn Arg Thr Met Ile Arg Ile Leu Phe Ala Phe Val Ser Ser Val
 280 285 290

GCA AGA AAC GAC ACC GGA TAC TAC ACT TGT TCC TCT TCA AAG CAT CCC
 Ala Arg Asn Asp Thr Gly Tyr Tyr Thr Cys Ser Ser Ser Lys His Pro
 295 300 305

AGT CAA TCA GCT TTG GTT ACC ATC GTA GGA AAG GGA TTT ATA AAT GCT
 Ser Gln Ser Ala Leu Val Thr Ile Val Gly Lys Gly Phe Ile Asn Ala
 310 315 320 325

ACC AAT TCA AGT GAA GAT TAT GAA ATT GAC CAA TAT GAA GAG TTT TGT
 Thr Asn Ser Ser Glu Asp Tyr Glu Ile Asp Gln Tyr Glu Glu Phe Cys
 330 335 340

Fig. 1b.3

TTT TCT GTC AGG TTT AAA GCC TAC CCA CAA ATC AGA TGT ACG TGG ACC
 Phe Ser Val Arg Phe Lys Ala Tyr Pro Gln Ile Arg Cys Thr Trp Thr
 345 350 355

TTC TCT CGA AAA TCA TTT CCT TGT GAG CAA AAG GGT CTT GAT AAC GGA
 Phe Ser Arg Lys Ser Phe Pro Cys Glu Gln Lys Gly Leu Asp Asn Gly
 360 365 370

TAC AGC ATA TCC AAG TTT TGC AAT CAT AAG CAC CAG CCA GGA GAA TAT
 Tyr Ser Ile Ser Lys Phe Cys Asn His Lys His Gln Pro Gly Glu Tyr
 375 380 385

ATA TTC CAT GCA GAA AAT GAT GAT GCC CAA TTT ACC AAA ATG TTC ACG
 Ile Phe His Ala Glu Asn Asp Ala Gln Phe Thr Lys Met Phe Thr
 390 395 400 405

CTG AAT ATA AGA AGG AAA CCT CAA GTG CTC GCA GAA GCA TCG GCA AGT
 Leu Asn Ile Arg Arg Lys Pro Gln Val Leu Ala Glu Ala Ser Ala Ser
 410 415 420

CAG GCG TCC TGT TTC TCG GAT GGA TAC CCA TTA CCA TCT TGG ACC TGG
 Gln Ala Ser Cys Phe Ser Asp Gly Tyr Pro Leu Pro Ser Trp Thr Trp
 425 430 435

AAG AAG TGT TCA GAC AAG TCT CCC AAC TGC ACA GAA GAG ATC ACA GAA
 Lys Lys Cys Ser Asp Lys Ser Pro Asn Cys Thr Glu Glu Ile Thr Glu
 440 445 450

GGA GTC TGG AAT AGA AAG GCT AAC AGA AAA GTG TTT GGA CAG TGG GTG
 Gly Val Trp Asn Arg Lys Ala Asn Arg Lys Val Phe Gly Gln Trp Val
 455 460 465

TCG AGC AGT ACT CTA AAC ATG AGT GAA GCC ATA AAA GGG TTC CTG GTC
 Ser Ser Ser Thr Leu Asn Met Ser Glu Ala Ile Lys Gly Phe Leu Val
 470 475 480 485

AAG TGC TGT GCA TAC AAT TCC CTT GGC ACA TCT TGT GAG ACG ATC CTT
 Lys Cys Cys Ala Tyr Asn Ser Leu Gly Thr Ser Cys Glu Thr Ile Leu
 490 495 500

TTA AAC TCT CCA GGC CCC TTC CCT TTC ATC CAA GAC AAC ATC TCA TTC
 Leu Asn Ser Pro Gly Pro Phe Pro Phe Ile Gln Asp Asn Ile Ser Phe
 505 510 515

TAT GCA ACA ATT GGT GTT TGT CTC CTC TTC ATT GTC GTT TTA ACC CTG
 Tyr Ala Thr Ile Gly Val Cys Leu Leu Phe Ile Val Val Leu Thr Leu
 520 525 530

Fig. 1b.4

CTA ATT TGT CAC AAG TAC AAA AAG CAA TTT AGG TAT GAA AGC CAG CTA
 Leu Ile Cys His Lys Tyr Lys Lys Gln Phe Arg Tyr Glu Ser Gln Leu
 535 540 545

CAG ATG GTA CAG GTG ACC GGC TCC TCA GAT AAT GAG TAC TTC TAC GTT
 Gln Met Val Gln Val Thr Gly Ser Ser Asp Asn Glu Tyr Phe Tyr Val
 550 555 560 565

GAT TTC AGA GAA TAT GAA TAT GAT CTC AAA TGG GAG TTT CCA AGA GAA
 Asp Phe Arg Glu Tyr Glu Tyr Asp Leu Lys Trp Glu Phe Pro Arg Glu
 570 575 580

AAT TTA GAG TTT GGG AAG GTA CTA GGA TCA GGT GCT TTT GGA AAA GTG
 Asn Leu Glu Phe Gly Lys Val Leu Gly Ser Gly Ala Phe Gly Lys Val
 585 590 595

ATG AAC GCA ACA GCT TAT GGA ATT AGC AAA ACA GGA GTC TCA ATC CAG
 Met Asn Ala Thr Ala Tyr Gly Ile Ser Lys Thr Gly Val Ser Ile Gln
 600 605 610

GTT GCC GTC AAA ATG CTG AAA GAA AAA GCA GAC AGC TCT GAA AGA GAG
 Val Ala Val Lys Met Leu Lys Glu Lys Ala Asp Ser Ser Glu Arg Glu
 615 620 625

GCA CTC ATG TCA GAA CTC AAG ATG ATG ACC CAG CTG GGA AGC CAC GAG
 Ala Leu Met Ser Glu Leu Lys Met Met Thr Gln Leu Gly Ser His Glu
 630 635 640 645

AAT ATT GTG AAC CTG CTG GGG GCG TGC ACA CTG TCA GGA CCA ATT TAC
 Asn Ile Val Asn Leu Leu Gly Ala Cys Thr Leu Ser Gly Pro Ile Tyr
 650 655 660

TTG ATT TTT GAA TAC TGT TGC TAT GGT GAT CTT CTC AAC TAT CTA AGA
 Leu Ile Phe Glu Tyr Cys Cys Tyr Gly Asp Leu Leu Asn Tyr Leu Arg
 665 670 675

AGT AAA AGA GAA AAA TTT CAC AGG ACT TGG ACA GAG ATT TTC AAG GAA
 Ser Lys Arg Glu Lys Phe His Arg Thr Trp Thr Glu Ile Phe Lys Glu
 680 685 690

CAC AAT TTC AGT TTT TAC CCC ACT TTC CAA TCA CAT CCA AAT TCC AGC
 His Asn Phe Ser Phe Tyr Pro Thr Phe Gln Ser His Pro Asn Ser Ser
 695 700 705

ATG CCT GGT TCA AGA GAA GTT CAG ATA CAC CCG GAC TCG GAT CAA ATC
 Met Pro Gly Ser Arg Glu Val Gln Ile His Pro Asp Ser Asp Gln Ile
 710 715 720 725

Fig. 1b.5

TCA GGG CTT CAT GGG AAT TCA TTT CAC TCT GAA GAT GAA ATT GAA TAT
 Ser Gly Leu His Gly Asn Ser Phe His Ser Glu Asp Glu Ile Glu Tyr
 730 735 740

GAA AAC CAA AAA AGG CTG GAA GAA GAG GAG GAC TTG AAT GTG CTT ACA
 Glu Asn Gln Lys Arg Leu Glu Glu Glu Asp Leu Asn Val Leu Thr
 745 750 755

TTT GAA GAT CTT CTT TGC TTT GCA TAT CAA GTT GCC AAA GGA ATG GAA
 Phe Glu Asp Leu Leu Cys Phe Ala Tyr Gln Val Ala Lys Gly Met Glu
 760 765 770

TTT CTG GAA TTT AAG TCG TGT GTT CAC AGA GAC CTG GCC GCC AGG AAC
 Phe Leu Glu Phe Lys Ser Cys Val His Arg Asp Leu Ala Ala Arg Asn
 775 780 785

GTG CTT GTC ACC CAC GGG AAA GTG GTG AAG ATA TGT GAC TTT GGA TTG
 Val Leu Val Thr His Gly Lys Val Val Lys Ile Cys Asp Phe Gly Leu
 790 795 800 805

GCT CGA GAT ATC ATG AGT GAT TCC AAC TAT GTT GTC AGG GGC AAT GCC
 Ala Arg Asp Ile Met Ser Asp Ser Asn Tyr Val Val Arg Gly Asn Ala
 810 815 820

CGT CTG CCT GTA AAA TGG ATG GCC CCC GAA AGC CTG TTT GAA GGC ATC
 Arg Leu Pro Val Lys Trp Met Ala Pro Glu Ser Leu Phe Glu Gly Ile
 825 830 835

TAC ACC ATT AAG AGT GAT GTC TGG TCA TAT GGA ATA TTA CTG TGG GAA
 Tyr Thr Ile Lys Ser Asp Val Trp Ser Tyr Gly Ile Leu Leu Trp Glu
 840 845 850

ATC TTC TCA CTT GGT GTG AAT CCT TAC CCT GGC ATT CCG GTT GAT GCT
 Ile Phe Ser Leu Gly Val Asn Pro Tyr Pro Gly Ile Pro Val Asp Ala
 855 860 865

AAC TTC TAC AAA CTG ATT CAA AAT GGA TTT AAA ATG GAT CAG CCA TTT
 Asn Phe Tyr Lys Leu Ile Gln Asn Gly Phe Lys Met Asp Gln Pro Phe
 870 875 880 885

TAT GCT ACA GAA GAA ATA TAC ATT ATA ATG CAA TCC TGC TGG GCT TTT
 Tyr Ala Thr Glu Glu Ile Tyr Ile Ile Met Gln Ser Cys Trp Ala Phe
 890 895 900

GAC TCA AGG AAA CGG CCA TCC TTC CCT AAT TTG ACT TCG TTT TTA GGA
 Asp Ser Arg Lys Arg Pro Ser Phe Pro Asn Leu Thr Ser Phe Leu Gly
 905 910 915

Fig. 1b.6

TGT CAG CTG GCA GAT GCA GAA GAA GCG ATG TAT CAG AAT GTG GAT GGC
Cys Gln Leu Ala Asp Ala Glu Glu Ala Met Tyr Gln Asn Val Asp Gly
920 925 930

CGT GTT TCG GAA TGT CCT CAC ACC TAC CAA AAC AGG CGA CCT TTC AGC
Arg Val Ser Glu Cys Pro His Thr Tyr Gln Asn Arg Arg Pro Phe Ser
935 940 945

AGA GAG ATG GAT TTG GGG CTA CTC TCT CCG CAG GCT CAG GTC GAA GAT
Arg Glu Met Asp Leu Gly Leu Leu Ser Pro Gln Ala Gln Val Glu Asp
950 955 960 965

TCG TAGAGGAACA ATTTAGTTTT AAGGACTTCA TCCCTCCACC TATCCCTAAC
Ser

AGGCTGTAGA TTACCAAAAC AAGATTAATT TCATCACTAA AAGAAAATCT ATTATCAACT

GCTGCTTCAC CAGACTTTTC TCTAGAAGCC GTCTGCGTTT ACTCTTGT TT TCAAAGGGAC

TTTTGTAAAA TCAAATCATC CTGTCACAAG GCAGGAGGAG CTGATAATGA ACTTTATTGG

AGCATTGATC TGCATCCAAG GCCTTCTCAG GCCGGCTTGA GTGAATTGTG TACCTGAAGT

ACAGTATATT CTTGTAAATA CATAAAACAA AAGCATTG CTAAGGAGAA GCTAATATGA

TTTTTTAAGT CTATGTTTA AAATAATATG TAAATTTTC AGCTATTTAG TGATATATT

TATGGGTGGG AATAAAATTT CTACTACAGA AAAAAAAA AAAAAAAA AAAAA

Fig. 2.1

CTGTGTCCCG CAGCCGGATA ACCTGGCTGA CCCGATTCCG CGGACACCCG TGCAGCCGCG
 GCTGGAGCCA GGGCGCCGGT GCCCGCGCTC TCCCCGGTCT TGCGCTGCAG GGGCCGATAC
 CGCCTCTGTG ACTTCTTGC GGGCCAGGGA CGGAGAAGGA GTCTGTGCCT GAGAAACTGG
 GCTCTGTGCC CAGGCGCGAG GTGCAGG ATG GAG AGC AAG GGC CTG CTA GCT
 Met Glu Ser Lys Gly Leu Leu Ala -19 -15
 GTC GCT CTG TGG TTC TGC GTG GAG ACC CGA GCC GCC TCT GTG GGT TTG
 Val Ala Leu Trp Phe Cys Val Glu Thr Arg Ala Ala Ser Val Gly Leu
 -10 -5 1 5
 CCT GGC GAT TTT CTC CAT CCC CCC AAG CTC AGC ACA CAG AAA GAC ATA
 Pro Gly Asp Phe Leu His Pro Pro Lys Leu Ser Thr Gln Lys Asp Ile
 10 15 20
 CTG ACA ATT TTG GCA AAT ACA ACC CTT CAG ATT ACT TGC AGG GGA CAG
 Leu Thr Ile Leu Ala Asn Thr Thr Leu Gln Ile Thr Cys Arg Gly Gln
 25 30 35
 CGG GAC CTG GAC TGG CTT TGG CCC AAT GCT CAG CGT GAT TCT GAG GAA
 Arg Asp Leu Asp Trp Leu Trp Pro Asn Ala Gln Arg Asp Ser Glu Glu
 40 45 50
 AGG GTA TTG GTG ACT GAA TGC GGC GGT GGT GAC AGT ATC TTC TGC AAA
 Arg Val Leu Val Thr Glu Cys Gly Gly Asp Ser Ile Phe Cys Lys
 55 60 65
 ACA CTC ACC ATT CCC AGG GTG GTT GGA AAT GAT ACT GGA GCC TAC AAG
 Thr Leu Thr Ile Pro Arg Val Val Gly Asn Asp Thr Gly Ala Tyr Lys
 70 75 80 85
 TGC TCG TAC CGG GAC GTC GAC ATA GCC TCC ACT GTT TAT GTC TAT GTT
 Cys Ser Tyr Arg Asp Val Asp Ile Ala Ser Thr Val Tyr Val Tyr Val
 90 95 100
 CGA GAT TAC AGA TCA CCA TTC ATC GCC TCT GTC AGT GAC CAG CAT GGC
 Arg Asp Tyr Arg Ser Pro Phe Ile Ala Ser Val Ser Asp Gln His Gly
 105 110 115
 ATC GTG TAC ATC ACC GAG AAC AAG AAC AAA ACT GTG GTG ATC CCC TGC
 Ile Val Tyr Ile Thr Glu Asn Lys Asn Lys Thr Val Val Ile Pro Cys
 120 125 130

Fig. 2.2

CGA GGG TCG ATT TCA AAC CTC AAT GTG TCT CTT TGC GCT AGG TAT CCA
 Arg Gly Ser Ile Ser Asn Leu Asn Val Ser Leu Cys Ala Arg Tyr Pro
 135 140 145

 GAA AAG AGA TTT GTT CCG GAT GGA AAC AGA ATT TCC TGG GAC AGC GAG
 Glu Lys Arg Phe Val Pro Asp Gly Asn Arg Ile Ser Trp Asp Ser Glu
 150 155 160 165

 ATA GGC TTT ACT CTC CCC AGT TAC ATG ATC AGC TAT GCC GGC ATG GTC
 Ile Gly Phe Thr Leu Pro Ser Tyr Met Ile Ser Tyr Ala Gly Met Val
 170 175 180

 TTC TGT GAG GCA AAG ATC AAT GAT GAA ACC TAT CAG TCT ATC ATG TAC
 Phe Cys Glu Ala Lys Ile Asn Asp Glu Thr Tyr Gln Ser Ile Met Tyr
 185 190 195

 ATA GTT GTG GTT GTA GGA TAT AGG ATT TAT GAT GTG ATT CTG AGC CCC
 Ile Val Val Val Val Gly Tyr Arg Ile Tyr Asp Val Ile Leu Ser Pro
 200 205 210

 CCG CAT GAA ATT GAG CTA TCT GCC GGA GAA AAA CTT GTC TTA AAT TGT
 Pro His Glu Ile Glu Leu Ser Ala Gly Glu Lys Leu Val Leu Asn Cys
 215 220 225

 ACA GCG AGA ACA GAG CTC AAT GTG GGG CTT GAT TTC ACC TGG CAC TCT
 Thr Ala Arg Thr Glu Leu Asn Val Gly Leu Asp Phe Thr Trp His Ser
 230 235 240 245

 CCA CCT TCA AAG TCT CAT CAT AAG AAG ATT GTA AAC CGG GAT GTG AAA
 Pro Pro Ser Lys Ser His His Lys Lys Ile Val Asn Arg Asp Val Lys
 250 255 260

 CCC TTT CCT GGG ACT GTG GCG AAG ATG TTT TTG AGC ACC TTG ACA ATA
 Pro Phe Pro Gly Thr Val Ala Lys Met Phe Leu Ser Thr Leu Thr Ile
 265 270 275

 GAA AGT GTG ACC AAG AGT GAC CAA GGG GAA TAC ACC TGT GTA GCG TCC
 Glu Ser Val Thr Lys Ser Asp Gln Gly Glu Tyr Thr Cys Val Ala Ser
 280 285 290

 AGT GGA CGG ATG ATC AAG AGA AAT AGA ACA TTT GTC CGA GTT CAC ACA
 Ser Gly Arg Met Ile Lys Arg Asn Arg Thr Phe Val Arg Val His Thr
 295 300 305

 AAG CCT TTT ATT GCT TTC GGT AGT GGG ATG AAA TCT TTG GTG GAA GCC
 Lys Pro Phe Ile Ala Phe Gly Ser Gly Met Lys Ser Leu Val Glu Ala
 310 315 320 325

Fig. 2.3

ACA GTG GGC AGT CAA GTC CGA ATC CCT GTG AAG TAT CTC AGT TAC CCA
Thr Val Gly Ser Gln Val Arg Ile Pro Val Lys Tyr Leu Ser Tyr Pro
330 335 340

GCT CCT GAT ATC AAA TGG TAC AGA AAT GGA AGG CCC ATT GAG TCC AAC
Ala Pro Asp Ile Lys Trp Tyr Arg Asn Gly Arg Pro Ile Glu Ser Asn
345 350 355

TAC ACA ATG ATT GTT GGC GAT GAA CTC ACC ATC ATG GAA GTG ACT GAA
Tyr Thr Met Ile Val Gly Asp Glu Leu Thr Ile Met Glu Val Thr Glu
360 365 370

AGA GAT GCA GGA AAC TAC ACG GTC ATC CTC ACC AAC CCC ATT TCA ATG
Arg Asp Ala Gly Asn Tyr Thr Val Ile Leu Thr Asn Pro Ile Ser Met
375 380 385

GAG AAA CAG AGC CAC ATG GTC TCT CTG GTT GTG AAT GTC CCA CCC CAG
Glu Lys Gln Ser His Met Val Ser Leu Val Val Asn Val Pro Pro Gln
390 395 400 405

ATC GGT GAG AAA GCC TTG ATC TCG CCT ATG GAT TCC TAC CAG TAT GGG
Ile Gly Glu Lys Ala Leu Ile Ser Pro Met Asp Ser Tyr Gln Tyr Gly
410 415 420

ACC ATG CAG ACA TTG ACA TGC ACA GTC TAC GCC AAC CCT CCC CTG CAC
Thr Met Gln Thr Leu Thr Cys Thr Val Tyr Ala Asn Pro Pro Leu His
425 430 435

CAC ATC CAG TGG TAC TGG CAG CTA GAA GAA GCC TGC TCC TAC AGA CCC
His Ile Gln Trp Tyr Trp Gln Leu Glu Glu Ala Cys Ser Tyr Arg Pro
440 445 450

GGC CAA ACA AGC CCG TAT GCT TGT AAA GAA TGG AGA CAC GTG GAG GAT
Gly Gln Thr Ser Pro Tyr Ala Cys Lys Glu Trp Arg His Val Glu Asp
455 460 465

TTC CAG GGG GGA AAC AAG ATC GAA GTC ACC AAA AAC CAA TAT GCC CTG
Phe Gln Gly Gly Asn Lys Ile Glu Val Thr Lys Asn Gln Tyr Ala Leu
470 475 480 485

ATT GAA GGA AAA AAC AAA ACT GTA AGT ACG CTG GTC ATC CAA GCT GCC
Ile Glu Gly Lys Asn Lys Thr Val Ser Thr Leu Val Ile Gln Ala Ala
490 495 500

AAC GTG TCA GCG TTG TAC AAA TGT GAA GCC ATC AAC AAA GCG GGA CGA
Asn Val Ser Ala Leu Tyr Lys Cys Glu Ala Ile Asn Lys Ala Gly Arg
505 510 515

Fig. 2.4

GGA GAG AGG GTC ATC TCC TTC CAT GTG ATC AGG GGT CCT GAA ATT ACT
 Gly Glu Arg Val Ile Ser Phe His Val Ile Arg Gly Pro Glu Ile Thr
 520 525 530

GTG CAA CCT GCT GCC CAG CCA ACT GAG CAG GAG AGT GTG TCC CTG TTG
 Val Gln Pro Ala Ala Gln Pro Thr Glu Gln Glu Ser Val Ser Leu Leu
 535 540 545

TGC ACT GCA GAC AGA AAT ACG TTT GAG AAC CTC ACG TGG TAC AAG CTT
 Cys Thr Ala Asp Arg Asn Thr Phe Glu Asn Leu Thr Trp Tyr Lys Leu
 550 555 560 565

GGC TCA CAG GCA ACA TCG GTC CAC ATG GGC GAA TCA CTC ACA CCA GTT
 Gly Ser Gln Ala Thr Ser Val His Met Gly Glu Ser Leu Thr Pro Val
 570 575 580

TGC AAG AAC TTG GAT GCT CTT TGG AAA CTG AAT GGC ACC ATG TTT TCT
 Cys Lys Asn Leu Asp Ala Leu Trp Lys Leu Asn Gly Thr Met Phe Ser
 585 590 595

AAC AGC ACA AAT GAC ATC TTG ATT GTG GCA TTT CAG AAT GCC TCT CTG
 Asn Ser Thr Asn Asp Ile Leu Ile Val Ala Phe Gln Asn Ala Ser Leu
 600 605 610

CAG GAC CAA GGC GAC TAT GTT TGC TCT GCT CAA GAT AAG AAG ACC AAG
 Gln Asp Gln Gly Asp Tyr Val Cys Ser Ala Gln Asp Lys Lys Thr Lys
 615 620 625

AAA AGA CAT TGC CTG GTC AAA CAG CTC ATC ATC CTA GAG CGC ATG GCA
 Lys Arg His Cys Leu Val Lys Gln Leu Ile Ile Leu Glu Arg Met Ala
 630 635 640 645

CCC ATG ATC ACC GGA AAT CTG GAG AAT CAG ACA ACA ACC ATT GGC GAG
 Pro Met Ile Thr Gly Asn Leu Glu Asn Gln Thr Thr Ile Gly Glu
 650 655 660

ACC ATT GAA GTG ACT TGC CCA GCA TCT GGA AAT CCT ACC CCA CAC ATT
 Thr Ile Glu Val Thr Cys Pro Ala Ser Gly Asn Pro Thr Pro His Ile
 665 670 675

ACA TGG TTC AAA GAC AAC GAG ACC CTG GTA GAA GAT TCA GGC ATT GTA
 Thr Trp Phe Lys Asp Asn Glu Thr Leu Val Glu Asp Ser Gly Ile Val
 680 685 690

CTG AGA GAT GGG AAC CGG AAC CTG ACT ATC CGC AGG GTG AGG AAG GAG
 Leu Arg Asp Gly Asn Arg Asn Leu Thr Ile Arg Arg Val Arg Lys Glu
 695 700 705

Fig. 2.5

GAT GGA GGC CTC TAC ACC TGC CAG GCC TGC AAT GTC CTT GGC TGT GCA
 Asp Gly Gly Leu Tyr Thr Cys Gln Ala Cys Asn Val Leu Gly Cys Ala
 710 715 720 725

 AGA GCG GAG ACG CTC TTC ATA ATA GAA GGT GCC CAG GAA AAG ACC AAC
 Arg Ala Glu Thr Leu Phe Ile Ile Glu Gly Ala Gln Glu Lys Thr Asn
 730 735 740

 TTG GAA GTC ATT ATC CTC GTC GGC ACT GCA GTG ATT GCC ATG TTC TTC
 Leu Glu Val Ile Ile Leu Val Gly Thr Ala Val Ile Ala Met Phe Phe
 745 750 755

 TGG CTC CTT CTT GTC ATT CTC GTA CGG ACC GTT AAG CGG GCC AAT GAA
 Trp Leu Leu Leu Val Ile Leu Val Arg Thr Val Lys Arg Ala Asn Glu
 760 765 770

 GGG GAA CTG AAG ACA GGC TAC TTG TCT ATT GTC ATG GAT CCA GAT GAA
 Gly Glu Leu Lys Thr Gly Tyr Leu Ser Ile Val Met Asp Pro Asp Glu
 775 780 785

 TTG CCC TTG GAT GAG CGC TGT GAA CGC TTG CCT TAT GAT GCC AGC AAG
 Leu Pro Leu Asp Glu Arg Cys Glu Arg Leu Pro Tyr Asp Ala Ser Lys
 790 795 800 805

 TGG GAA TTC CCC AGG GAC CGG CTG AAA CTA GGA AAA CCT CTT GGC CGC
 Trp Glu Phe Pro Arg Asp Arg Leu Lys Leu Gly Lys Pro Leu Gly Arg
 810 815 820

 GGT GCC TTC GGC CAA GTG ATT GAG GCA GAC GCT TTT GGA ATT GAC AAG
 Gly Ala Phe Gly Gln Val Ile Glu Ala Asp Ala Phe Gly Ile Asp Lys
 825 830 835

 ACA GCG ACT TGC AAA ACA GTA GCC GTC AAG ATG TTG AAA GAA GGA GCA
 Thr Ala Thr Cys Lys Thr Val Ala Val Lys Met Leu Lys Glu Gly Ala
 840 845 850

 ACA CAC AGC GAG CAT CGA GCC CTC ATG TCT GAA CTC AAG ATC CTC ATC
 Thr His Ser Glu His Arg Ala Leu Met Ser Glu Leu Lys Ile Leu Ile
 855 860 865

 CAC ATT GGT CAC CAT CTC AAT GTG GTG AAC CTC CTA GGC GCC TGC ACC
 His Ile Gly His His Leu Asn Val Val Asn Leu Leu Gly Ala Cys Thr
 870 875 880 885

 AAG CCG GGA GGG CCT CTC ATG GTG ATT GTG GAA TTC TCG AAG TTT GGA
 Lys Pro Gly Gly Pro Leu Met Val Ile Val Glu Phe Ser Lys Phe Gly
 890 895 900

Fig. 2.6

AAC CTA TCA ACT TAC TTA CGG GGC AAG AGA AAT GAA TTT GTT CCC TAT
 Asn Leu Ser Thr Tyr Leu Arg Gly Lys Arg Asn Glu Phe Val Pro Tyr
 905 910 915

AAG AGC AAA GGG GCA CGC TTC CGC CAG GGC AAG GAC TAC GTT GGG GAG
 Lys Ser Lys Gly Ala Arg Phe Arg Gln Gly Lys Asp Tyr Val Gly Glu
 920 925 930

CTC TCC GTG GAT CTG AAA AGA CGC TTG GAC AGC ATC ACC AGC AGC CAG
 Leu Ser Val Asp Leu Lys Arg Arg Leu Asp Ser Ile Thr Ser Ser Gln
 935 940 945

AGC TCT GCC AGC TCA GGC TTT GTT GAG GAG AAA TCG CTC AGT GAT GTA
 Ser Ser Ala Ser Ser Gly Phe Val Glu Glu Lys Ser Leu Ser Asp Val
 950 955 960 965

GAG GAA GAA GAA GCT TCT GAA GAA CTG TAC AAG GAC TTC CTG ACC TTG
 Glu Glu Glu Ala Ser Glu Glu Leu Tyr Lys Asp Phe Leu Thr Leu
 970 975 980

GAG CAT CTC ATC TGT TAC AGC TTC CAA GTG GCT AAG GGC ATG GAG TTC
 Glu His Leu Ile Cys Tyr Ser Phe Gln Val Ala Lys Gly Met Glu Phe
 985 990 995

TTG GCA TCA AGG AAG TGT ATC CAC AGG GAC CTG GCA GCA CGA AAC ATT
 Leu Ala Ser Arg Lys Cys Ile His Arg Asp Leu Ala Ala Arg Asn Ile
 1000 1005 1010

CTC CTA TCG GAG AAG AAT GTG GTT AAG ATC TGT GAC TTC GGC TTG GCC
 Leu Leu Ser Glu Lys Asn Val Val Lys Ile Cys Asp Phe Gly Leu Ala
 1015 1020 1025

CGG GAC ATT TAT AAA GAC CCG GAT TAT GTC AGA AAA GGA GAT GCC CGA
 Arg Asp Ile Tyr Lys Asp Pro Asp Tyr Val Arg Lys Gly Asp Ala Arg
 1030 1035 1040 1045

CTC CCT TTG AAG TGG ATG GCC CCG GAA ACC ATT TTT GAC AGA GTA TAC
 Leu Pro Leu Lys Trp Met Ala Pro Glu Thr Ile Phe Asp Arg Val Tyr
 1050 1055 1060

ACA ATT CAG AGC GAT GTG TGG TCT TTC GGT GTG TTG CTC TGG GAA ATA
 Thr Ile Gln Ser Asp Val Trp Ser Phe Gly Val Leu Leu Trp Glu Ile
 1065 1070 1075

TTT TCC TTA GGT GCC TCC CCA TAC CCT GGG GTC AAG ATT GAT GAA GAA
 Phe Ser Leu Gly Ala Ser Pro Tyr Pro Gly Val Lys Ile Asp Glu Glu
 1080 1085 1090

Fig. 2.7

TTT TGT AGG AGA TTG AAA GAA GGA ACT AGA ATG CGG GCT CCT GAC TAC
 Phe Cys Arg Arg Leu Lys Glu Gly Thr Arg Met Arg Ala Pro Asp Tyr
 1095 1100 1105

ACT ACC CCA GAA ATG TAC CAG ACC ATG CTG GAC TGC TGG CAT GAG GAC
 Thr Thr Pro Glu Met Tyr Gln Thr Met Leu Asp Cys Trp His Glu Asp
 1110 1115 1120 1125

CCC AAC CAG AGA CCC TCG TTT TCA GAG TTG GTG GAG CAT TTG GGA AAC
 Pro Asn Gln Arg Pro Ser Phe Ser Glu Leu Val Glu His Leu Gly Asn
 1130 1135 1140

CTC CTG CAA GCA AAT GCG CAG CAG GAT GGC AAA GAC TAT ATT GTT CTT
 Leu Leu Gln Ala Asn Ala Gln Gln Asp Gly Lys Asp Tyr Ile Val Leu
 1145 1150 1155

CCA ATG TCA GAG ACA CTG AGC ATG GAA GAG GAT TCT GGA CTC TCC CTG
 Pro Met Ser Glu Thr Leu Ser Met Glu Glu Asp Ser Gly Leu Ser Leu
 1160 1165 1170

CCT ACC TCA CCT GTT TCC TGT ATG GAG GAA GAG GAA GTG TGC GAC CCC
 Pro Thr Ser Pro Val Ser Cys Met Glu Glu Glu Val Cys Asp Pro
 1175 1180 1185

AAA TTC CAT TAT GAC AAC ACA GCA GGA ATC AGT CAT TAT CTC CAG AAC
 Lys Phe His Tyr Asp Asn Thr Ala Gly Ile Ser His Tyr Leu Gln Asn
 1190 1195 1200 1205

AGT AAG CGA AAG AGC CGG CCA GTG AGT GTA AAA ACA TTT GAA GAT ATC
 Ser Lys Arg Lys Ser Arg Pro Val Ser Val Lys Thr Phe Glu Asp Ile
 1210 1215 1220

CCA TTG GAG GAA CCA GAA GTA AAA GTG ATC CCA GAT GAC AGC CAG ACA
 Pro Leu Glu Glu Pro Glu Val Lys Val Ile Pro Asp Asp Ser Gln Thr
 1225 1230 1235

GAC AGT GGG ATG GTC CTT GCA TCA GAA GAG CTG AAA ACT CTG GAA GAC
 Asp Ser Gly Met Val Leu Ala Ser Glu Glu Leu Lys Thr Leu Glu Asp
 1240 1245 1250

AGG AAC AAA TTA TCT CCA TCT TTT GGT GGA ATG ATG CCC AGT AAA AGC
 Arg Asn Lys Leu Ser Pro Ser Phe Gly Gly Met Met Pro Ser Lys Ser
 1255 1260 1265

AGG GAG TCT GTG GCC TCG GAA GGC TCC AAC CAG ACC AGT GGC TAC CAG
 Arg Glu Ser Val Ala Ser Glu Gly Ser Asn Gln Thr Ser Gly Tyr Gln
 1270 1275 1280 1285

Fig. 2.8

TCT GGG TAT CAC TCA GAT GAC ACA GAC ACC ACC GTG TAC TCC AGC GAC
 Ser Gly Tyr His Ser Asp Asp Thr Asp Thr Thr Val Tyr Ser Ser Asp
 1290 1295 1300

 GAG GCA GGA CTT TTA AAG ATG GTG GAT GCT GCA GTT CAC GCT GAC TCA
 Glu Ala Gly Leu Leu Lys Met Val Asp Ala Ala Val His Ala Asp Ser
 1305 1310 1315

 GGG ACC ACA CTG CAG CTC ACC TCC TGT TTA AAT GGA AGT GGT CCT GTC
 Gly Thr Thr Leu Gln Leu Thr Ser Cys Leu Asn Gly Ser Gly Pro Val
 1320 1325 1330

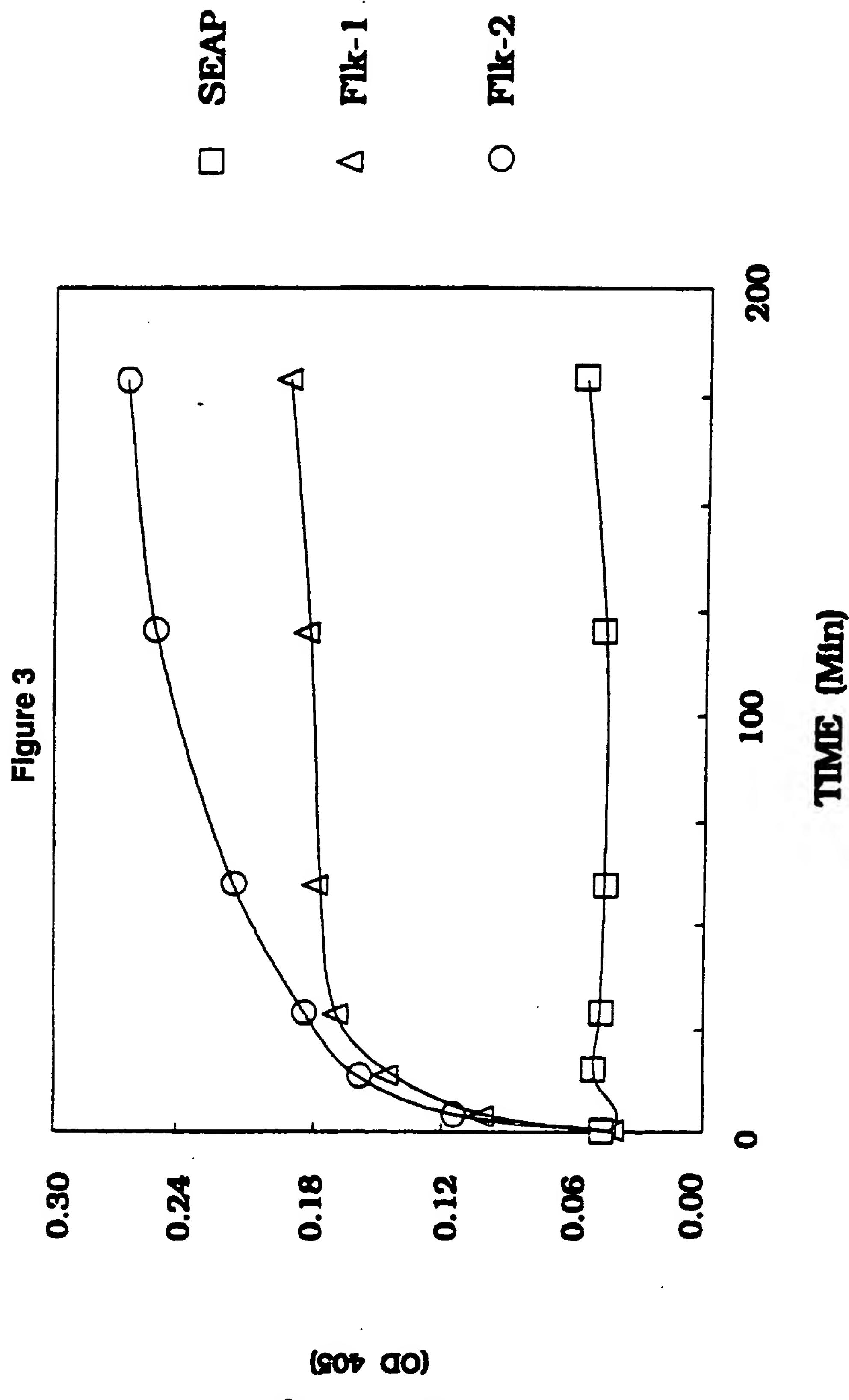
 CCG GCT CCG CCC CCA ACT CCT GGA AAT CAC GAG AGA GGT GCT GCT TAG
 Pro Ala Pro Pro Pro Thr Pro Gly Asn His Glu Arg Gly Ala Ala
 1335 1340 1345

 ATTTCAAGT GTTGTCTTT CCACCACCCG GAAGTAGCCA CATTGATT TCATTTTGG
 AGGAGGGACC TCAGACTGCA AGGAGCTTGT CCTCAGGGCA TTTCCAGAGA AGATGCCAT
 GACCCAAGAA TGTGTTGACT CTACTCTCTT TTCCATTCA TAAAGTCC TATATAATGT
 GCCCTGCTGT GGTCTCACTA CCAGTTAAAG CAAAGACTT TCAAACACGT GGACTCTGTC
 CTCCAAGAAG TGGCAACGGC ACCTCTGTGA AACTGGATCG AATGGGCAAT GCTTGTGTG
 TTGAGGATGG GTGAGATGTC CCAGGGCCGA GTCTGTCTAC CTTGGAGGCT TTGTGGAGGA
 TGCAGGCTATG AGCCAAGTGT TAAGTGTGGG ATGTGGACTG GGAGGAAGGA AGGCGCAAGC
 CGTCCGGAGA GCGGTTGGAG CCTGCAGATG CATTGTGCTG GCTCTGGTGG AGGTGGGCTT
 GTGGCCTGTC AGGAAACGCA AAGGCGGCCG GCAGGGTTTG GTTTGGAAG GTTGCCTGC
 TCTTCACAGT CGGGTTACAG GCGAGTTCCC TGTGGCGTT CCTACTCCTA ATGAGAGTTC
 CTTCCGGACT CTTACGTGTC TCCTGGCCTG GCCCCAGGAA GGAAATGATG CAGCTTGCTC
 CTTCCCTCATC TCTCAGGCTG TGCCTTAATT CAGAACACCA AAAGAGAGGA ACGTCGGCAG
 AGGCTCCTGA CGGGGCCGAA GAATTGTGAG AACAGAACAG AAACTCAGGG TTTCTGCTGG
 GTGGAGACCC ACGTGGCGCC CTGGTGGCAG GTCTGAGGGT TCTCTGTCAA GTGGCGGTAA
 AGGCTCAGGC TGGTGTCTT CCTCTATCTC CACTCCTGTC AGGCCCCCAA GTCCTCAGTA
 TTTAGCTTT GTGGCTTCCT GATGGCAGAA AAATCTTAAT TGGTTGGTTT GCTCTCCAGA

Fig. 2.9

TAATCACTAG CCAGATTCG AAATTACTTT TTAGCCGAGG TTATGATAAC ATCTACTGTA
TCCTTTAGAA TTTAACCTA TAAAACATATG TCTACTGGTT TCTGCCTGTG TGCTTATGTT
AAAAAAAAAA AAAAAA

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(cont'd)

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Figure 4

